Metal Nanostructure-Enhanced Fluorescence and its Applications in Bioassays and Bioimaging

By

Wei Deng

A THESIS SUBMITTED TO MACQUARIE UNIVERSITY FOR THE DEGREE OF DOCTOR OF PHILOSOPHY DEPARTMENT OF DEPARTMENT OF PHYSICS AND ENGINEERING APRIL 2011



EXAMINER'S COPY

© Wei Deng, 2011.

Typeset in $\mathbb{A}T_{\mathbb{E}} X 2_{\mathcal{E}}$.

Except where acknowledged in the customary manner, the material presented in this thesis is, to the best of my knowledge, original and has not been submitted in whole or part for a degree in any university.

Wei Deng

Acknowledgements

Many people have offered me valuable help in my thesis writing, my supervisors, my colleagues, my family and my friends. Firstly, I would like to give my sincere gratitude to my principal supervisor, Prof. Ewa M Goldys, with her extraordinary patience and consistent encouragement. She gave me great help by providing me with necessary materials, advice of great value and inspiration of new ideas. It is her suggestions that draw my attention to a number of deficiencies and make many things clearer. Without her strong support, this thesis could not have been the present form. Then, I am pleased to acknowledge my co-supervisors, Ms. Krystyna Drozdowicz-Tomsia and Dr. Dayong Jin, for their invaluable assistance throughout the whole PhD project. We have had plenty of fruitful discussion ranging from technical issues to theoretical issues dealing with my PhD project. Special thanks to Prof. Jingli Yuan and his group of Dalian University of Technology, China for his important collaboration work, which provided me with Eu chelates used in this thesis. Without this important collaboration over the continents, this thesis would not exist! Last my thanks would go to my beloved family for their loving considerations and great confidence in me all through these years. I also owe my sincere gratitude to my friends and my fellow colleagues who gave me their help and time in listening to me and helping me work out my problems during the difficult course of the thesis.

List of Publications

- Wei Deng, Dayong Jin, Krystyna Drozdowicz-Tomsia, Jingli Yuan, Jing Wu, Ewa M Goldys, Ultrabright Eu-doped Plasmonic Ag@SiO2 Nanostructures: Time-gated Bioprobes with Single Particle Sensitivity and Negligible Background. Adv. Mater. 23, 4649–4654 (2011).
- Wei Deng, Sudheendra Lakshmana, Junxiang Fu, Jiangbo Zhao, Ian Kennedy, Dayong Jin, Ewa M Goldys, *Upconversion in NaYF4 : Yb,Er Nanoparticles Amplified by Metal Nanostructures*. Nanotechnology, **22**, **325604 (2011)**.
- Wei Deng, Dayong Jin, Krystyna Drozdowicz-Tomsia, Jingli Yuan, Ewa M Goldys, Metal Nanostructure Enhanced Fluorescence of Europium Chelate BHHCT-Eu³⁺ Applied to Bioassays and Time-gated Bioimaging. Langmuir, 26, 10036-10043(2010).
- Wei Deng, Krystyna Drozdowicz-Tomsia, Dayong Jin, Ewa M Goldys, *Enhanced Flow Cytometry Based Bead Immunoassays by Using Metal Nanostructures*. Anal. Chem., **81**, 7248-7255 (2009).
- Wei Deng, Krystyna Drozdowicz-Tomsia, Dayong Jin, Ewa M Goldys, *Silver Nanostructure Coated Beads Enhance Fluorescence for Sensitive Immunoassays and Bio-imaging*. IEEE Explorer (accepted).
- Wei Deng, Dayong Jin, Ewa M. Goldys, "Metal Nanostructure-enhanced Fluorescence and its Biological Applications" in the book "Nanotechnology in Australia: Showcase of Early Career Research" edited by D M Kane, A Micolich and J Rabeau (ISBN: 9789814310024) by Pan Stanford Publishing Pte. Ltd
- Varun K. A. Sreenivasan, Ekaterina A. Ivukina, Wei Deng, Timothy A. Kelf, Tatyana A. Zdobnova, Sergey V. Lukash, Boris V. Veryugin, Oleg A. Stremovskiy, Andrei V. Zvyagin, Sergey M. Deyev, *Barstar: Barnase-a Versatile Platform for Colloidal Diamond Bioconjugation*. J. Mater. Chem., 21, 65-68 (2011).
- Yiqing Lu, Dayong Jin, Robert C. Leif, Wei Deng, James A Piper, Jingli Yuan,

Yusheng Duan, Yujing Huo, Automated Detection of Rare-Event Pathogens through Time-gated Luminescence Scanning Microscopy. Cytometry Part A, **79A**, 349-355 (2011).

- Ewa M. Goldys, Wei Deng, Nils P. Calander, Krystyna Drozdowicz-Tomsia, Dayong Jin, *Nanoscale Plasmonic Resonators with High Purcell Factor: Spontaneous and Stimulated Emission.* SPIE Proceedings (accepted).
- Wei Deng, Dayong Jin, Krystyna Drozdowicz-Tomsia, Jingli Yuan, Jing Wu, Ewa M Goldys, *Plasmonic Ag/SiO*₂ *Composite Nanoparticles Doped with Europium Chelate and Their Metal Enhanced Fluorescence*. Proceedings of SPIE, **7909**, 79090Z (2011).

Abstract

Over the last two decades, fluorescence based detection has become one of the leading sensing technologies in biomedical, biological and related sciences. Its sensitivity makes it possible to detect a single biomolecule through labeling with a suitable fluorophore. Two principal fluorophore properties, brightness and photostability, are fundamentally important to achieve a high level of sensitivity and in many conventional fluorophores these often fall short of the requirements. Among the methods used to improve the sensitivity of fluorescence detection, the metal-enhanced fluorescence (MEF) technique has been recently actively developed. The MEF phenomenon occurs when an excited fluorophore is located in close proximity to metals, and it is particularly pronounced near noble metal nanostructures. Electrons in such metal nanostructures exhibit strong resonances often located in the visible part of the spectrum (also known as surface plasmon resonance). They can interact with proximal fluorophores modifying their optical properties and producing increased quantum yield (fluorescence efficiency) and improved photostability. It has been experimentally demonstrated that the MEF technique can increase fluorescence intensity up to several hundreds times. The work through my PhD project mainly focus on siver nanostructures and its potential in fluorescence-based applications requiring very high sensitivity because their strong surface plasmon resonance in the visible matches the absorption and emission bands of most fluorophores.

I began with synthesis for Ag nanostructure-coated silica beads in the solution-based

platform as a new MEF substrate. My first study employed these nanostructures to enhance the fluorescence readout on individual silica beads. These Ag nanostructure were deposited on micrometer size silica beads. The fluorescence enhancement was investigated using a model AlexaFluor 430 IgG immunoassay and AlexaFluor 430 labeling. Approximately 8.5-fold and 10.1-fold higher fluorescence intensities at 430 nm excitation were, respectively, observed from silvered 400 nm and 5 μ m silica beads deposited on glass as compared to the control sample. This achievement allowed us to demonstrate for the first time MEF immunoassays on silica beads by using high-throughput flow cytometry. Furthermore, we discovered that these Ag nanostructure-coated silica beads are able to modify the luminescence decay lifetime of lanthanide fluorophores $(BHHCT - Eu^{3+})$ for time-gated luminescence bioimaging applications. The fluorescence enhancement factor achieved about 11 times, while the simultaneously measured fluorescence lifetime was reduced twofold. The fluorophore stability was also improved by a factor of three. We applied such bead substrates to time-gated fluorescence imaging of Giardia lamblia cells stained by $BHHCT - Eu^{3+}$ with improvement in brightness by a factor of two. This will open up a broad range of opportunities for ultrasensitive and low background fluorescence detection using lanthanide fluorophores. Additionally, I applied these Ag nanostructures to another type of luminophor, upconversion (UC) nanoparticles via a simple bioassy. Specifically, streptavdin (SA)-labeled UC nanoparticles were bound to the biotinlyated anti-mouse IgG antibody which was attached to Ag nanostructure-coated silica beads in advance. These Ag nanostructures produced the strong luminescence enhancement (~ 4.4 -fold for the green mission and ~ 3.5 -fold for the red emission) from $NaYF_4$: Yb, Er nanoparticles, while the simultaneously measured luminescence lifetime was reduced 2-fold for the green mission and 2.2-fold for the red emission. These findings open a new pathway to rationally modulate the UC emission, and broadly impact areas such as bioimageing and bioassays, as well as enable new opportunities for energy harvesting and conversion.

In addition, I developed another kind of MEF substrate in this project, Eu chelatedoped Ag@SiO₂ nanocomposites, and systematically study the interaction between Ag core and Eu chelate (BHHCT-Eu-DPBT) by tuning the distance between them and Ag core sizes. At high excitation intensities, these nanocomposites showed greatly increased fluorescence enhancement factors of up to 145, due to significantly increased radiative rates in samples with metal cores, from about 700 s^{-1} in control samples to over 5000 s^{-1} . They are bright enough to be observed as single particles and are compatible with low background time-gating by using simple detection systems. A simple bioassay using avidin-biotin binding system was also carried out with the luminescence enhancement factor of ~120, demonstrating the potential of these bioconjugated nanocomposites to be used in a range of biological applications.

Based on these experimental results, we also presented the theoretical analysis for the high fluorescence enhancement factor. This phenomenon observed on the nanostructured silver surfaces is a result of two effects: an increase of a local electromagnetic field near silver nanostructures, leading to increased excitation rate of fluorophores and an increase of the radiative decay rate Γ of fluorophores close to silver nanostructures, reflected both in the fluorescence lifetime and quantum yield. The local electromagnetic field enhancement produces a higher excitation rate but it does not change the lifetime of the fluorophore; this effect is referred as excitation enhancement (E_{ex}). The second effect referred as emission enhancement (E_{em}), increases the quantum yield and reduces the lifetime of the fluorophore. The detailed mechanism was systematically analysed in the following chapters.

In summary, this work mainly covered the fabrication of nanoscaled silver with different geometries and the MEF effect induced by them in biological applications such as bioassays, immunoassays and bioimaging.

Contents

1	Acknowledgements	v
2	List of Publications	vii
3	Abstract	ix
Lis	st of Figures	xv
Lis	st of Tables	xv
4	Introduction	1
5	Fluorescence and Fluorescence Instrumentation (review)	7
6	Metal-enhanced Fluorescence and its Biological Application (review)	33
7	Enhanced Flow Cytometry-Based Bead Immunoassays Using Metal	
	Nanostructures	55
8	Metal Nanostructure Enhanced Fluorescence of Europium Chelate	
	$BHHCT - Eu^{3+}$ applied to Bioassays and Time-gated Bioimaging	75
9	Upconversion Phosphor Nanoparticles and its Application in Bioas-	
	says (review)	95

- 11 Ultrabright Eu-doped Plasmonic $Ag@SiO_2$ Nanostructures: Time-gated Bioprobes with Single Particle Sensitivity and Negligible Background 129

12 Conclusion	147
References	153

List of Figures

5.1	Example of Jablonski diagram	8
5.2	A simplified Jablonski diagram to illustrate the meaning of quantum	
	yields and lifetimes	10
5.3	Jablonski diagram with collisional quenching and fluorescence resonance	
	energy transfer (FRET). The term $\sum k_i$ is used to represent non-radiative	
	paths to the ground state aside from quenching and FRET	12
5.4	Schematic diagram of a spectrofluorometer	14
5.5	Corrected and uncorrected excitation spectra of fluorescein	16
5.6	Monochromators based on a plane (top) or concave (bottom) grating. $\ .$	17
5.7	Transmission spectra of some typical colored glass filters	18
5.8	Long-pass filters designed to reject light from a helium-cadmium laser	
	at 325 nm or an argon ion laser at 488 nm	19
5.9	Schematic diagram of a photomultiplier tube and its dynode chain. $\ .$.	19
5.10	Schematic diagram of a flow cytometer	21
5.11	Hydrodynamic focusing of the sample core through a flow cell	22
5.12	Light-scattering properties of a cell	23
5.13	Schematic of a typical epifluorescence microscopy.	25
5.14	The schematic confocal microscopy.	26
5.15	Basic principle of the SPC-830 TCSPC Imaging module	30
5.16	System configuration of the Leica FLIM systems.	32

6.1	Schematic of surface plasmons on a metal surface. Adapted from $\left[40\right]$.	36
6.2	Surface plasmon oscillation in a metallic colloid. Adapted from $\left[39\right] $.	36
6.3	Classical Jablonski diagram: E_m is metal enhanced excitation rate, Γ_m is radiative rate in the presence of metal, k_{nr} is nonradiative decay rate of fluorophores. The weak effect of metals on k_{nr} is ignored	37
6.4	Effects of metals on the steady-state intensity, intensity decay, and pho- tobleaching of a nearby fluorophore. From top to bottom the panels show quenching by the metal, the effect of an increased excitation field, and the effects of an increased radiative decay rate. The dashed lines indicate the absence of metal and the solid line indicates the presence of metal, adopted from [34].	39
6.5	(Left) silver nanostructures with a size of 47 nm on the glass slide. In- serts show the morphology of individual particles. (Right) Fluorescence emission intensity of FITC-HSA (from top to bottom): on the surface of silver nanostructures with a size of 47 nm; on the surface of silver nanostructures with a size of 19 nm; on the clean glass surface without silver nanostructures; on the surface of gold colloid monolayer, adopted from [56].	43
6.6	Scheme of the model immunoassay, adopted from [59]	44
6.7	Scheme of the model immunoassay (A) and myoglobin (sandwich-format) immunoassay (B) on the SiF-modified slide surface. Ab, antibody, adopted from [60].	45
6.8	Fluorescence spectra of the labeled (AlexaFluor 55) anti-rabbit antibod- ies bound to the antigen immobilized on various slide supports used for immunoassay: glass only, SiFs on aluminum mirror, SiFs on bare glass, SiFs on gold mirror, and SiFs on silver mirror, adopted from [60]	46

6.9	(a) Dependence of the enhancement factor on the thickness of silver
	film coated by a 5 nm thickness of silica upon excitation at 514 nm;
	(b)Dependence of the enhancement factor on the thickness of silica
	coated on a 10 nm thickness of silver film; (c)Dependence of the en-
	hancement factor on the emission wavelength of a fluorophore-labeled
	antibody on a 10 nm thickness of a silver film coated by a 5 nm thick-
	ness of silica, adopted from [61]

- 6.10 Scheme showing DNA hybridization in solution versus on a surface. (A)Solution hybridization; (B) Surface hybridization, adopted from [70]... 48
- 6.11 Fluorescence image of labeled oligonucleotide targets hybridized to MEF and glass DNA arrays. Probe oligonucleotides (23 mer) were arrayed onto the substrates at different spotting concentrations: each row represents seven replicate spots at a given concentration; the rows are 2-fold serial dilutions starting with 50 mM (upper row). The image shows the Cy5 fluorescence as a result of co-hybridization with complementary Cy5- and Cy3-labeled targets (23 mer), adopted from [70]. 49
- 6.12 Upper panel: Representative emission intensity images of PM1 cells labeled with Alexa Fluor 680-dextran conjugates adhered to the cell membranes on a glass coverslip and on SiFs. Lower panel: Corresponding lifetime images of the intensity images in the upper panel, adopted from [77].
 50

6.14 Schematic for the preparation of fluorescent core-shell Ag@SiO₂ (MEF)
nanoballs and fluorescent nanobubbles, adopted from [79].
52

47

6.16	Preparation, displacement by thiolate oligonucleotides, and hybridiza-	
	tion with fluorescein-labeled complementary oligonucleotides of tiopronin- monolayer protected silver nanoparticle, adopted from [80]	53
7.1	TEM images of Au-coated silica beads produced using different amounts of gold colloid solution: (a) 0.5 mL, (b) 1 mL, (c) 2 mL	61
7.2	Absorption spectra of the pure silica beads and samples at different Ag enhancing times, as indicated in the graph	61
7.3	Absorption spectra of the Ag layer on the surface of silica beads \ldots	62
7.4	SEM images of silica beads with silver deposition for (a) 10sec, (b)1 min, (c)3 min and (d)5 min	63
7.5	Fluorescence intensities observed from the supernatant (a)(from top to bottom: original AlexaFluor 430 solution, without Ag enhancement, 10sec, 1 min and 3 min Ag enhancement)and the glass surfaces(b) (from top to bottom: 3 min, 1 min, 10 sec and without Ag enhancement) for different samples with silica beads of 400nm	65
7.6	Fluorescence intensities from the supernatant (a) (from top to bottom: original AlexaFluor 430 solution, glass surface without Ag enhancement and glass surface with 3 min Ag enhancement) and from the glass sur- faces (b) (from top to bottom: 5 μ m silica beads with 3 min Ag en- hancement and without Ag enhancement.)	66
7.7	Laser scanning microscopy images and corresponding histograms of sam- ples with 3 min Ag enhancement (a) and without Ag enhancement (b). The horizontal axis of the histogram shows brightness in arbitrary units.	67
7.8	Representative fluorescence lifetime decay curves of AlexaFluor 430- labelled immunoassay measured on samples with different Ag enhancing	
	times	68

7.9	Flow cytometry results for 5 μ m silica beads: AlexaFluor 430-antibody on the silica beads without Ag enhancing (a), with 10 sec Ag enhanc-	
	ing (b), with 1 min Ag enhancing (c) and with 3 min Ag enhancing(d). Left, forward scattering (FSC) versus side scattering (SSC); Right,	
	fluorescence histograms.	72
7.10	Flow cytometry scanning for AlexaFluor 430 labelled -silica beads (400nm) without Ag enhancement (a) and with 3 minutes Ag enhancement (b). Left, forward scattering vs. side scattering; Right, fluorescence histogram.	73
7.11	Fluorescence intensities from the glass surfaces for 400 nm (a) and 5 $\mu {\rm m}$	
	(b) silica beads under the 488 nm excitation. (from top to bottom: with3 min Ag enhancement and without Ag enhancement)	73
8.1	Structure of BHHCT – Eu^{3+} chelate	77
8.2	Schematic representation of conjugation of SA-BSA with BHHCT $-Eu^{3+}$ chelate (a) and the bioassay using BHHCT $-Eu^{3+}$ as a label (b)	79
8.3	Schematic representation of immunostaining Giardia lamblia cells with $BHHCT - Eu^{3+}$.	81
8.4	Absorption spectra of pure BHHCT, SA-BSA conjugates and SA-BSA conjugates with BHHCT (from top to bottom).	83
8.5	SEM images of silica beads at different Ag enhancing times (a) 10sec, (b) 1 min, (c) 3 min and (d) 5 min	84
8.6	Fluorescence intensities measured from the glass slides (a) and silica beads (b) with different Ag enhancing time and without Ag (from top to bottom: 3 min, 1 min, 10 sec and without Ag enhancement)	86
8.7	Fluorescence intensities measured in supernatants (sample a: original BHHCT – Eu^{3+} -labeled SA-BSA solution; sample b: without Ag enhancement; sample c: 10 sec Ag enhancement; sample d: 1 min Ag	
	enhancement; sample e: 3 min Ag enhancement)	86

8.8	Fluorescence spectrum from BHHCT – Eu^{3+} on the silica beads with 3 min Ag enhancement after non-specific binding under the same experi-	
	mental conditions.	87
8.9	Representative fluorescence lifetime decay curves of BHHCT – Eu^{3+} - labelled assay measured on the glass slides silica beads (a) and silica beads (b) with and without Ag enhancement steps	88
8.10	Photostability comparison of BHHCT $- Eu^{3+}$ -labelled assay with and without Ag enhancing step with power adjusted to provide the same initial fluorescence intensity	92
8.11	Representative images of Giardia lamblia stained by BHHCT $- Eu^{3+}$ without Ag enhancement as reference (a) and with 3 min Ag enhance- ment (b). All the images were collected under the same experimental conditions	93
9.1	Schematic representation of the most prominent upconversion mecha- nisms: ground-state absorption (GSA), excited-state absorption (ESA) and energy transfer up-conversion (ETU), respectively. The dotted ar- rows represent nonradiative energy transfer (ET) processes. The straight arrows indicate radiative transitions. The transients indicated in (b) and (d) describe the time-evolution of the emission of the upconversion lu- minescence after a short excitation pulse, adopted from [161]	99
9.2	Principal of the two-photon up-conversion process (a) and three-photon up-conversion (b), adopted from [163]	100
9.3	Detection of PSA in paraffin-embedded sections of human prostate tissue with biotinylated antibodies and neutravidin blue phosphors. a) Tissue section after exposure to both blue and IR excitation light. The green autofluorescence coincides with the PSA specific blue phosphor lumines- cence; b) IR excitation of the blue phosphor labelled PSA, adopted from [146]	106
		100

- 9.4 UCP lateral flow format. The architecture of the UCP lateral flow strip is designed to accommodate up to 12 distinct test lines. In addition, each strip also contains two control lines, adopted from [148] 107

- 9.8 In vivo imaging of rat: quantum dots (QDs) injected into translucent skin of foot (a) show fluorescence, but not through thicker skin of back (b) or abdomen (c); PEI/NaYF₄ nanoparticles injected below abdominal skin (d), thigh muscles (e), or below skin of back (f) show luminescence, adopted from [195] 112
- 9.9 False color two-photon images of C. elegans at 980 nm excitation with red representing the bright field and green for the phosphor emission. The worms were deprived of food over a period of 24 h, showing little or no change at (a) 0 h, (b) 4 h, and (c) 24 h, adopted from [220] . . . 113

9.10	UCP Instrumentation. a) Schematic representation of a Leica DM epi- fluorescence microscope modified to excite UC particles with 980nm light	
	from a xenon XBO 75W lamp and to visualise blue, green or red phos- phor luminescence. b) Schematic representation of a modified Packard	
	FluoroCount reader, adopted from [143, 222]	113
10.1	Schematic representation of conjugation SiO_2 -coated nanoparticles with streptavidin (SA)(a) and a SA-biotin-based assay (b)	120
10.2	TEM images of pure (a) and SiO_2 -coated $NaYF_4$: Yb, Er (b). (c) The UC luminescence spectra of SiO_2 -coated and uncoated $NaYF_4$: Yb, Er nanoparticles.	122
10.3	The TEM image of gold shell-coated $NaYF_4$: Yb, Er nanocrystals	123
10.4	Schematic illustration of the UC process of Er^{3+} in $NaYF_4$: Yb, Er under 980 nm excitation. CR: cross relaxation	124
10.5	TEM image of Ag nanostructure-coated silica beads (a) and absorption spectra of the pure and Ag nanostructure-coated silica beads (b). \ldots	125
10.6	The UC luminescence spectra observed from samples with and without Ag nanostructures (a) and with and without gold nanoshells (b)	125
10.7	Representative UC luminescence decay times of the green(a) and the red(b) emissions from $NaYF_4$: Yb, Er nanoparticles with and without Ag nanostructures.	126
10.8	Representative UC luminescence decay times of the green(a) and the $red(b)$ emissions from $NaYF_4$: Yb, Er nanoparticles with and without Au shells.	127
11.1	Schematic structure of europium chelate, BHHCT-Eu-DPBT (a) and steps required in the formation of BHHCT-Eu-DPBT-doped $Ag@SiO_2$	
	nanocomposites (b).	131

11.2 TEM images of $Ag@SiO_2$ nanocomposites with different thickness of	
silica shells (upper panel, from left to right: ${\sim}12$ nm, ${\sim}25$ nm and ${\sim}57$	
nm) and Ag-core sizes (lower panel, from left to right: ${\sim}33$ nm, ${\sim}52$ nm	
and $\sim 81 \text{ nm}$)	;
11.3 UV-visible absorption spectra of $Ag@SiO_2$ nanocomposites with differ-	
ent silica shell thickness (a) , Ag core sizes (b) and hollow silica nanoshells	
(c). Insets are TEM images of the corresponding $Ag@SiO_2$ nanocom-	
posites and SiO_2 nanoshells	3
11.4 Luminescence intensities observed from BHHCT-Eu-DPBT-doped Ag@SiO_2 $$	
nanocomposites with ${\sim}25~\mathrm{nm}$ silica shell and ${\sim}52~\mathrm{nm}$ Ag-core size (solid	
line) and nanoshells without Ag core (dashed line))
11.5 Representative fluorescence lifetime decay curves of BHHCT-Eu-DPBT	
in $Ag@SiO_2$ nanocomposites with different silica shell thickness(a) and	
with different Ag-core size(b)	L
11.6 Time-gated luminescence images of single nanocomposites. a) BHHCT-	
Eu-DPBT-doped $Ag@SiO_2$ with 25 nm silica shell and 52 nm Ag core.	
b) BHHCT-Eu-DPBT-doped hollow nanocomposites with 25 nm silica	
shell and without 52 nm Ag core. c) Original red spots from BHHCT-	
Eu-DPBT-doped Ag@SiO ₂ single nanocomposites. d) The enlarged im-	
age of spots in the square. e) Scanning electron microscope image of	
BHHCT-Eu-DPBT-doped $\mathrm{Ag}@\mathrm{SiO}_2$ nanocomposites. Data taken with	
$\times 60$ magnification in the time-gated fluorescence microscope and 2 s	
exposure time. \ldots \ldots \ldots \ldots \ldots \ldots \ldots \ldots \ldots 14^4	F
11.7 Schematic representation of conjugation of BHHCT-Eu-DPBT-doped	
$Ag@SiO_2$ nanocomposites with biotinylated antibody(a) and bioassay	
using $Ag@SiO_2$ nanocomposites as labels	ý
11.8 Luminescence spectra of nanocomposite-labeled antibody after bioas-	
say. Solid line - BHHCT-Eu-DPBT-doped ${\rm Ag}@{\rm SiO}_2$ with 12 nm silica	
shell and 52 nm Ag core, dotted line - BHHCT-Eu-DPBT-doped hollow	
nanocomposites with 12 nm silica shell and without 52 nm Ag core 146 $$;

List of Tables

7.1	Binding efficiencies and corrected enhancement factors for 400 nm and	
	5 $\mu \mathrm{m}$ silica beads at different silver enhancing times. The data marked	
	with a * were taken on 5 $\mu \mathrm{m}$ silica beads. Measurements taken at 430	
	nm excitation.	65
7.2	Fluorescence intensity decay analysis. $\tau_{1,2}$ denotes decay times of the	
	two lifetime components, τ is the average lifetime	67
7.3	Calculated enhancement factor for samples with different Ag enhancing	
	times by using flow cytometry	69
8.1	Fluorescence intensity decay analysis. $\tau_{1,2}$ denotes decay times of the	
	two lifetime components, τ is the average lifetime	88
8.2	Lifetime measurements for each sample and the calculated values of rate	
	(Γ/Γ_m) , yield ratios (Q_m/Q) , and modified quantum yield (Q_m) ; τ is the	
	average lifetime value of the control sample (380 ms for the silica beads	
	and 382 ms for the glass slides); τ_m is the modified lifetime; Q is the	
	average quantum yield of the control sample (0.27). \ldots \ldots \ldots	90
8.3	Values of the excitation enhancement (E_{ex}) and emission enhancement	
	(E_{em}) for each sample. E_t is the calculated enhancement factor for each	
	sample.	91
9.1	Composition of Upconversion Phosphors	114

11.1	Fluorescence intensity enhancement analysis. $t_{\rm SiO2}$ denotes thickness of	
	the silica shell, λ denotes the wavelength of the emission peak from Eu	
	chelate	140
11.2	Fluorescence intensity enhancement analysis. $\mathrm{D}_{\mathrm{Ag-core}}$ denotes denotes	
	the diameter of Ag core, λ denotes the wavelength of the emission peak	
	from Eu chelate	140
11.3	Fluorescence intensity decay analysis. $\mathrm{t}_{\mathrm{SiO2}}$ denotes thickness of the	
	silica shell, $\tau_{1,2}$ denotes decay times of the two lifetime components, τ is	
	the average lifetime.	141
11.4	Fluorescence intensity decay analysis. $\mathrm{D}_{\mathrm{Ag-core}}$ denotes the diameter of	
	Ag core, $\tau_{1,2}$ denotes decay times of the two lifetime components, τ is	
	the average lifetime.	142