# Proteomics of Wheat Bran

## (*Triticum aestivum* var. Babbler)

A thesis submitted in fulfilment of the requirements

for the Honours Degree of Master of Science

By

**Ante Jerkovic** 

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## Declaration

The work presented in this thesis was carried out between March 2005 and October 2006 on a full-time basis. This work represents original research which has not been submitted for any other degree. All work was carried out by the author unless otherwise acknowledged.

Candidates Signature

Ante Jerkovic

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### Abstract

Wheat is a major crop in Australia with around 25 million tonnes of grain harvested in an average year. Improved wheat grain cultivars and wheat grain milling can result in higher biological yields and flour quality. The introduction covers the general aspects of the wheat grain from bran development and structure through to milling and the importance of flour quality in flour-based products. It also highlights the problem with bran contamination in flour during milling and other factors that may have an effect on flour quality. Proteomics was used to identify proteins in three separate bran tissue fractions: the inner fraction (aleurone), intermediate fraction (nucellar tissue, testa, tube cells and cross cells) and the outer fraction (hypodermis and epidermis). The aim of the project was to identify proteins in bran tissue fractions which may potentially be useful in improvements in wheat quality for farmers and consumers and flour yield for millers. The results show that more than 80% of the identified proteins in the outer and intermediate tissue fractions are defence-and stress-related proteins (chitinase, xylanase, thaumatinlike protein, wheatwin 1, lipid-transfer protein, oxalatae oxidase (OXO), polyphenol oxidase (PPO), peroxidase (POX). Almost 60% of the proteins identified in the inner tissue fraction are 7S Globulin storage proteins and around 15% are protein synthesis-and energy-related. Water-soluble proteins were also identified and it was found that endochitinase, OXO, PPO and POX all leach out from the grain during imbibition. This study has added to the knowledge of bran tissue-specific proteins and has broad implications for improving crop yield and flour quality.

## Abbreviations

1-DE	One-dimensional gel electrophoresis
<b>2-DE</b>	Two-dimensional gel electrophoresis
ABA	Abscisic acid
ABI	Applied Biosystems International
ACTH	Adrenocorticotropic hormone
APS	Ammonium persulphate
AR	Acquired resistance
ATP	Adenosine triphosphate
AX	Arabinoxylan
BSA	Bovine serum albumin
BLAST	Basic Local Alignment Tool (from NCBI)
ВТН	Benzothiadiazole
CHAPS	3-[]-1-propanesulfonate
DPA	Days post anthesis
DTT	1,4-Dithio-DL-threitol
ESI	Electrospray ionization
EST	Expressed sequence tag
GA	Gibberellic acid
IAA	Indole-3-acetic acid
IDA	Information dependent acquisition
IEF	Iso-electric focussing
IPG	Immobilised pH gradient
kDa	kilo Da
LC	Liquid chromatography
LOX	Lipoxygenase
LTP	Lipid transfer protein
MALDI-TOF MS	Matrix-assisted laser desorption/ionization time-of-flight mass
	spectrometry
MS	Mass spectrometry
MS/MS	Tandem mass spectrometry

MudPIT	Multi dimensional protein identification technology					
NCBI	National Center for Biotechnology Information					
OXO	Oxalate oxidase					
PAGE	Polyacrylamide gel electrophoresis					
PCD	Programmed cell death					
РСТ	Patent Cooperation Treaty					
PDI	Protein disulfide isomerase					
PHYLIP	Phylogeny Inference Package					
PMF	Peptide mass fingerprinting					
РОХ	Peroxidase					
PPO	Polyphenol oxidase					
PR	Pathogenesis related					
SDS	Sodium dodecyl sulfate					
TBP	Tributyl phosphine					
ТСА	Trichloroacetic acid					
TEMED	N,N,N',N'-Tetramethylethylenediamine					
TFA	Trifluoroacetic acid					
TL	Thaumatin like					
TOF	Time of flight					
VDAC	Voltage dependant anion channel					
A. fatua	Avena fatua					
A. oryzae	Aspergillus oryzae					
H. vulgare	Hordeum vulgare					
M. viride	Mesostigma viride					
O. sativa	Oryza sativa					
P. glaucum	Pennisetum glaucum					
P. miliaceum	Panicum miliaceum					
S. cereale	Secale cereale					
T. aestivum	Triticum aestivum					
X. oryzae	Xanthomonas oryzae					
Z. mays	Zea mays					

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5. Conclusion and future directions

 Electronic copy of thesis "Proteomics of Wheat Bran (*Triticum aestivum* var. Babbler)"

- Protein ID Tables (Excel format) with links to mass spectra
- Mass spectra link file

## **1. Introduction**

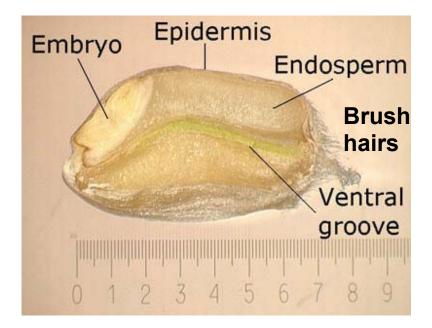
#### 1.1 Wheat in Australia

Wheat is a major food crop in Australia. Wheat growers produced around 25 million tonnes of grain in 2004, worth around five billion dollars for that year (Bread Research Institute (BRI) Australia, www.bri.com.au; Commonwealth Scientific and Industrial Research Organisation (CSIRO), www.csiro.au; Grain Foods Cooperative Research Centre (GFCRC), www.grainfoodscrc.com.au). The major economic and mass component of wheat grain is the starchy endosperm which is extracted in the milling process to produce flour and used in turn to make breads, pastas, cakes, biscuits and other baking products. Other parts of the mill stream, such as bran, germ and pollard are used in making products such as cereals, biscuits and animal feed (Every et al., 2002). To reduce the costs of maintenance and to increase profitability of crops, farmers choose to grow wheat based on yield and a number of factors such as resistance to disease, climate stress and watering requirements, whereas millers choose wheat based on flour quality and milling properties (Dupont and Altenbach, 2003). Thus there is great importance in understanding the physical and biochemical characteristics of wheat grain which make it suitable for all of these purposes. This information can potentially be used to provide improvements in yield and management of crops for farmers and increased flour yield and quality for millers.

#### 1.1.1 Wheat grain structure

Wheat grain is generally ready for harvesting at around 30 days post anthesis (DPA) depending on the cultivar. The basic structural composition of the grain at this stage is

shown in Figure 1.1. The three main components of whole grain are bran tissue, germ (which contains the embryo and scutellum), and endosperm. The distinct external feature of the grain is its 'blimp'-like shape. A cleft/crease runs longitudinally along the grain and indents approximately half way inside the grain. The embryo is indented at one end of the grain and the brush hairs are located at the opposite end.



**Figure 1.1** Longitudinal section showing wheat grain structure (modified from Home-Grown Cereals Authority (HGCA), <u>www.wheatbp.net</u>).

When grain is being milled, the ideal outcome is to separate the bran and embryo from the endosperm. The quality of flour in terms of bran contamination will largely depend on how well the bran separates from the endosperm and how much the bran fractures into smaller fragments. Conditioning grain (addition of small quantities of water) prior to milling has the effect of increasing bran strength and improving bran separation from the endosperm, resulting in a reduction of bran contamination in the flour (Butcher and Stenvert, 1973; Moss et al., 1980).

#### 1.1.2 Milling process

After wheat is harvested, it is cleaned from rocks, dirt and unwanted vegetation matter. The grains are then stored in silos where they are kept dry until later distribution. When the grain reaches the flour mill, it is firstly conditioned by addition of water for around 16 h to increase moisture content of the grain from around 10% to around 15%, depending on the wheat cultivar. As mentioned previously, the accepted wisdom is that the conditioning process toughens the bran by making it more elastic, giving it more integrity when passing through the break rollers. The result is better endosperm separation from bran during milling and reduction in bran contamination in the flour (Butcher and Stenvert, 1973; Moss et al., 1980; Every et al., 2002). After conditioning, the grains are passed through the first break rollers. The first break mostly removes semolina (flour and germ with minimal bran contamination). At this stage the cleanest fractions are collected, however the yield is quite low. Semolina and bran are then separated by fluting. The heavier bran/endosperm pieces are recycled back through to the break rollers and are passed through twice more to remove as much endosperm from the bran as possible. The remaining bran/endosperm pieces are then passed through reduction rollers, which have a smooth surface unlike the break rollers, and their purpose is to crush the starch granules into very fine flour (Moss et al., 1980; Every et al., 2002; Fang and Campbell, 2002). The

main products collected after milling are bran, semolina, flour and pollard (Every et al., 2002).

'Ash content' in flour is a name used to indicate bran contamination. When there is a high ash content, the flour is considered to have high levels of tiny fragments of bran (Peyron et al., 2002). This gives flour a slight brownish colour and can cause a reduction in dough elasticity resulting in poor baking qualities. The aim of the flour miller is to minimise the ash content as much as possible. This can mostly be achieved by conditioning the wheat prior to milling as discussed earlier. However, most of the cleanest flour with low ash content is collected after the first break rollers with relatively low flour yield. Developing a method to keep the bran intact and reduce bran/endosperm adhesiveness is thus economically important.

#### 1.1.3 Hard and soft wheat

During milling the grains are crushed and sheared between two break rollers rotating towards each other at a differential speed. The hardness of the wheat cultivar will determine how the starch granules break during this process. Soft and hard wheat have different fracturing patterns when they are passed through the break rollers. Soft wheats tend to produce less mechanically damaged starch granules than hard wheats (Oliver, 1998; Fang and Campbell, 2002). This form of starch damage generally occurs when the starch granules break into smaller fragments caused by the shearing forces of the break rollers. In this case, fracturing tends to occur along the starch 'cell wall', and is termed 'intra-cellular' fracturing. In hard wheat, the shearing force from the break rollers causes

'inter-cellular' fracturing, a fracturing that occurs through the starch granule (Moss et al., 1980).

The two main properties of mechanical starch damage are increased water absorption and a reduction in dough viscosity (Evers et al., 1999). The consequence of starch damage for bakers is that more water will be required when forming dough with flour that has high starch damage when compared to flour with low starch damage (Oliver, 1998).

In bread baking, the baker needs to be certain that when making dough, the flour will require a set volume of water which will create a reasonable dough texture every time. This is important in getting a consistent final baked bread of predictable loaf volume, air pocket size, wall thickness and crumb texture. To achieve a standard flour quality, grains are blended prior to milling and/or flour is blended after milling (Bread Research Institute (BRI) Australia, <u>www.bri.com.au</u>; Kuakpetoon et al., 2001). Grains are first tested for quality, protein content, and hardness. Grains with high protein content and hardness are blended to produce flour for bread baking, whereas soft wheats with low protein content are blended for flour used in baking cakes and biscuits. Blending of wheat cultivars and/or flour is thus important in providing a standard flour for each specific baking product and also to eliminate uncertainty when adding water to get the required dough texture (Personal Communication M. Southen, BRI; Kuakpetoon et al., 2001).

#### 1.2 Protein distribution

Starch grain size in the endosperm is generally larger around the centre of the grain (type-A) and tends to be smaller (type-B) towards the edges adjacent to the aleurone cells (Fig. 1.2). The endosperm cell walls (Fig. 1.2) surrounding the starch granules are primarily comprised of arabinoxylans (AX) and  $(1\rightarrow3, \text{ and } 1\rightarrow4)$ - $\beta$ -glucans. Protein concentration is low around the centre of the endosperm, increasing as the starch granule size decreases nearer to the aleurone cell wall (Tervilä-Wilo et al., 1996; Evers et al., 1999). For whole grains, most of the total protein pool is found in the large endosperm fraction; however, when based on milligrams of protein per gram of tissue fraction, the protein content in the embryo, scutellum and aleurone tissue fraction is much higher than the endosperm and pericarp/testa (see Table 1.1).

 Table 1.1 Distribution of proteins, fraction weight and starch in the wheat grain

Grain Fraction	% of grain Weight	% Total protein	mg protein per gram of tissue fraction*	% of Total starch
Outer and intermediate fraction (Pericarp, Testa)	8	4.5	56	0
Inner fraction (aleurone cells)	7	15.5	221	0
Endosperm	82.5	72	87	100
Scutellum	1.5	4.5	300	0
Embryo	1	3.5	350	0

(Spurway, 1998)

\* Based on milligrams of protein per gram of grain fraction

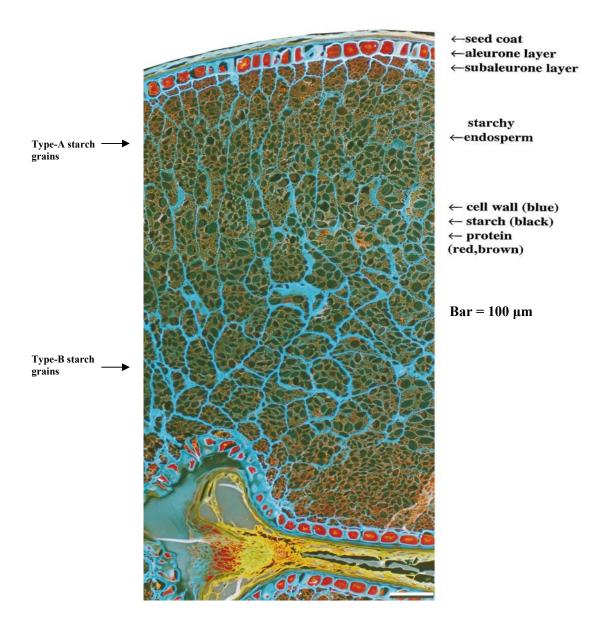


Figure 1.2 Cross section of wheat grain (Tervilä-Wilo et al., 1996).

#### 1.2.1 Major hydrolytic enzymes

*α*-amylase and *β*-amylase are hydrolytic enzymes found in wheat. They are both classified as *α*-(1→4)-D-glucanases and are involved in hydrolysis of the starchy endosperm into oligo-and monosaccharides to feed the developing plant during germination (Evers et al., 1999; Every et al., 2002). *α*-amylase is an endo-acting enzyme, in that it hydrolyses starch at random points along the polysaccharide chain. *β*-amylase however, can hydrolyse starch only at the reducing end of the polysaccharide chain. *α*amylase is mainly found in the scutellum of the embryo. At the begining of germination, *α*-amylase is secreted into the endosperm and begins starch hydrolysis. The aleurone cells then start to express and secrete *α*-amylase, as this whole cascade is triggered by hormones that are released from the scutellum upon hydration. *β*-amylase is contained in the endosperm and it is inactive when the grain is dry. Upon hydration, hydrolysis of starch by *β*-amylase is quite slow, however it is greatly increased when *α*-amylase is secreted by the scutellum and aleurone cells into the endosperm. *α*-amylase to hydrolyses starch into smaller fragments to expose more reduced ends for *β*-amylase to hydrolyse it further into maltose (Cejudo et al., 1995; Every et al., 2002).

Apart from starch, arabinoxylans (AX) and  $(1 \rightarrow 3, \text{ and } 1 \rightarrow 4)$ - $\beta$ -glucans are the most abundant carbohydrates in the remnant cell walls of the endosperm (Evers et al., 1999). Water-extractable AX's give flour its viscous property when adding water to make the dough. AX's which cannot be extracted with water give flour its water absorptive properties (Courtin and Delcour, 2002). Endoxylanase is found in wheat endosperm at very low levels. During germination, the AX cell walls are hydrolysed by endoxylanase,

providing the amylases greater access to in turn hydrolyse starch (Courtin and Delcour, 2002).

#### 1.2.2 *a*-amylase and its effect on flour quality

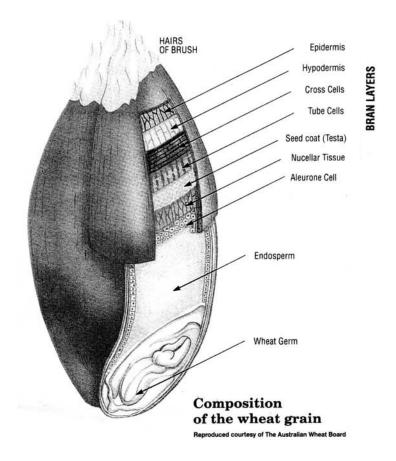
The presence of high levels of  $\alpha$ -amylase in flour is an indicator of potentially poor quality flour.  $\alpha$ -amylase greatly increases the breakdown of starch into smaller fragments, resulting in flour losing dough viscosity (Evers et al., 1999). The presence of high levels of  $\alpha$ -amylase indicates that germination has begun. This may be a result of late harvesting and/or water damage in the field or exposure to moisture during storage. A standard measure of grain quality, called the "falling number" is related to the level of  $\alpha$ -amylase present in grain. The number is the time in seconds it takes a plunger device to fall to the bottom of a mixture of water and ground-up whole grain. High falling number indicates better quality grain, as the mixture will be more viscous, thus decreasing the speed at which the plunger is falling. For good quality flour, the falling number should not be below 250 (Grain Foods Cooperative Research Centre (GFCRC),

<u>www.grainfoodscrc.com.au</u>). Other factors may also affect the falling number, such as protein and ash content and starch damage (Evers et al., 1999; Every et al., 2002).

#### 1.3 Bran layers in grain development

#### 1.3.1 Bran layers

The outermost component of the grain is the bran, which is composed of about seven distinct tissue layers as mentioned previously (Fig. 1.3). The outermost layers of the bran are collectively called the pericarp. The pericarp consists of the epidermis (being the outermost layer), hypodermis, cross cells and finally the tube cells. The next three tissue layers are the testa, nucellar and aleurone. The aleurone tissue is in contact with the endosperm and it is the only bran tissue layer that is still alive and functional at the cellular level in mature grain and is critical during germination. Bran has an important function as a protective barrier for the grain. The grain is a potential food for insects, fungi, and bacteria, and it is also exposed to many environmental stresses, thus the bran must have properties which protect it from all these factors. Much work has been done on the extraction and isolation of defence proteins in wheat. These proteins are categorized into major classes of defence proteins are termed pathogenesis-related proteins (PR), and include PR-1, PR-2 (1, 3- $\beta$ -glucanases), PR-3 (chitinases), PR-4 (wheatwin), and PR-5 (thaumatin-like proteins) (Selitrennikoff 2001; Desmond et al. 2006).



**Figure 1.3** Bran tissue layers (modified from The Australian Wheat Board (AWB), www.awb.com.au).

#### 1.3.2 Formation of bran layers during grain development

Wheat is a self-pollinating plant, where the pollen is released from the anther (at anthesis) and attaches to the stigma. The gamete is then delivered to the ovum which then becomes fertilized. The triploid gamete and ovum fuse together to form a hexaploid zygote. The zygote begins cell division and forms into an embryo, which contains the primordial plant organs. During grain development, the embryo develops together with food and nutrient

storage compartments, and protective outer tissue layers. Development is complete at around 30 to 40 DPA. The grain becomes dormant at the end of development until the right conditions allow it to germinate (Home-Grown Cereals Authority (HGCA), <u>www.wheatbp.net</u>). There are many physiological changes that occur during grain development, however for the purpose of this paper only bran tissue layers will be discussed.

During grain development, there are around 16 different cell types that ultimately form the outer bran tissue. Once grain development is complete at around 30 DPA, only seven distinct layers can be seen (Fig. 1.4). The early tissue layers are present up to around 11 DPA, and are involved in providing food and nutrients for the developing grain. The green colour of the developing grain is due to chloroplasts inside the cross cells. They gradually collapse towards the end of grain development and give the grain a brown colour. At around 11 DPA grain filling begins, accumulating starch to form the endosperm. The outermost cells of the endosperm differentiate to become aleurone cells (Drea et al., 2005). At this stage, all of the tissue layers present will become part of the bran. The changes that occur are due to compression caused by grain filling. The inner epidermis is compressed as the endosperm fills out and is torn and crushed, forming the tube cell layer. The inner integuments collapse and form the seed coat (testa), whereas the outer integuments are absorbed and disappear. Finally, the nucellus and nucellar epidermis compact together to form the nucellar tissue that surrounds the aleurone layer (Fig. 1.4) (Home-Grown Cereals Authority (HGCA), www.wheatbp.net).

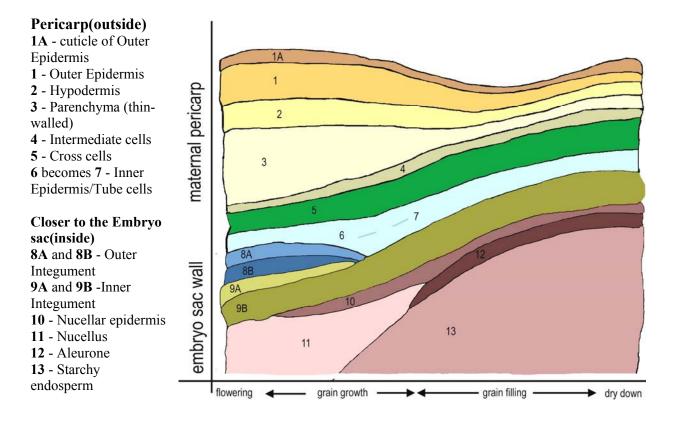


Figure 1.4 Bran tissue layer formation during wheat grain development (from Home-

Grown Cereals Authority (HGCA), www.wheatbp.net).

#### 1.3.3 Effects on bran layers during germination

Grain is harvested at around 30 DPA when it is in the dormant stage. Given that the moisture content of the grain remains low at around 10%, the grain should not germinate. However, once the moisture content increases, many biochemical processes begin and the embryo begins to grow.

There are many hormones involved at the early stages of development. The hormones abscisic acid (ABA) and gibberellic acid (GA) are important in determining if the grain remains dormant. ABA acts as a germination suppressant and GA has the opposite effect. The embryo of the grain is the initiator of germination and is where growth-promoting hormones are produced and or released. Experiments have shown that when the embryo was dissected from a grain and then imbibed in a solution containing GA, germination mechanisms are activated in the aleurone cells (Bethke et al., 1998). When ABA levels decrease and GA levels increase, the aleurone cells begin to breakdown and release  $\alpha$ -amylases into the endosperm. As discussed earlier,  $\alpha$ -amylase works together with  $\beta$ -amylase to break down the starchy endosperm into simpler sugars. This dual process in starch hydrolysis is important in supplying the embryo with a carbon source. Another hormone that is involved in germination is auxin, or indole-3-acetic acid (IAA). It is involved in the mobilization and breakdown of phytin (potassium, magnesium, calcium salt of inositol hexaphosphate), which is a macronutrient storage compound found mainly in the aleurone cells (Eastwood and Laidman, 1971; Fincher, 1989; Evers et al., 1999).

#### 1.4 Analysis of protein composition in wheat grain

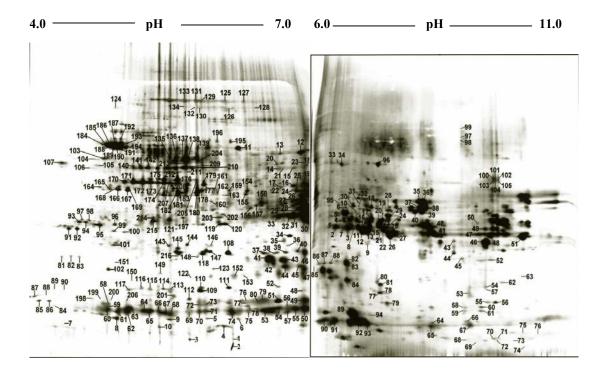
#### 1.4.1 High resolution 2-dimensional electrophoresis (2-DE)

The first step in proteomic work is to extract proteins from the tissue sample. The extracted proteins are then solubilised into appropriate buffers for first-dimension separation.

The first-dimension separation in 2-DE separates proteins based on their intrinsic pI by isoelectric focusing (IEF). The pI of each protein represents the pH required for it to have a neutral charge. Immobilised pH gradient (IPG) strips can range from pH 1 to pH 13. An electric current is applied across the strip which mobilizes the proteins. Each different protein will stop migrating when it reaches its corresponding pI on the IPG strip. In situations where there is an accumulation of many proteins around a particular pI, focusing over a narrow pH range can improve separation and resolution (Fig. 1.5). The second dimension of 2-DE is the separation of proteins based on their molecular weight.

Once proteins have been separated by their pI, the IPG strip is equilibrated with sodium dodecyl sulfate (SDS) to coat the proteins with an overall negative charge. The strip is then transferred onto an SDS polyacrylamide gel for electrophoresis (PAGE). As the proteins travel electrophoretically through the polyacrylamide gel matrix, they begin to separate according to molecular weight. Low-molecular-weight proteins travel faster through the gel matrix and higher-molecular-weight proteins travel slower. After electrophoresis the gel is stained and ready for further analysis. Figure 1.5 shows the

complex mixture of proteins separated by 2-DE, using two narrow pH bands to improve resolution.



**Figure 1.5** Protein spots on 2-DE gel of wheat grain endosperm at 17 DPA. Approximately 200 ug of protein extract was loaded per gel and was stained with diamine silver (Skylas et al., 2000).

#### 1.4.2 Identification of protein spots in 2-D gels

Protein spots are excised from a 2-D gel and residual protein stain is washed out. Gel pieces are then dehydrated and digested with trypsin which fragments proteins at arginine and lysine residues (except those that are followed by a proline). After trypsin digestion, the peptides are extracted from the gel pieces for further analysis. Peptides can then be analysed using matrix assisted laser desorption/ionization time-of-flight mass spectrometry combined with tandem mass spectrometry (MALDI-TOF MS/MS) or electrospray ionization triple-quadrapole tandem mass spectrometry (ESI MS/MS).

MALDI-TOF MS can be used for peptide mass fingerprinting (PMF) and selected peptide masses can be further analysed by (MS/MS). The sample is co-crystalised with a matrix on a metal plate and a laser is used to ionize and desorb the peptides into the TOF analyzer. Resolution of a large mass range can be achieved using TOF as it measures the flight time of an ion between the source and the detector to give a calculated charge-tomass ratio. The effective length of the flight tube in TOF mass spectrometers can be increased by ion reflectors, which increase the resolution and mass range (Yates III, 1998; Chernushevich et al., 2001).

ESI triple-quadrapole MS/MS is different to MALDI TOF MS in that the sample is in solution prior to analysis and the ion masses are detected by selecting ions using a frequency-to-current ratio. The sample is sprayed into an electric field using an inert carrier gas. This causes the particles to desolvate and ionize without fragmenting. The ionized particles accelerate into a constant direct current electric field generated by two

longitudinal poles. Perpendicular to this electric field plane are two poles which have an alternating current. A frequency-to-current ratio is used to stabilize an ion with particular mass-to-charge ratio as it travels through the electric field. Stabilized ions which pass through the electric field are then detected, and ions that are not transmitted collide into the poles. A scan of different frequencies allows for different mass-to-charge ratio ions to pass through and hit the detector. The resolution of this type of MS is dependent on the electric field current and frequency. High frequency and current increases an ion's oscillation in its trajectory through the quadrapole and thus increases mass resolution (Yates III, 1998; Chernushevich et al., 2001; Lay Jr, 2001; Mak et al., 2006b).

Peptide masses and MS/MS peptide ion masses can then be used together to search against cereal and wheat expressed sequence tag (EST) databases through Mascot (Mascot, <u>www.matrixscience.com</u>). MS/MS ion masses can also be used to search against proteins in the wheat EST database, using The Grand Proteome Machine (thegpm) (thegpm, http://137.111.103.157/tandem/thegpm\_tandem.html).

Peptides can be sequenced by Edman degradation, also called *N*-terminal sequencing. The sequences can then be used for protein identification. Some of the limitations in using this method are that it takes more time to identify each protein and there is potential for the *N*-terminal of proteins to be blocked. Although this method is still in use, highthroughput techniques like peptide fragment sequencing using multi-dimensional protein identification technology (MudPIT), and the types of MS mentioned previously are more commonly used (Skylas et al., 2005).

#### 1.4.3 Wheat grain proteomics

Proteomics is essentially the separation and identification of proteins from any part of an organism. The total number of genes in an organism's genome is generally larger than the number of expressed proteins. Proteomics reveals those genes that are currently, or recently, expressed in different tissues and cell types (Skylas et al., 2000; Wong et al., 2004; Skylas et al., 2005).

As mentioned previously, it is important to understand the biochemical processes of wheat to determine favorable characteristics which can improve crop yield and management for farmers, and flour quality and yield for millers. Some of the recent studies on wheat using a proteomic approach have identified proteins in wheat grain endosperm, enzyme-modifying proteins in the endosperm, and proteins in the germ (Skylas et al., 2000; Wong et al., 2004; Mak et al., 2006b).

Proteomic studies on wheat grain endosperm by Skylas et al., (2000) were made at two stages of grain development, at 17 DPA and 45 DPA. More protein spots were observed at 17 DPA (around 1298 protein spots) than to at 45 DPA (1125 protein spots). Protein disulfide isomerase (PDI) isoforms and 60S ribosomal proteins are abundant during grain development and less abundant when the grain matures, thus possibly explaining the reduction in protein spots at 45 DPA. Because proteins were sequenced by *N*-terminal Edman degradation, *N*-terminal blocking prevented 28% of proteins in the endosperm at 17 DPA being sequenced. This problem may be overcome by using the mass spectrometry protein identification procedures mentioned previously. Nonetheless, this work highlights the dynamic changes in gene expression that occur during wheat grain development together with a large number of identified endosperm proteins. The identification of these proteins may be useful in flour trait correlation with protein quantity and type.

Proteomics has also been used to identify thioredoxin target proteins in wheat grain endosperm by Wong et al. (2004). Thioredoxin is present in plants and acts as an enzyme regulator, a process which is called redox regulation. It does this by activating enzymes or altering the enzyme activity via disulfide bond reshuffling. This work has identified a large number of thioredoxin target proteins, thus increasing knowledge of which proteins are modified during the development of the endosperm.

Proteomics of germ and whole grain (Mak et al., 2006b) has recently identified many major proteins in the germ, and has shown major protein class differences in the germ compared to proteins in the endosperm identified by Skylas et al. (2000). This work has added to knowledge concerning major proteins in the germ and is potentially useful in breeding programs where nutritional value and wheat quality can be improved.

Further proteomics work has compared the germ of wheat affected by Black Point with non-affected wheat (Mak et al., 2006a). Black Point disease affects many wheat growing countries. The disease causes an undesirable discoloration of the germ and the bran in severe cases, and as a result the value of the crop is down-graded. The study has shown that cultivars not affected by Black Point had higher levels of stress related proteins compared to non affected cultivars (Mak et al., 2006a). This could lead to better understanding of the function of these proteins in the germ and their role in Black Point resistance.

#### 1.4.4 Proteomics of bran components

Proteomic analysis of bran and bran tissue fractions have not been reported. The difficulty with collecting clean bran from dry grain is that there is a strong bond between the endosperm and aleurone cells. This makes it difficult to cleanly separate the aleurone cells from the endosperm, to make the bran free from contaminants and suitable for protein analysis. Another problem is separating the bran tissue fractions from each other. It is difficult to separate the bran tissue layers from dry grain as the layers are brittle and firmly bonded together.

There are methods for collecting aleurone cells and other tissue fractions by various techniques such as floating, differential centrifugation and electrostatics. However, they mostly result only in tissue enrichment, which still may contain contaminants from other tissue fractions (Bacic and Stone 1981; Stone et al. 1988). This makes these methods unsuitable for proteomics studies.

Imbibing whole grain in water will soften the bran, making it more flexible and increasing bran/endosperm separation. Manual bran tissue separation in this way has been reported by Antoine et al. (2004). The idea is to initiate germination and begin the endogenous breakdown of endosperm at the aleurone endosperm interface by  $\alpha$ -and  $\beta$ -

amylases. This then softens the endosperm around the aleurone surface and allows the bran to be removed easily and washed further with water. Softening of the bran tissue allows it to be more easily separated manually into three distinct tissue layers, the outer layer (epidermis and hypodermis), intermediate layer (cross cells, tube cells, testa and nucellar tissue), and inner layer (aleurone cells). Any water-soluble proteins that may leach out during grain imbibition can be collected and analysed.

The bran tissue collected using this method will be representative of bran at the early stages of germination. For the purpose of this study, the focus is on protein composition of bran at this early stage, as it will be similar to bran from conditioned grain. Collecting bran tissues from dry (dormant) grain and comparing them to bran at the early stages of germination will give more information of the changes in gene expression. Improvements in bran tissue separation from dry grain would be required to do further work in this area.

As the conditioning step in milling is about stabilizing bran and improving the separation of endosperm from bran, knowledge of unique proteins which may aid in this process could potentially provide insights into how the conditioning and milling process could be optimised. A very optimistic goal in milling would be to achieve complete endosperm and bran separation. However, even a slight increase in flour yield and quality after milling would have a large impact, as millions of tonnes of wheat grain are milled annually.

# 2. Materials and Methods

# Materials 2.1

# 2.1.1 Wheat sample

Wheat grain *Triticum aestivum* cultivar Babbler, was supplied by the Bread Research Institute Australia Limited (BRI).

#### 2.1.2 Chemicals

#### 2.1.2.1 Protein extraction

Acetone (AJAX Finechem), ammonium acetate (BDH Chemicals), glycerol (AJAX Finechem), 2-mercaptoethanol (Sigma-Aldrich), methanol (AJAX Finechem), SDS (Amresco), sucrose (AJAX Chemicals), trichloroacetic acid (TCA) (BDH Chemicals), Tris (Amresco), Tris-buffered phenol at pH 8.0 (Sigma-Aldrich).

## 2.1.2.2 Protein quantification

Bovine serum albumin (BSA) (Sigma-Aldrich), Bradford reagent (Bio-Rad), nitrocellulose paper (0.2 μm pore size, Whatman, Schleicher and Schuell).

#### 2.1.2.3 SDS-PAGE

Bromopheonol blue (Edward Gurr), glycerol (AJAX Finechem), Pre-cast SDS 4-20% gradient polyacrylamide gels (0.1 cm x 10 cm x 8 cm) (Life Therapeutics, Long Life Gels), 2-Mercaptoethanol (Sigma-Aldrich), SDS (Amresco), Tricine (Sigma-Aldrich), Tris (Amresco).

#### 2.1.2.4 IEF and SDS-PAGE

Acrylamide (Bio-Rad), agarose (Bio-Rad), ammonium persulphate (APS) (Bio-Rad), bisacrylamide (Bio-Rad), bromopheonol blue (Bio-Rad), carrier ampholytes (Amersham Biosciences, GE), 3-[(3-cholamidopropyl) dimethylammonio]-1-propanesulfonate (CHAPS) (Bio-Rad), 1,4-dithio-DL-threitol (DTT) (Bio-Rad), glycerol (Merck), glycine (Bio-Rad), IPG strips pH 4-7 (Bio-Rad, Catalogue number 163-2008), IPG strips pH 6-11 (Amersham Biosciences, Lot number 309990), SDS (Bio-Rad), sulfobetaine 3-10 (Sigma-Aldrich), N,N,N',N'-tetramethylethylenediamine (TEMED) (Bio-Rad), thiourea (Sigma-Aldrich), tributyl phosphine (TBP) (Sigma-Aldrich), Tris (Sigma), urea (Bio-Rad).

# 2.1.2.5 Gel staining

Acetic acid (Merck), Deep Purple (Amersham GE Health Care), G-250 Coomassie (Bio-Rad), methanol (Merck), phosphoric acid (BDH Chemicals), sodium carbonate (Sigma-Aldrich).

# 2.1.2.6 Sample preparation for MS

Acetonitrile (Merck), ammonium bicarbonate (Sigma-Aldrich), C-18 Zip Tips (Eppendorf), formic acid (AJAX Finechem), N-terminal sequencing grade 25% trifluoroacetic acid (TFA) (Applied Biosystems), Prep mix (bradykinin, angiotensin I, neurotensin and adrenocorticotropic hormone (ACTH) fragment) (Sigma-Aldrich), recrystalised α-cyano-4-hydroxycinnamic acid (Sigma-Aldrich), sequencing grade trypsin (Promega, Catalogue number V5111).

# 2.2 Tissue fraction collection

#### 2.2.1 Grain treatment prior to tissue separation

Whole grains were imbibed in water for two days at room temperature. Once the endosperm had softened, each end of the grain was cut off with a fine pair of scissors, removing the embryo and brush hairs. The cleft running along the grain was also cut out using scissors. Finally, the endosperm was removed and the bran was washed with water. The washed bran was kept in fresh milli-Q water and stored at approximately 4°C.

#### 2.2.2 Bran cleaning and tissue separation of inner and intermediate fractions

Bran tissue was further washed under a dissecting microscope using forceps to scrape off any remaining endosperm. During dissection, the bran was kept in a volume of water (approximately 200 µl) on the surface of the dissecting microscope (Kyowa, Tokyo). This was needed to collect the inner fraction (aleurone cells) and to stop the bran from drying out. The inner fraction was scraped off into water on the glass dissecting surface of the microscope with forceps and collected using a pipette. The cells were then placed into a 1.5-ml plastic tube and centrifuged for 2 min at 14000 rpm. Excess water was removed and the cells were left to dry inside the tube. The outer layer of the remaining bran was removed and collected into a plastic tube. Finally, the remaining intermediate tissue fraction was further washed with water under the dissecting microscope and was also collected into a plastic tube. All tissue fractions were left to dry overnight. Once the tissue fractions were dry, they were stored at -20°C. A light microscope was used to visualize each bran fraction to confirm its purity. The method used to separate the tissue fractions was successful in that all the tissue fractions had unique cell structure, indicating the tissue types present. Micrographs of the tissue fractions were all captured at 200x magnification using a light microscope (Olympus BX50) and digital microscope camera (Sony DFW-SX700) and were edited using Image J software. The same digital camera was used with a stereo zoom microscope (Olympus SZH) to capture larger whole images of the bran fractions (Figs. 3.1, 3.2, and 3.3a). An Olympus BH2 epifluorescence microscope equipped with a mercury lamp and a filter set for UV excitation at 395 nm and emission at 420 nm was used to image the autofluorescent inner fraction. The UV image was captured using a Nikon DXM 1200F (Fig. 3.3b).

The separation and collection of inner and intermediate bran tissue fractions was successful, and enough of each tissue sample was collected for protein extraction. However, much more of the outer tissue fraction was required because of its low protein content.

#### 2.2.3 Improved method for collecting outer fraction

Grains were imbibed in water for 5 min, followed by freezing with dry ice. Once frozen, the grains were left to thaw out at room temperature. After thawing, the outer fraction was easily removed using forceps with the aid of a dissecting microscope (Fig. 2.1). The brush hairs were cut off with scissors and the remaining tissue was stored in a plastic tube to dry overnight at room temperature. The dry tissue was then stored at -20°C.

Amount of tissue collected for protein extraction – inner fraction (Aleurone) (45 mg), intermediate fraction (42 mg), and outer fraction (200 mg).



**Figure 2.1** Removal of the outer fraction (Hypodermis and epidermis) using the 'freeze thaw' method.

# 2.3 Protein extraction and quantification

#### 2.3.1 Protein extraction from bran fractions

#### 2.3.1.1 Tissue preparation and washing

Samples were kept on ice during all steps in the protein extraction. All centrifugation was done using a 1.5-ml plastic tube centrifuge (Sigma).

The method used for protein extraction from plant tissue was followed as described by Wang et al. (2003).

The tissue sample was placed into a 2-ml screw-cap plastic tube and washed with 1 ml cold acetone. The sample was vortexed for 30 s and then centrifuged at 14000 rpm for 3 min. Acetone was removed and this step was repeated. The tissue was then left to dry inside the plastic tube in a fume hood. Once dry, the tissue was placed into a mortar, together with a small amount of acid-washed sand, and was ground down to a fine powder. The fine powder was collected back into the 2-ml plastic tube.

One millilitre of cold 10% TCA in acetone was added to the ground-up tissue. The sample was then vortexed for 30 s and then centrifuged at 11000 rpm for 3 min. The solution was removed and this step was repeated twice. The sample was then washed twice with cold 10% TCA in water and then finally washed twice with cold 80% acetone. Again for each step, the sample was vortexed for 30 s and then centrifuged at 11000 rpm for 3 min. The tissue was left to dry at room temperature overnight.

#### 2.3.1.2 Protein extraction

To the washed and dried tissue was added 800 µl of Tris-buffered phenol at pH 8.0, followed by 800 µl of SDS buffer (30% sucrose, 2% SDS, 0.1M Tris-HCl at pH 8.0, and 5% 2-mercaptoethanol). The sample was then vortexed for 30 s and centrifuged at 11000 rpm for 3 min. The top phenol layer was removed and placed into a new 2-ml plastic tube.

# 2.3.1.3 Protein precipitation

To the phenol was added five volumes of cold methanol, 0.1 M ammonium acetate. The tube was then stored at -20°C for 1 hr to precipitate the protein. The precipitated protein was centrifuged at 11000 rpm for 5 min and the supernatant was removed. The protein pellet was washed twice with cold methanol, 0.1 M ammonium acetate and then twice with cold 80% acetone. Each time, the pellet was vortexed for 30 s and centrifuged for 5 min at 11000 rpm. After final wash, the pellet was left to air dry to evaporate any acetone.

#### 2.3.1.4 Solubilising protein pellet in rehydration solution

All protein pellet samples were solubilised in 500 µl rehydration buffer (5 M urea, 2 M thiourea, 65 mM DTT, 2% CHAPS, 2% sulfobetaine 3-10, 1% carrier ampholytes, 40 mM tris, 0.002% bromophenol blue dye, milli-Q water).

#### 2.3.2 Protein collection from grain and outer fraction supernatant

Approximately 35 to 40 whole grains, and 37 mg of isolated outer fraction collected from 50 grains, were placed into separate 1.5-ml plastic tubes and filled with Milli-Q water. The tubes were stored at 4°C overnight to allow for any water-soluble proteins to leach out. The samples were then stored at -80°C prior to freeze drying. 50 µl 2x loading buffer was added to the freeze-dried samples to solubilise the proteins. The samples were then centrifuged for 5 min at 14000 rpm to remove any tissue material. The supernatant was collected into fresh 1.5-ml plastic tubes.

#### 2.3.3 Protein quantification

#### 2.3.3.1 Protein quantification of tissue fractions

Nitrocellulose paper was pre-wet with milli-Q water for 15 min. The paper was then placed onto a 96-well Bio-Dot micro filtration apparatus. An aliquot of water (100  $\mu$ l) was loaded into each well and was drawn through the paper using a low vacuum. This was done to check if the apparatus is sealed properly.

Bovine serum albumin (BSA) was diluted in water for one dilution series and rehydration buffer for a comparison as a standard curve. The dilutions were 200 µg.ml<sup>-1</sup>, 50 µg.ml<sup>-1</sup>, 12.5 µg.ml<sup>-1</sup>, 3.125 µg.ml<sup>-1</sup>, 781 ng.ml<sup>-1</sup>, 195 ng.ml<sup>-1</sup>, 48.8 ng.ml<sup>-1</sup> and a blank. An aliquot (100 ul) of each dilution was loaded onto the 96-well Bio Dot apparatus in triplicate. Samples were diluted 1:10, 1:100, and 1:1000. Each well was then washed with 150 µl water four times, each time allowing the vacuum to draw the water through the membrane. The membrane was removed from the apparatus and rinsed several times with water. It was then placed in fresh water on rocker for 10 min and rinsed again. The membranes were placed in between two filter papers wrapped in foil and stored in the fridge overnight.

The Deep Purple staining method was followed as described in the manufacturer's instructions for nitrocellulose blot staining (Amersham Biosciences). Membrane was washed with 200 mM sodium carbonate (Na<sub>2</sub>CO<sub>3</sub>) for 15 min. The wash solution was replaced with 25 ml 1:200 dilution of Deep Purple stain in water. The membrane was left on a rocker to stain for 1 h and kept covered from light. The membrane was left to dry in the dark and then scanned using a Typhoon variable mode imager (Amersham Biosciences). The membrane image was analysed using Image J software to measure the relative intensities of the protein spots. From the relative intensities, a standard curve was calculated for BSA dilutions in water and the same for BSA dilution in rehydration buffer. The blanks were used to subtract background intensities and then protein concentrations were determined.

#### 2.3.3.2 Protein quantification of water-soluble proteins

The water-soluble proteins in the supernatant were quantified using Bio-Rad protein assay dye reagent (Catalogue number 500-0006). Proteins from the supernatant were extracted as described earlier. After freeze drying, the samples were solubilised in 250 µl of Milli-Q water. BSA dilutions (160 µg.ml<sup>-1</sup>, 80 µg.ml<sup>-1</sup>, 40 µg.ml<sup>-1</sup>, 20 µg.ml<sup>-1</sup>, 10 µg.ml<sup>-1</sup>, 5 µg.ml<sup>-1</sup>) for a standard curve were prepared. Protein assay dye was diluted, one part to four parts Milli-Q water. An aliquot (200 µl) of this dilution was added to wells in a 96-well microplate (Greiner, Catalogue number 655101). Ten microlitres of each BSA standard and sample was added to the protein assay dye in duplicate. The plate was left to incubate at room temperature for 5 min prior to reading absorbance at a wavelength of 620 nm (Labsystems Multiskan).

# 2.4 Iso-electric focusing (IEF) (First dimension in 2-DE)

### 2.4.1 Sample preparation

Prior to IEF, samples in rehydration buffer were reduced by adding 5 mM tributyl phosphine (TBP) and alkylated by adding 10 mM acrylamide. The sample was then left on the bench to incubate for 1 h. After incubation, the samples were centrifuged (Eppendorf centrifuge 5417R) at 14000 rpm for 10 min at 4°C. The required total volume of sample in buffer was dependent on the type of immobilized pH gradient (IPG) strip used. For the 17 cm pH 4-7 IPG strip, the sample volume per strip was 300 µl and for the 18 cm pH 6-11 IPG strip, the sample volume was 120 µl, as it is recommended to use cup loading (see below) in the alkaline pH range.

### 2.4.2 IPG strip rehydration

Sample loading onto IPG strips was different for acidic pH ranges and alkaline pH ranges. For the 4-7 pH range samples could be loaded onto the IPG strip during rehydration. On the other hand, the manufacturer (Amersham Biosciences, GE) recommends cup loading for the 6-11 pH range. Firstly, the IPG strip was rehydrated with the rehydration solution. When placing the IPG strip onto the IEF machine, a cup

attachment was placed onto the strip at the acidic end. The sample was then loaded into the cup and the sample was electrophoresed out from an opening at the bottom of the cup and into the strip. This means that only proteins with an alkaline pI migrated onto the IPG strip and not the acidic pI proteins. The result of using this method was better protein focusing in the alkaline pH range.

The 4-7 pH IPG strips were rehydrated for 4 h with 300  $\mu$ l of the prepared sample. The 6-11 pH IPG strips were rehydrated for 4 h with 300  $\mu$ l of rehydration buffer alone. An aliquot (120  $\mu$ l) of the prepared sample in buffer was cup loaded during IEF.

# 2.4.3 IEF

IPG strips were focused overnight at 8000 V for 11.5 h, on an IEF machine (Ettan IPGphor3, GE Healthcare) (method: 300 V for 4 h, ramp to 8000 V over 8 h, finally remained at 8000 V for 11.5 h).

Total V hr for each IEF run:

Inner fraction	pH range 4-7 – 129000	
Inner fraction	pH range 6-11 – 118649	
Intermediate fraction and outer fraction	pH range 4-7 - 127820	
Intermediate fraction and outer fraction	pH range 6-11 – 107818	

After focusing, the IPG strips were placed into a tray and sealed with cling wrap to be stored at -80°C overnight or when ready for SDS-PAGE. It is recommended to store the IPG strips at -80°C at least overnight for better second-dimension resolution.

#### 2.4.4 IPG strip equilibration

The IPG strips were removed from the -80°C freezer and were thawed at room temperature. The tray was placed onto a rocker and equilibration solution (6 M urea, 2% SDS, 0.375 M Tris/HCl pH 8.8, 20% glycerol, 5 mM TBP, 2.5% acrylamide) was poured over the strips and left to equilibrate for 15 min. This was repeated twice, and after each time, the equilibration solution was poured off and replaced with fresh solution.

# 2.5 SDS-PAGE (Second dimension in 2-DE)

# 2.5.1 SDS-PAGE of IPG strips from tissue fractions

#### 2.5.1.1 Casting 8-18% polyacrylamide gradient gels

Glass plates, spacers and gel cast chambers (Protean II systems chamber) were used to cast six gels (17 x 17 cm) at a time. An 8% acrylamide buffer solution was prepared containing 44 ml 5x Tris/HCl buffer, 44 ml 40% bis-acrylamide, and 132 ml Milli-Q water. An 18% acrylamide buffer solution was prepared containing, 44 ml 5x Tris/HCl buffer, 99 ml 40% bis-acrylamide, and 77 ml 50% glycerol. To polymerise the acrylamide, 36 µl TEMED followed by 363 µl 10% APS was added to each buffer solution. The solutions were then stirred and immediately poured into two separate gradient pouring chambers (Bio-Rad, Gradient Former Model 395). The 18% acrylamide solution was poured into the reservoir chamber and the 8% acrylamide solution into the mixing chamber which contained a magnetic stir bar rotated gently by a magnetic stirrer (Selby Australia). A cartridge pump (Masterflex L/S) was used to draw the acrylamide solution out of the mixing chamber and into the gel casts. The 8% acrylamide solution

was first pumped into the gel casts at a rate of 60-75 ml.min<sup>-1</sup> until there was an even layer of about 1 cm on the bottom of the casting chamber. The 18% acrylamide solution in the reservoir chamber was released into the mixing chamber. The gels were poured until the acrylamide solution reached 2 cm from the top of the glass casts. The pump was then stopped and the inlet hose was clamped. Isobutanol was poured on top of the gels to create an even layer. Once the acrylamide had polymerized, the isobutanol was washed off with water and the gels were ready to be used.

#### 2.5.1.2 Electrophoresis

IPG strips were imbedded on top of the 8-18% gradient polyacrylamide gels (17 cm x 17 cm) using hot agarose (0.5% agarose, 0.001% bromopheonol blue, 192 mM glycine, 0.1% SDS, 24.8 mM Tris base pH 8.3). The gels were then electrophoresed in Protean II multi-cell tanks (Bio-Rad) using a power box (Bio-Rad Power Pac 3000), set at 5 mA per gel for 30 min, and then at 40 mA per gel for approximately 4.5 h, or until the dye front had run off the gel. Once the dye front had run off, gels were removed from their casts and placed into fixing solution (30% methanol, 7.5% acetic acid) for at least 1 h to prepare for Deep Purple staining.

#### 2.5.2 SDS-PAGE and protein analysis of water-soluble proteins from supernatant

An aliquot (50  $\mu$ l) of 2x 1D SDS loading buffer (100 mM Tris-HCl pH 6.8, 0.2% bromophenol blue, 20% glycerol, 4% SDS, 200 mM 2-mercaptoethanol) was added to the tube to solubilise the protein. The tube was vortexed for 1 min and then centrifuged for 5 min at 14000 rpm. The supernatant was collected and placed into a fresh plastic tube. The

sample was then reduced by adding 1  $\mu$ l 2-mercaptoethanol, incubated in a boiling water bath for 2 min to denature the proteins, and centrifuged for 2 min at 14000 rpm. An aliquot (30  $\mu$ l) of this sample was then loaded onto a 4-20% gradient pre-cast SDS polyacrylamide gel (0.1 cm x 10 cm x 8 cm). The sample was run using a Bio-Rad Power Pac 200 at constant 150 V for approximately 1 h. The gel was then stained with Coomassie and selected bands were excised for MS identification.

# 2.6 Staining, imaging and protein spot excision

# 2.6.1 Deep Purple staining

After fixing, the solution was poured off and replaced with 200 mM Na<sub>2</sub>CO<sub>3</sub> for 1 h to basify the gels. Sodium carbonate was then poured off and replaced with water, approximately 10x the gel volume. Deep Purple stain was added to the water to make a final dilution of 1 in 200. The gels were covered with foil and left to stain overnight on a rocker. After staining, the stain was poured off and destained twice with 1% acetic acid. The gels were immediately scanned using a Typhoon variable mode imager (Amersham Biosciences). The gel image was scanned in fluorescence mode, 610 BP Deep Purple emission filter, green (532) laser, and with 100 micron pixel resolution.

#### 2.6.2 Protein spot selection

The scanned images were transferred and uploaded into image analysis software Progenesis Discovery version 2005 (Nonlinear Dynamics LTD). The uploaded images were transferred to, and edited using, Progenesis PG240 version 2006 (Nonlinear Dynamics LTD) software to match and number protein spots in each of the triplicate gels. This software was used to match spots only in the intermediate and outer fraction samples to identify as many spots as possible. The protein spots in the inner fraction gel images were selected on the basis of proteins visible with Coomassie stain. The images were scanned on the spot cutter (Bio-Rad, EXQuest) and only spots that could be clearly seen with Coomassie stain were manually selected for excision.

# 2.6.3 Coomassie staining

After Deep Purple staining and scanning, the gels were counter stained with colloidal Coomassie Blue G-250 stain (17% ammonium sulfate, 3% phosphoric acid, 0.1% Coomassie G-250, 34% methanol, Milli-Q water to make up 1 L total) (Neuhoff et al. 1988) and left on a rocker overnight. The gels were then destained with 1% acetic acid and bagged in seal tight plastic bags provided in APAF lab. All gels were stored at 4°C until needed.

#### 2.6.4 Spot cutting

Prior to spot cutting, all gels were placed in Milli-Q water for at least 1 h. This was important to minimise any swelling of the gel during spot cutting. An image was taken of the gel and all spots visible with Coomassie stain were numbered and selected for excision. A spot cutter (Bio-Rad, EXQuest) was used to excise 96 spots at a time. It was found that the cutting becomes less accurate over time if more than 96 spots at a time were selected. Spots were placed into 96 well plates for further processing.

# 2.7 Peptide extraction for MS

#### 2.7.1 Gel plug destaining

Gel plugs were manually destained 3x with 120 µl wash solution (50% (v/v) acetonitrile, 25 mM ammonium bicarbonate). For each wash, the gel plug-in wash solution was placed onto an orbital shaker and incubated at 37°C for 10 min. The solution was removed and replaced with fresh wash solution each time. After destaining, the gel plugs were dried using a Savant Speed Vac Plus SC210A. For large gel samples, automated destaining was performed on Xcise - Automatated gel processing platform (Shimadzu Biotech) using standard "Wash and Destain" program. The same destaining solutions were used as described previously; however, the final drying step used 100% acetonitrile to dehydrate the gel plugs instead of the Speed Vac.

# 2.7.2 Passive trypsin digest

To each dry gel plug was added 8  $\mu$ l of 15 ng. $\mu$ l<sup>-1</sup> sequencing grade trypsin in 25 mM ammonium bicarbonate, pH 7.8. The plugs were then placed in the refrigerator for 1 h to allow the trypsin to be absorbed into the gel. After incubation, any excess trypsin was removed. The gel plugs were then sealed in the 96-well plate and incubated over night at 37°C.

#### 2.7.3 Peptide extraction from gel plugs (for Zip Tip clean up)

To each gel plug was added 10  $\mu$ l of extraction solution (10% acetonitrile, 0.1% TFA). The wells were sealed and the plate was placed in a water bath sonicator (Transsonic 700/H, Elma) for 20 min. After sonication, the seal on the well plate was removed to allow any excess acetonitrile to evaporate off for 30 min.

#### 2.7.4 C18 column Zip Tip clean up

Zip Tips were first washed with 10  $\mu$ l of 70% acetonitrile, 0.1% TFA by pipetting 10  $\mu$ l up and down three times. The tips were then further washed in the same manner using 0.1% TFA. The peptide extraction solution was taken up into the Zip Tip, drawing up and down 8 $\mu$ l. This was repeated ten times to concentrate the peptides onto the column. Once the peptides were loaded onto the Zip Tip, the tip was further washed three times with 10  $\mu$ l of 0.1% TFA.

# 2.7.5 Loading peptides onto ABI plate for MS

Four microlitres of extraction solution (4 mg.ml<sup>-1</sup> Matrix,  $\alpha$ -cyano-4-hydroxycinnamic acid, 70% acetonitrile, 0.1% TFA) was drawn up into the Zip Tip. The extraction solution in the tip was drawn up and down forming a drop at the end of the tip at least five times to elute the peptides from the column. Finally, 2 µl of this solution was spotted onto a designated circle marked on the ABI plate. A standard (prep mix with matrix) was also spotted after each sample on the ABI plate to externally calibrate using near point calibration with four peptide standards (bradykinin, angiotensin I, neurotensin and adrenocorticotropic hormone (ACTH) fragment). Samples on the plate were allowed to dry/crystallise, ready to be analysed using an Applied Biosystems 4700 MALDI TOF MS MS/MS (Foster City, CA).

# 2.8 Protein identification

#### 2.8.1 MALDI TOF MS

Protein peptide samples were analysed using an Applied Biosystems 4700 MALDI MS/MS with TOF/TOF optics (Foster City, CA) in reflector mode for positive ion detection. A Nd.YAG laser with wavelength and repetition rate of 355 nm and 200 Hz, respectively, was used. All MS spectra resulted from accumulation of 4000 laser shots (20 sub-spectrum were accumulated with 200 shots per sub-spectrum). Laser intensity varied between 3000 and 4000. Data was collected over a mass range of 750 to 3500 Da. Mass spectral data was analysed using Mascot (Matrixscience). Peak detection criteria for mass lists were MS: mass range 500-4000 Da, maximum 30 peaks per 200 Da, minimum signal to noise ration (S/N) 20, minimum area 200, maximum peak/spot 200, and for MS/MS: mass range 60 Da to precursor -15, maximum 20 peaks per 200 Da, minimum S/N 18, minimum area 300, maximum peak/spot 60. This converts the mass lists into Mascot and The Global Proteome Machine (GPM) compatible text files.

MS searches were done on cereal and wheat EST data bases on Mascot (Matrix Science, London, UK), which uses both MS and MS/MS data for protein identification. Searches using The Global Proteome Machine (GPM) wheat EST database uses MS/MS data for protein identifications. Identification of proteins with high scores and low e-values with good peptide matches and coverage were then tabulated with corresponding spot number and location on the 2D gel. Spots that showed no significant protein identification matches were further analysed using electro-spray ionization (ESI) MS/MS.

40

#### 2.8.2 ESI MS/MS

Digested peptides were separated by nano-LC using a CapLC system (Agilent 1100 Series, Agilent Technologies, Germany). Sample (39  $\mu$ l) was injected onto a peptide trap (Michrome peptide Captrap) for preconcentration and desalted with 0.1% formic acid at 10  $\mu$ l.min<sup>-1</sup>. The peptide trap was then switched into line with the analytical column containing C18 RP silica (SGE ProteCol C18, 300A, 3  $\mu$ m, 150  $\mu$ m x 10 cm). Peptides were eluted from the column using a linear solvent gradient, with steps, from H<sub>2</sub>O:CH<sub>3</sub>CN (95:5; + 0.1% formic acid) to H<sub>2</sub>O:CH<sub>3</sub>CN (20:80 + 0.1% formic acid) at 600 nl.min<sup>-1</sup> over 45 min. The LC eluent was subject to positive ion nanoflow electrospray analysis on an Applied Biosystems QSTAR XL mass spectrometer (ABI, CA, USA). The QSTAR was operated in an information dependant acquisition mode (IDA).

In IDA mode a TOF/MS survey scan was acquired (m/z 400-2000, 1.0 s), with the four largest multiply charged ions (counts >25) in the survey scan sequentially subjected to MS/MS analysis. MS/MS spectra were accumulated for 1 s (m/z 50-2000).

The LC/MS/MS data was searched using Mascot (Matrix Science, London, UK). Mascot was used to search cereal entries in the NCBI non-redundant protein database. High scores in the database searches indicated a likely match, confirmed or qualified by inspection of the spectra and search results.

#### 2.8.3 Construction of Wheat EST Database for searching using XTandem and Mascot

Approximately 620,000 wheat EST sequences were available through GenBank in February 2006. These sequences were assembled into contigs at the plant genome database

(www.plantgdb.org/search/misc/plantlistconstruction.php?mySpecies=Triticum%20aesti yum) which reduced the dataset to approximately 200,000 contig sequences. These contig sequences were downloaded as a single file in FASTA format. A biopython program was written to translate these EST-contigs in all six reading frames and also to give each translated product a unique name while maintaining the FASTA format (see Appendix-3). These translated EST-contigs were assembled into a single file which was searchable using Mascot and XTandem (thegpm,

http://137.111.103.157/tandem/thegpm\_tandem.html) (Dong et al., 2004; Dong et al., 2005).

### 2.8.4 Wheat EST alignments

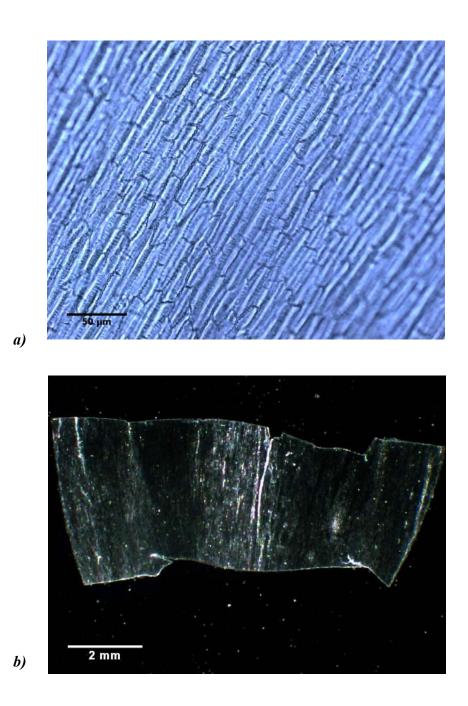
Wheat EST sequences were aligned using Clustal W (1.81) to create multiple sequence alignments. Phylogenetic relationships were created using PHYLIP and alignments were viewed using SEAVIEW (Galtier et al., 1996) software. NJplot was used to draw an unrooted phylogentic tree diagram from the PHYLIP output (Perriere and Gouy, 1996).

# 3. Results

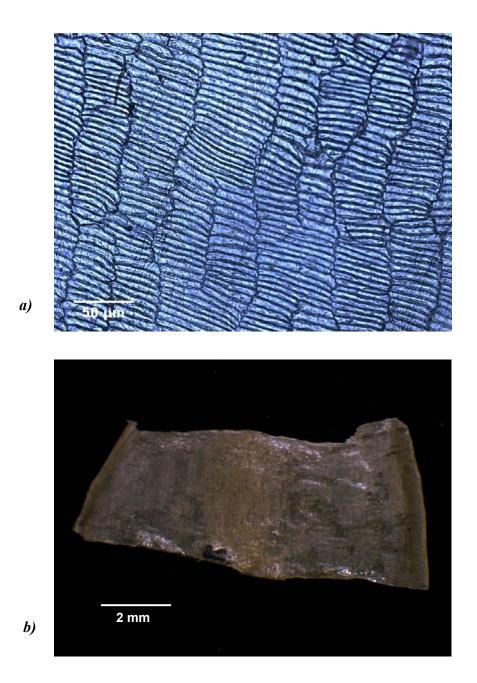
# 3.1 Collection of tissue fractions and microscopy

Bran was separated into three separate tissue fractions based on ease of separation. Firstly, the outer fraction was removed, which comprised the epidermis and hypodermis (Fig. 3.1). This fraction was the easiest to remove using the 'freeze-thaw' technique. Next, the inner fraction was scraped off and collected in water. This fraction contained aleurone cells and possibly some connecting tissue between the endosperm and the nucellar tissue (Fig. 3.4). Lastly, the remaining tissue after removing the inner fraction was the intermediate fraction. This fraction is comprised of nucellar tissue, seed coat (testa), tube cells and cross cells, together forming a thin compact multilayer tissue fraction (Fig. 3.2).

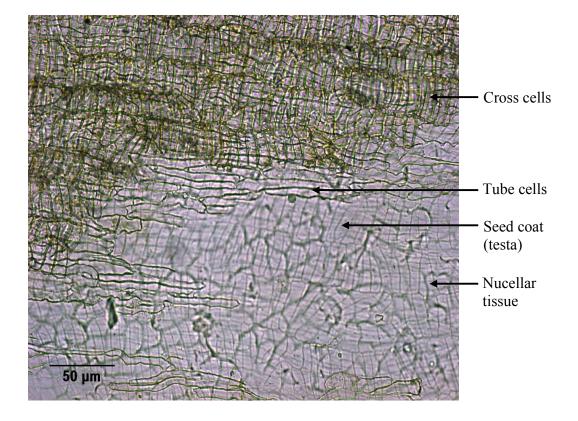
All of the tissue fractions were checked for purity using light microscopy. Photomicrographs showed the distinctive cell patterns of the outer fraction epidermis and hypodermis (Fig. 3.1) and the intermediate fraction cross cells (Fig. 3.2). The individual tissue layers of the intermediate fraction could be seen (Fig. 3.3). The majority of the inner-fraction cells (aleurone cells) were still relatively intact, showing cell walls and cell contents (Fig. 3.4).



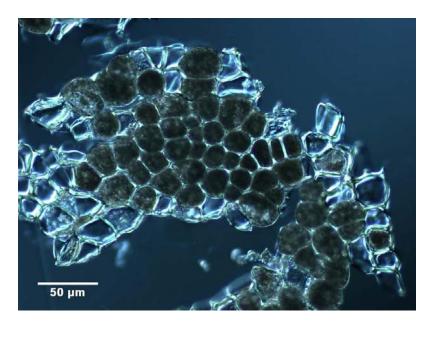
**Figure 3.1** Mechanically stripped-off outer fraction (epidermis and hypodermis): a) viewed using a light microscope, b) whole outer fraction viewed under a dissecting microscope with variable zoom.



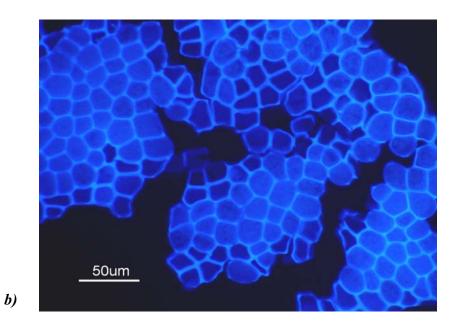
**Figure 3.2** Mechanically stripped-off intermediate fraction (cross cells, tube cells, testa, and nucellar tissue): a) viewed using a light microscope, b) whole intermediate fraction viewed under a dissecting microscope with variable zoom.



**Figure 3.3** Photomicrograph of the intermediate fraction (cross cells, tube cells, testa, and nucellar tissue), viewed using a light microscope.



a)



**Figure 3.4** Mechanically stripped-off inner fraction (aleurone cells): a) viewed under dark field microscopy, b) viewed under fluorescence during exitation with UV (395 nm).

# 3.2 Extraction and quantification of proteins from bran tissue fractions and supernatant from imbibed grain

Proteins extracted from the intermediate and outer tissue fractions (see Table 3.1) represented less than 10% yield predicted from the data of Spurway (1998). In contrast, the inner fraction showed a much higher concentration of protein using the same extraction method used by Wang et al. (2003). The amount of protein extracted was 70% of the expected protein content in the inner fraction shown in Table 3.1; thus, there is a large difference in the percentage of proteins extracted between the intermediate and outer tissue fractions and the inner fraction.

To account for any loss of water-soluble protein during tissue collection, the supernatant was collected from the imbibed grain and also from the imbibed isolated outer tissue fraction. The amount of protein that leached out from the whole grain into the supernatant was higher (1.3  $\mu$ g.grain<sup>-1</sup>) than the amount protein that leached out from the isolated outer fraction (0.5  $\mu$ g.grain<sup>-1</sup> equivalent) (see Table 3.1). This difference in protein amounts can be seen in the 1D gels where a number of protein bands seen in the whole-grain supernatant were missing from extracts of isolated outer fraction (Fig. 3.11). The extra protein bands seen in the whole-grain supernatant are likely to be due to proteins leaching out from inner bran tissue fractions, or more likely from the germ.

It was estimated that 0.6 mg of protein per gram of outer tissue fraction had leached out into the supernatant. This is a relatively large amount of protein per gram of tissue material when compared to the 0.4 mg.gram<sup>-1</sup> of protein detected in the water-insoluble

component of the outer tissue fraction (see Table 3.1). This probably indicates that supernatant proteins were highly water-soluble, or that the protein extraction method did not accurately reflect actual protein extracted from these fractions.

**Table 3.1** Estimated protein content in tissue fractions (Spurway, 1998): a) protein extracted from bran tissue fractions using the method from Wang et al., (2003) (see Appendix A-1.1); b) water-soluble protein extracted from supernatant of imbibed whole grain and isolated outer fraction (see Appendix A-1.2)

Fraction and protein extraction method	Protein content mg.g <sup>-1</sup>	Protein content µg.grain <sup>-1</sup>	Reference
Reference			
Outer and intermediate (pericarp and testa)	56.0	NA	Spurway, 1998
Inner (aleurone)	221.0	NA	Spurway, 1998
Protein extraction by Wang et al., (2003) method			
a) Outer	0.4	NA	This study
Intermediate	3.6	NA	This study
Inner (aleurone)	156.0	NA	This study
Water extraction			
b) Isolated outer fraction (pericarp)	0.6*	0.5*	This study
Whole grain	NA	1.3*	This study

NA – not applicable

\* Water-soluble proteins

# 3.3 Protein spot distribution in 1D and 2D gels

#### 3.3.1 Water-soluble proteins

Two methods were used to discriminate the location of water-soluble proteins that leached out from the grain during imbibition. The first was imbibing the whole grain in water and the second was to imbibe isolated outer tissue fraction alone. The proteins were collected from each method and were separated by SDS-PAGE. The protein bands that were present in both gels were assumed to originate from the outer fraction alone. Other proteins that were not present in the isolated outer fraction gel were assumed to have either leached out from the inner bran tissue fractions or the germ (Fig. 3.11).

#### 3.3.2 Outer fraction

The results of 2D electrophoresis analysis of the outer fraction using both the 4 to 7 pH (Fig. 3.5) and the 6 to 11 pH (Fig. 3.6) range gels showed very few protein spots. The majority of proteins were in the acidic pH range of 4 to 5, and around a molecular weight of 25 kDa (Fig. 3.5). The only major proteins in the alkaline range were around pH 10, with a low molecular weight of around 5 kDa (Fig. 3.6). There were also very faint protein spots around pH 7 with molecular weights around 20 kDa.

#### 3.3.3 Intermediate fraction

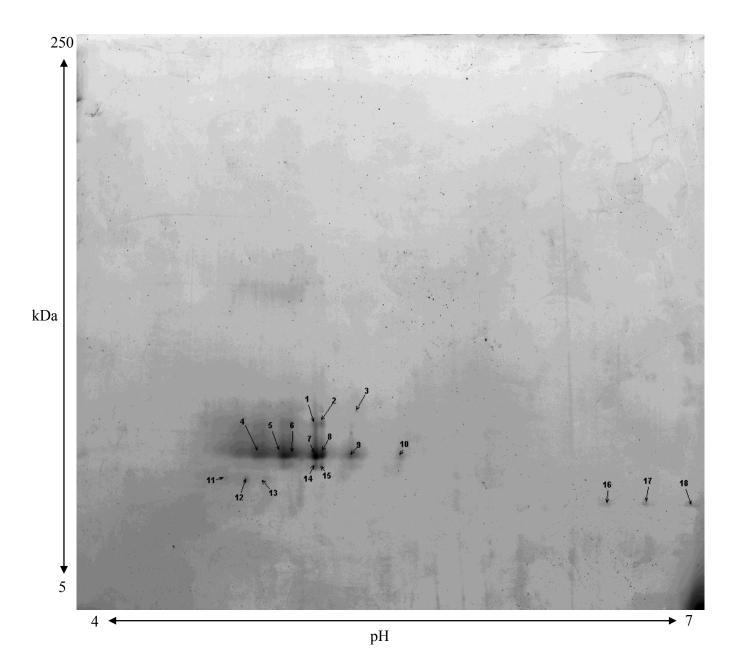
In contrast to the outer fraction, almost all of the protein spots in the 2D gels of the intermediate fraction (Figs. 3.7 and 3.8) were in the alkaline pH range 6 to 11 with molecular weights ranging from 20 to 40 kDa (Fig. 3.8). There were only a few protein spots in the 4 to 7 pH range, mainly between 5 and 10 kDa (Fig. 3.7).

#### 3.3.4 Inner fraction

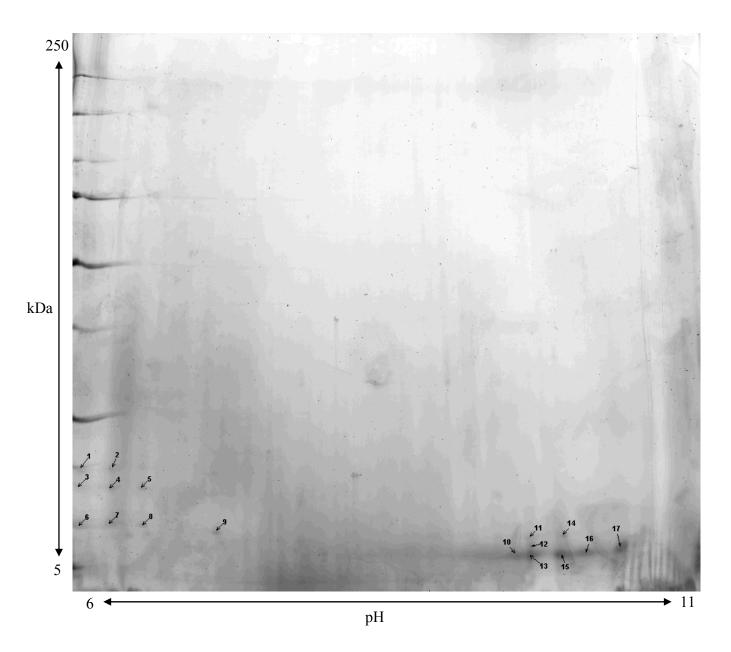
Two-dimensional electrophoresis analysis of the inner fraction, show a clear pattern of protein distribution across the pH range from 4 to 11 (Figs. 3.9 and 3.10). In the acidic pH range between pH 4 and pH 7, the majority of protein spots were of low molecular weight (Fig. 3.9). In the alkaline pH range between pH 6 and pH 11, the majority of proteins were of high molecular weight (Fig. 3.10). As expected based on the quantity of protein extracted, the 2D gels of the inner tissue fraction yielded more proteins than in the outer and intermediate fractions.

The purity of the inner tissue fraction was a concern due to the difficulty of separating it from the endosperm with minimal endosperm protein contamination. The distribution patterns in these gels (Figs. 3.9 and 3.10) differed significantly from patterns in endosperm 2D gels across a similar pH range (Skylas et al., 2000). Proteins in the 2D gels of endosperm were of relatively high molecular weight in the acidic pH range and clustered around pH 5, while in the alkaline pH range the majority of proteins were of medium molecular weight and located around pH 8 and pH 10. Comparison of displays between the endosperm and inner fraction 2D gels showed major differences. Thus the inner tissue fraction collection technique was successful as there were no obvious similarities between the 'endosperm' and 'inner fraction' 2D gels.

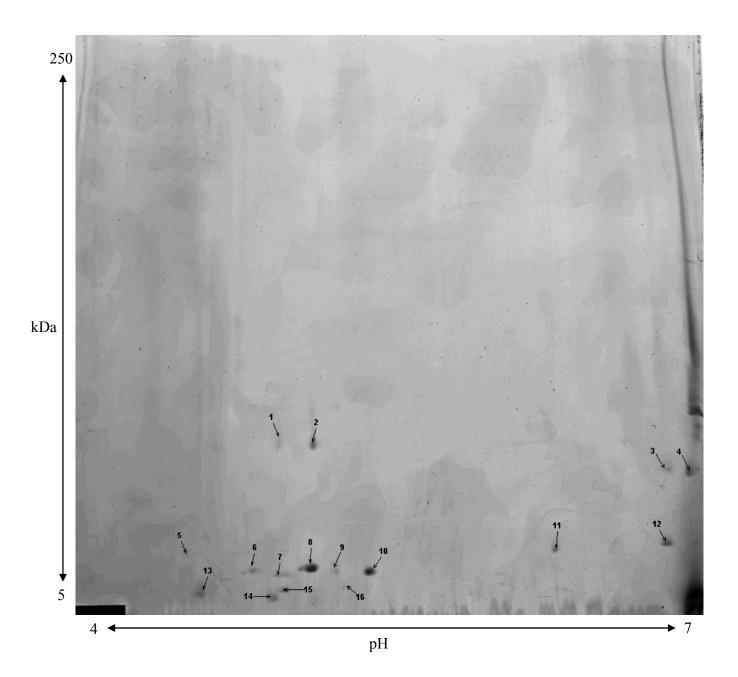
51



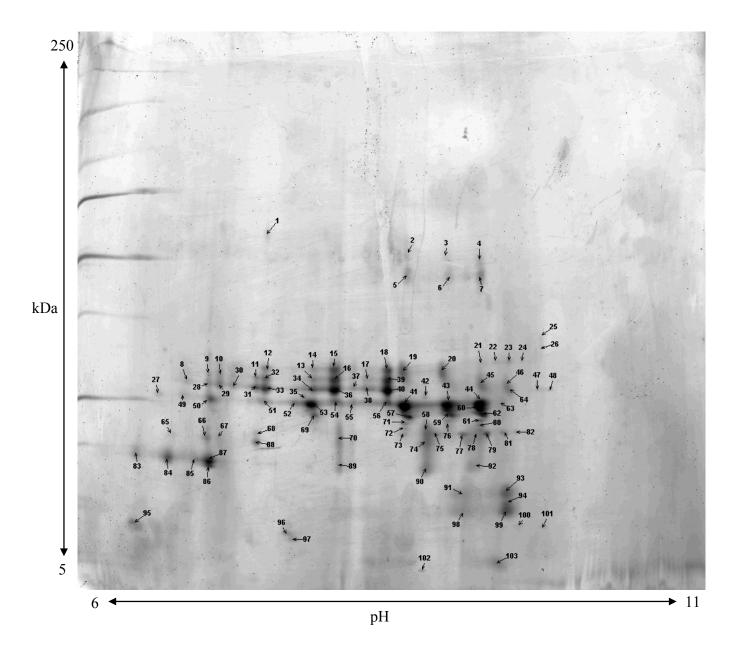
**Figure 3.5** Two-dimensional gel of the outer tissue fraction (epidermis and hypodermis) between the pH range 4 to 7 stained with Deep Purple. Protein spot numbers refer to Appendix-2, Table A-2.1.



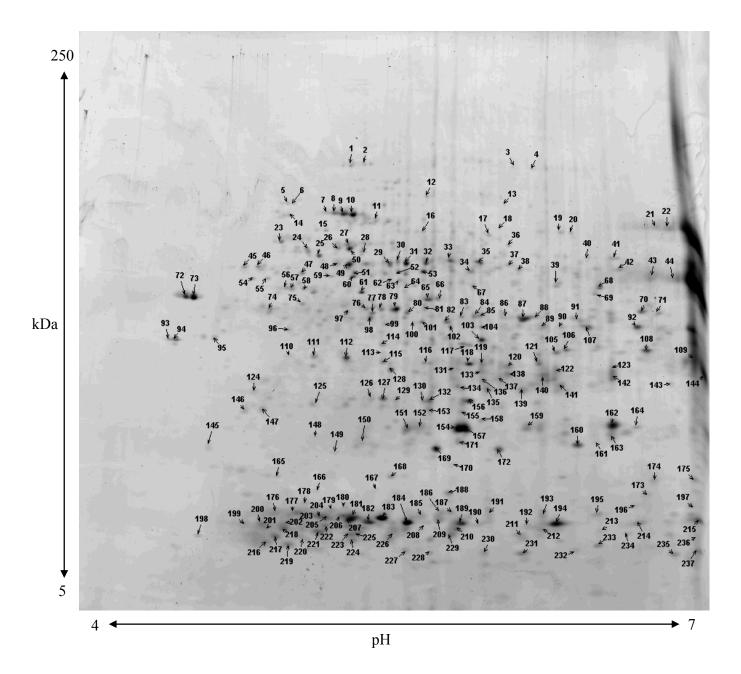
**Figure 3.6** Two-dimensional gel of the outer tissue fraction (epidermis and hypodermis) between the pH range 6 to 11 stained with Deep Purple. Protein spot numbers refer to Appendix-2, Table A-2.2.



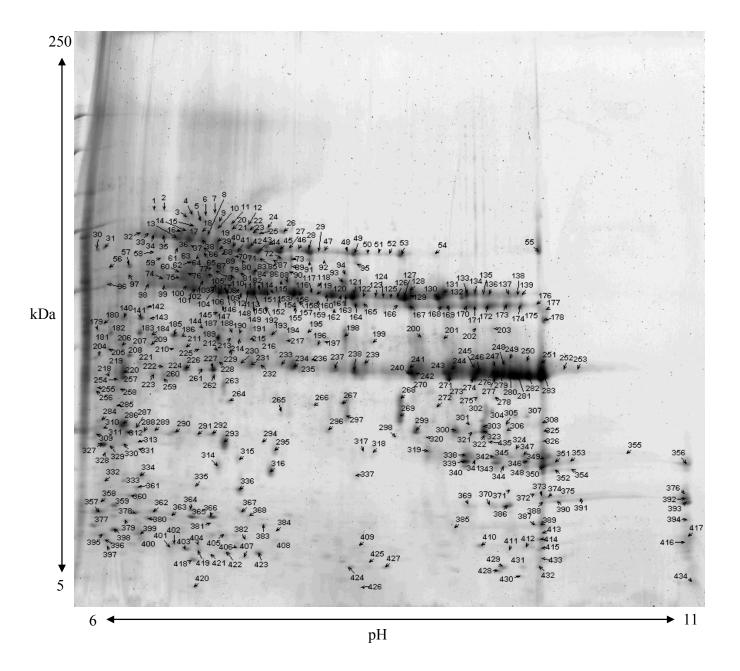
**Figure 3.7** Two-dimensional gel of the intermediate tissue fraction (nucellar, testa, tube cells and cross cells) between the pH range 4 to 7 stained with Deep Purple. Protein spot numbers refer to Appendix-2, Table A-2.3.



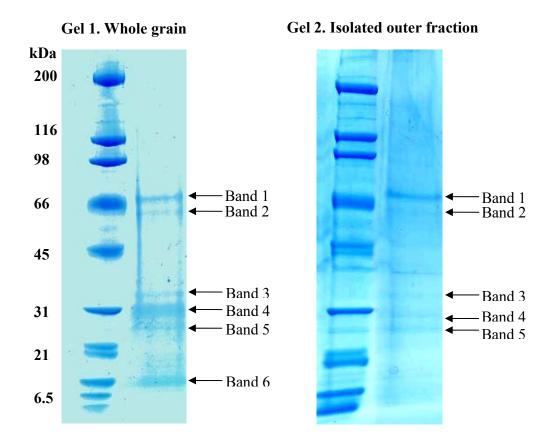
**Figure 3.8** Two-dimensional gel of the intermediate tissue fraction (nucellar, testa, tube cells and cross cells) between the pH range 6 to 11 stained with Deep Purple. Protein spot numbers refer to Appendix-2, Table A-2.4.



**Figure 3.9** Two-dimensional gel of the inner tissue fraction (aleurone cells) between the pH range 4 to 7 stained with Deep Purple. Protein spot numbers refer to Appendix-2, Table A-2.5.



**Figure 3.10** Two-dimensional gel of the inner tissue fraction (aleurone cells) between the pH range 6 to 11 stained with Deep Purple. Protein spot numbers refer to Appendix-2, Table A-2.6.



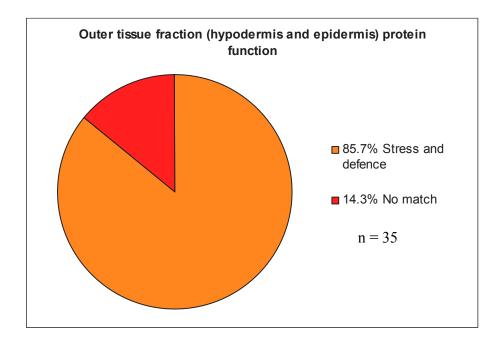
**Figure 3.11** Gel 1, proteins extracted from supernatant of whole grains; Gel 2, proteins extracted from supernatant of the outer fraction only. Gels were stained with Coomassie Blue G-250. Protein band numbers refer to Appendix-2, Table A-2.7.

# 3.4 Proteins identified in bran tissue fractions and supernatant from imbibed grain

Criteria for all the protein identifications (matches) were based on high scores (generally above 70) from Mascot searches, however low Mascot scores below 70 were still considered based on good peptide-to-protein coverage and low log (e) values (below -1) from 'thegpm' searches together with a combination of the number of peptide matches and percent peptide coverage of the protein. Wheat EST's identified were searched against NCBI BLAST and matched to proteins in GenBank with e-values less than 0.0001. All of the protein summary tables refer to the complete protein identification tables in Appendix-2. The complete protein identification tables in Appendix-2 show further information such as search scores as described above, the pI and MW of each protein and its position on the 2D gels, the protein homology to species, matching peptides and percent coverage of the identified protein and accession numbers for each protein. The 'protein summary tables' show a summary of all the identified proteins in Appendix-2 grouped based on their major function and with a more specific sub-class function together with spot number references to 1D and 2D gels.

#### 3.4.1 Proteins identified in the outer fraction

A summary of the proteins identified in the outer fraction from Appendix-2, Tables A-2.1 and A-2.2 are shown in Table 3.2. In total 35 spots were selected for identification. The major proteins identified in this fraction were oxidative-stress and defence-related proteins (30 spots or 85.7%) (Fig. 3.12). All of the identified proteins were matched to wheat species with two spots matching to wheat EST's. Of these, there were no matches to five spots. The mass spectra data of the unmatched proteins were poor, with very few peptide masses and MS/MS ion peaks together with low peak intensities. The poor spectral data could be a result of low amount of protein as the spots excised for analysis were very faint.



**Figure 3.12** Functional distribution of protein spots on the 2D gel of the outer tissue fraction.

Defence proteins	Spot number*	Sub-class
Oxalate oxidase precursor	1a, 1b, 2a, 2b, 3a, 4a, 5a, 6a, 7a, 7b, 8a, 9a, 10a, 11a, 12a, 13a, 14a, 15a, 16a, 17a, 18a	Cupin domain-containing proteins
Oxalate oxidase GF-3.8 precursor (EC 1.2.3.4) (Germin GF-3.8)	4b	
Oxalate oxidase GF-2.8 precursor (EC 1.2.3.4) (Germin GF-2.8)	3b, 5b, 6b, 8b, 9b	
PLAT (Polycystin-1, Lipoxygenase, Alpha-Toxin) domain or LH2 (Lipoxygenase homology 2) domain	11b	Oxidative stress and defence
Type 1 non-specific lipid transfer protein precursor	12b, 17b	Transport and defence
No match	10b, 13b, 14b, 15b, 16b	

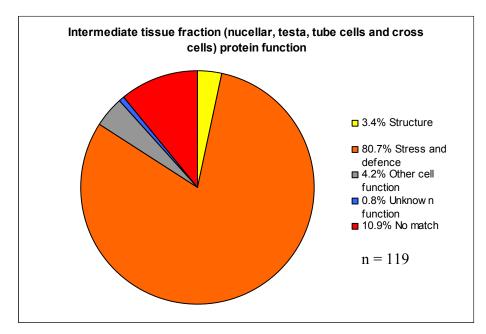
 Table 3.2
 Summary of proteins identified in the outer tissue fraction from Appendix-2 (see Tables A-2.1 and A-2.2)

\* Spot numbers with suffix 'a' and 'b' correspond to pH range 4-7 and 6-11 gels respectively displayed in Figures 3.5 and 3.6

#### 3.4.2 Proteins identified in the intermediate fraction

A summary of the proteins identified in the intermediate fraction from Appendix-2, Tables A-2.3 and A-2.4 is shown in Table 3.3. There were 119 spots selected for identification. The major proteins identified in this fraction are also oxidative-stress and defence-related proteins (106 spots or 89.1%; Fig. 3.13). The identified proteins (84 spots or 79.2%) were matched to wheat species with 22 (20.8%) of these matched to wheat EST's. Proteins that were not identified to wheat species were identified to cereal or plant homologues (22 spots or 20.8%). Of the selected spots, 13 were unable to be identified. The mass spectral data of the unmatched proteins were poor, with very few peptide masses and few MS/MS ion peaks together with low peak intensities. Oxalate oxidase (OXO) was the only protein identified in the intermediate fraction that was also identified in the outer fraction and supernatant. Thus OXO may be located in the intermediate fraction or it might have leached across from the outer fraction into the intermediate fraction during grain imbibition.

The major defence-related proteins xylanase inhibitor, chitinase and endochitinase,  $\alpha$ amylase/subtilisin inhibitor, wheatwin1, thaumatin like protein (TL) and benzothiadiazole (BTH)- clone of a wheat chemically-induced protein (cWCI-5) were all identified in this fraction (see Table 3.3).



**Figure 3.13** Functional distribution of protein spots on the 2D gel of the intermediate tissue fraction.

(see Tables A-2.3 and A-2.4)		
Defence proteins	Spot number*	Sub-class
26 kDa endochitinase 1 precursor	20b, 41b, 43b, 50b, 52b,	Chitinases
	61b, 62b, 76b, 78b, 80b,	
	91b	
26 kDa endochitinase 2 precursor (EC 3.2.1.14) (CHI-	9b, 19b, 26b	
26)		
Basic endochitinase A precursor (EC 3.2.1.14) (Rye	25b	
seed chitinase-a) (RSC-a)		
Chitinase a	63b	
Class II chitinase (EC 3.2.1.14)	27b, 28b, 29b, 34b, 42b,	
	44b, 45b, 48b, 49b, 56b, 59b, 72b, 73b, 79b, 86b,	
	100b, 101b	
	1000, 1010	
Chain B, Crystal Structure of Chitinase At 1.91a	5b, 55b	
Resolution	50, 550	
Xylanase inhibitor protein 1 precursor (Class III	3b, 4b, 8b, 10b, 13b, 14b,	Xylanase inhibitors
chitinase homolog) (XIP-I protein)	15b, 18b, 22b, 51b, 54b,	
	66b, 67b	
Xylanase inhibitor protein I	16b, 30b, 32b, 37b, 38b,	
	39b, 40b, 71b, 102b	
Vulanasa inhibitar VID III	(0h	
Xylanase inhibitor XIP-III Chain A. Cristal Structure of Family 11 Videnase in	69b	
Chain A, Crystal Structure of Family 11 Xylanase in Complex with Inhibitor (Xip-I)	12b, 65b, 85b, 87b, 89b, 90b, 92b, 94b, 103b	
Chain A, Crystal Structure Of Xylanase Inhibitor	36b, 46b, 53b, 57b	
Protein (Xip-I)	500, 400, 550, 570	
Xylanase inhibitor precursor (Xylanase inhibitor	64b	
TAXI-I)		
Chain B, Crystal Structure Of The Triticum aestivum	6a, 8a	
Xylanase Inhibitor-I In Complex With Aspergillus		
Niger Xylanase-I		
Xylanase inhibitor	7a, 10a	
Alaba ana lara inhibitan	2- 4- 22h	Alaba and the interview
Alpha amylase inhibitor Endogenous alpha-amylase/subtilisin inhibitor	3a,4a, 33b	Alpha amylase inhibitors
(WASI)	23b, 35b, 60b, 68b, 83b, 84b	
נעהיין)	UTU	
Benzothiadiazole-induced protein (clone	75b, 82b	Other defence proteins
WCI-5)		
Oxalate oxidase precursor	1a, 2a	
Pathogenesis-related protein 4 (Fragment)	5a, 12a, 95b, 96b, 97b	
Thaumatin-like protein	17b	
Wheatwin1	11a	

# **Table 3.3** Summary of identified proteins in the intermediate tissue fraction from Appendix-2(see Tables A-2.3 and A-2.4)

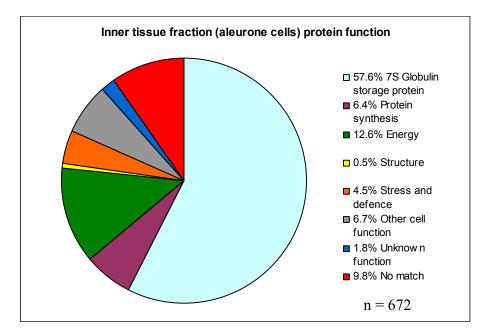
Other	Spot number*	Sub-class
Plant Basic Secretory Protein	31b	Transport
Secretory protein	11b, 77b, 88b	
Actin (065316)	2b, 6b, 7b, 21b	Structural proteins
Uncoupling protein	13a	Other
Unknown	81b	
No match	1b, 9a, 14a, 15a, 16a,	24b,
	47b, 58b, 70b, 74b, 9	3b,
	98b, 99b	

Table 3.3 Summary of identified proteins in the intermediate tissue fraction (continued)

\* Spot numbers with suffix 'a' and 'b' correspond to pH range 4-7 and 6-11 gels respectively displayed in Figures 3.7 and 3.8

#### 3.4.3 Proteins identified in the inner fraction (Aleurone cells)

A summary of the proteins identified in the inner fraction from Appendix-2, Tables A-2.5 and A-2.6 is shown in Table 3.4. There were 672 spots selected for identification. The major proteins identified in this fraction were 7S globulin storage proteins (387 spots or 57.6%) and cell function proteins (128 spots or 19%) involved in protein synthesis and carbohydrate metabolism (Fig. 3.14). Of the identified proteins, 480 spots (79.2%) were matched to wheat species with 223 spots (36.8%) of these matched to wheat EST's. Proteins that were not identified to wheat species were identified to cereal or plant homologues (126 spots or 20.8%). Of the selected spots, 66 were unable to be identified. The mass spectral data of the unmatched proteins were good, showing many MS peaks and MS/MS ion peaks together with high peak intensities similar to spectral data of matched to proteins.



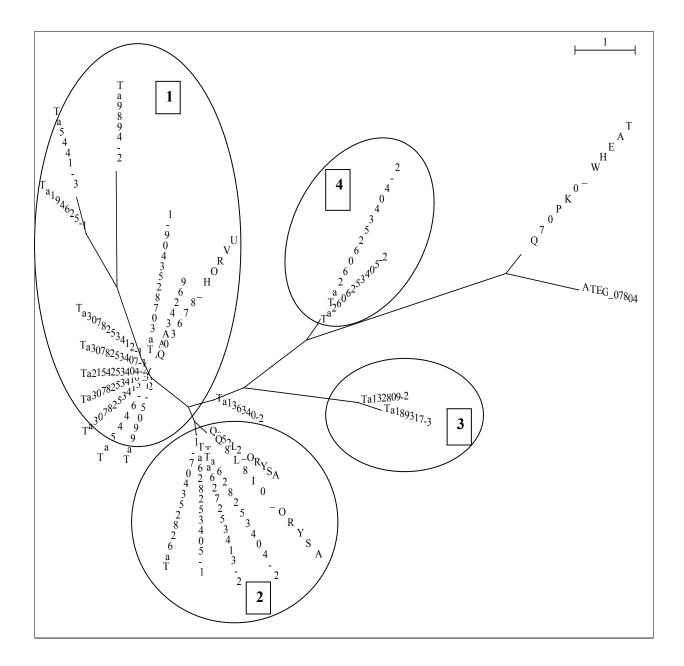
**Figure 3.14** Functional distribution of protein spots on the 2D gel of the inner tissue fraction.

#### 3.4.4 Further analysis of major protein classes in inner fraction

#### 3.4.4.1 Cupin domain-containing proteins

The majority of proteins identified from the inner tissue fraction 2D gel images (Figs. 3.9 and 3.10) were found to be cupin domain-containing proteins from the cupin super family (387 spots or 58%). Mascot search against the cereal database identified 207 spots (53%) that were wheat 7S globulin storage proteins, four spots (1%) matched to rice globulin-like protein, one spot (0.3%) matched to a rice putative globulin and finally one spot (0.3%) matched to a barley embryo globulin. Wheat EST sequences identified were searched against NCBI BLAST; 163 spots (42%) were matched to the wheat 7S globulin storage protein and 11 spots (3%) were matched to a cupin family expressed protein (Table 3.4).

Alignments were performed of all the translated wheat EST amino acid sequences that matched to cupin family proteins, together with amino acid sequences from wheat 7S globulin storage protein, rice globulin-like protein, barley-embryo globulin protein and also cupin family oxalate-metabolizing proteins, oxalate oxidase (identified in the outer tissue fraction) and an oxalate decarboxylase sequence (Appendix A-3). An unrooted phylogenetic tree diagram was drawn showing the relationship of the alignments (Fig. 3.15). There were three main clusters of wheat EST sequence orthologous to three different groups of cupin family proteins. Cluster (1) represents all orthologs of the wheat 7S globulin storage protein together with the barley embryo globulin protein. Cluster (2) represents all orthologs of the rice globulin-like and putative globulin proteins. Finally, clusters (3 and 4) represent orthologs of the oxalate-metabolizing proteins oxalate oxidase and oxalate decarboxylase (Table 3.4).



**Figure 3.15** Unrooted phylogenetic tree showing clusters of wheat EST sequence alignments to all cupin containing protein showing closest ortholog.

Cluster	Protein	Wheat EST orthologues
1	7S globulin storage protein (Q7DMU0_WHEAT)	Ta3078253413-3
	Embryo globulin (Q03678_HORVU)	Ta136340-2
		Ta194625-1
		Ta2154253404-2
		Ta2606253405-2
		Ta3078253409-1
		Ta3078253410-1
		Ta3078253411-3
		Ta3078253412-1
		Ta3078253413-3
		Ta3079253404-3
		Ta5441-3
		Ta5446-1
		Ta628253407-1
		Ta9894-2
		Ta9905-2
2	Globulin-like protein (Q8L8I0 ORYSA)	Ta3078253407-1
	Putative globulin (Q852L2 ORYSA)	Ta628253405-1
		Ta627253413-2
		Ta628253404-2
3 and 4	Oxalate oxidase precursor (Q70PK0 WHEAT)	Ta132809-2
	Oxalate decarboxylase (gi:114190366)	Ta189317-3
		Ta2606253405-2
		Ta2606253404-2

 Table 3.4
 Clusters of wheat EST sequence alignments to cupin containing protein showing closest ortholog

#### 3.4.4.2 Cell function proteins

The other major proteins identified in the inner fraction 2D gels are involved in protein synthesis, folding and stability, carbohydrate metabolism (128 spots or 19%) as mentioned previously and the stress-and defence-related proteins (31 spots or 4.5%) that were similarly identified in the intermediate tissue fraction (Fig. 3.14).

# **Table 3.5** Summary of identified proteins in the inner tissue fraction from Appendix-2(see Tables A-2.5 and A-2.6)

Storage	Spot number*	Sub-class
7S Globulin storage protein	(370 spots)	Cupin domain containing storage
		protein
Cupin family protein, expressed	152a, 154a, 157a, 171a,	
	256b, 286b, 293b, 295b,	
	314b, 316b, 337b	
Embryo globulin	410b	
Putative globulin	73b	
Globulin-like protein	93b, 105b, 109b, 215b	
-		
Protein synthesis		2
Elongation factor 1-alpha (EF-1-alpha)	137b, 138b, 139b, 431b	Gene expression
Elongation factor 1-alpha, putative, expressed	432b	
Elongation factor Tu	76a	
Putative ribophorin I homologue	198a	
R2R3MYB-domain protein (Fragment)	381b	
RNA recognition motif family protein, expressed	193b	
6.9 kDa class I heat shock protein (Low molecular	192a, 209a, 380b	Protein synthesis and folding
veight heat shock protein)		
30S ribosomal protein S2	272b	
10S subunit ribosomal protein	387b	
50S ribosomal protein L31	418b	
50 kDa chaperonin	25a	
Chaperonin CPN60-1, mitochondrial precursor HSP60-1	27a	
Heat shock protein 16.9B	186a, 359b	
Heat shock protein 17.9.	195a	
Heat shock protein HSP26	148a, 150a, 170a, 167a	
HSP70	7a, 8a, 9a, 10a, 84a, 85a,	
	86a, 87a, 124a	
Putative 40S ribosomal protein S3	308b	
Putative 60S ribosomal protein L12	386b	
Putative 60S ribosomal protein L36	111a	
Putative alpha 1 subunit of 20S proteasome	164a, 287b	

<b>Table 3.5</b> Summary of identified proteins in the inner tissue fraction (continued)	

Protein synthesis	Spot number*	Sub-class
Small heat shock protein Hsp23.5 precursor	166a	
Small heat shock protein Hsp23.6 precursor	149a	
Protein disulfide isomerase (EC 5.3.4.1) (Fragment)	23a	
Histone H3	434b	DNA binding
Enzymes		
Citrate synthase, eukaryotic	142b	Citric acid cycle
Malate dehydrogenase	204b	
Putative malate dehydrogenase	105a, 208b	
Cytosolic malate dehydrogenase (EC 1.1.1.37) (Fragment)	103a	
Aconitate hydratase, cytoplasmic, putative, expressed	4a	
NADP-specific isocitrate dehydrogenase (EC 1.1.1.42)	88a	
Putative dihydrolipoamide dehydrogenase	40a, 41a, 57b	
Triosephosphate-isomerase	151a	
Aldose reductase-related protein (EC 1.1.1.21)	144a, 226b	Glycolysis and gluconeogenesis
Cytoplasmic aldolase	179b	
Cytosolic glyceraldehyde-3-phosphate dehydrogenase	89a, 90a, 91a, 92a, 107a,	
GAPDH (Fragment)	121a, 189b, 207b, 212b,	
	216b	
Enolase (EC 4.2.1.11)	29a, 31a, 32a, 180b	
Glyceraldehyde-3-phosphate dehydrogenase, cytosolic (EC 1.2.1.12)	145b, 182b, 183b, 185b	
2,3-bisphosphoglycerate-independent phosphoglycerate mutase	16a	
Putative 2,3-bisphosphoglycerate-independent phosphoglycerate mutase	17a, 36a	
Putative aldose reductase	222b, 227b, 261b, 262b, 288b, 290b, 291b	
Phosphoglycerate kinase, cytosolic (EC 2.7.2.3)	65a, 82a, 101a	
Putative glyceraldehyde-3-phosphate dehydrogenase	211b	

nzymes	Spot number*	Sub-class
lucose and ribitol dehydrogenase homolog - barley	123a, 142a, 219b, 220b,	
	256b, 257b, 260b, 263b	
hosphoglucomutase (EC 5.4.2.2) (Fragment)	18a	
ruvate orthophosphate dikinase (Fragment)	2a	
tative S-formylglutathione hydrolase	218b	Oxidative stress
lutathione transferase (EC 2.5.1.18)	160a, 161a, 327b	
tative cytosolic 6-phosphogluconate dehydrogenase	67a	Pentose phosphate pathway
tative fructose 1-,6-bisphosphate aldolase	184b, 206b	
ragment)		
Pglucose-1-phosphate uridylyltransferase (EC	51a	Carbohydrate metabolism
7.7.9)		
tosolic NADP malic enzyme	19a, 20a, 29a, 30b, 31b	C4 pathway
spartate aminotransferase precursor (EC 2.6.1.1)	181b	C+ paulway
lanine aminotransferase	96b	
P synthase beta subunit	48a, 49a	ATP synthesis
p1 protein	48a, 49a 33a, 35a, 37a	ATF Synthesis
cleoside diphosphate kinase (EC 2.7.4.6)	357b	
ragment)		
cleoside diphosphate kinase 1, putative, expressed	233a	
tative nucleoside diphosphate kinase	377b, 383b, 396b, 398b	
rmate dehydrogenase (Fragment)	70a, 141b	C1 Metabolism
prmate dehydrogenase, mitochondrial precursor (EC	140b	

Table 3.5	Summary	of identified	proteins	in the	inner	tissue	fraction	(continued)	

Enzymes	Spot number*	Sub-class
Subtilisin-like protease	66a	Protease
Aspartic proteinase	146a, 147a	
12-oxo-phytodienoic acid reductase	61a	Other
Glutamine synthetase isoform GS1c	99a	
Manganese superoxide dismutase (EC 1.15.1.1)	329b, 330b	
O-methyltransferase	205b	
Putative beta-N-acetylhexosaminidase (Fragment)	45a	
Putative proteasome subunit alpha type 3	158a	
Putative serine/threonine protein kinase	264b	
Transposase	161b, 236a, 433b	
Xylose isomerase (EC 5.3.1.5)	30a	
Putative glyoxalase I	128a	
Glyoxalase family protein, expressed	227a, 232a	
Putative rubisco subunit binding-protein alpha	24a	
subunit precursor		
Structural proteins		
Actin and related proteins	302b, 303b, 304b, 339b	Cytoskeletal
Defence and stress related proteins	100 2001 2111 2121	<u>0</u> ,
1-Cys-peroxiredoxine	122a, 309b, 311b, 312b	Stress
Stress responsive protein	221b	
Stress-inducible membrane pore protein	364b	
Translationally controlled tumor protein	145a	
26 kDa endochitinase 1 precursor (EC 3.2.1.14)	301b, 306b, 435b	Defence
Basic endochitinase C precursor (EC 3.2.1.14) (Rye seed chitinase-c)	323b	
Class II chitinase (EC 3.2.1.14)	273b, 296b, 299b, 300b, 305b, 322b	
Disease resistance protein (Fragment)	106a, 108a	
Disease-resistent-related protein	416b	
Endogenous alpha-amylase/subtilisin inhibitor	175a, 315b, 335b, 336b	
(WASI)		
PR-4 (Fragment)	235a	
Xylanase inhibitor (Fragment)	198b	
Xylanase inhibitor precursor (Xylanase inhibitor TAXI-I)	201b	
Xylanase inhibitor protein 1 precursor (Class III chitinase homolog)	268b, 269b, 297b	

### **Table 3.5** Summary of identified proteins in the inner tissue fraction (continued)

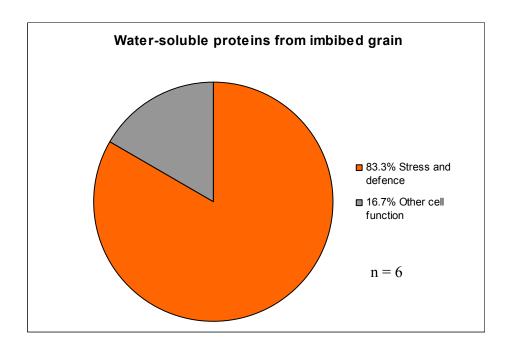
Other	Spot number*	Sub-class
ABA inducible protein	230a	Programmed cell death (PCD)
Putative cell death associated protein	90b	
Voltage dependent anion channel (VDAC)	278b	
(Fragment)		
Amylogenin	104a	Other cell function
Annexin	59a, 294a	
Caleosin 1	223b, 284b	
Cyclophilin A-2 (EC 5.2.1.8) (Cyclophilin A)	13a	
DNA-binding protein (Fragment)	110a	
Em protein H5	39a	
Globosa	42a	
Lipoprotein-like	267b	
MLo protein	50a	
OJ991113_30.23 protein	197b	
Oleosin (Fragment)	389b	
OSJNBa0027P08.9	191a, 384b	
OSJNba0093F12.16 protein	370b	
P0481E12.28	320b	
Prohibitin protein Wph	313b, 369b	
Reversibly glycosylated polypeptide	100b	
Secretory protein	342b, 345b	
Squamosa promoter binding protein 2-like	292b	
Hypothetical protein OJ1124 E11.8.	405b	Unknown
Hypothetical protein OSJNBb0094P23.22	367b	
Hypothetical protein P0431E05.13	47b	
Hypothetical protein P0686H11.1 (Hypothetical protein P0605H02.41)	209b, 285b	
Unknown protein	59b, 412b, 414b	
Unnamed protein product	169a, 172a, 174a, 332b	

**Table 3.5** Summary of identified proteins in the inner tissue fraction (continued)

\* Spot numbers with suffix 'a' and 'b' correspond to pH range 4-7 and 6-11 gels respectively displayed in Figures 3.9 and 3.10

#### 3.4.5 Proteins identified in supernatant from imbibed grain

A summary of the water-soluble proteins identified in the supernatant from imbibed grain from Appendix-2, Table A-2.7 are shown in Table 3.6. There were six protein bands selected from the 1D gel for identification (Fig. 3.11). The major group of proteins identified were oxidative-stress and defence-related proteins (five bands or 83.3%); the other band was a histone (Fig. 3.16). All of the identified proteins were homologous to wheat proteins.



**Figure 3.16** Functional distribution of protein bands on the 1D gel of the water-soluble proteins

Enzyme	Band number	Sub-class
Endochitinase precursor	4	Defence
Oxalate oxidase precursor	1	Oxidative stress and defence
PSBGer3 protein	5	
Polyphenol oxidase (Fragment)	2	
Peroxidase (EC 1.11.1.7) (Fragment)	3	
Other		
Histone H4 variant TH011	6	DNA folding

#### Table 3.6 Summary of identified proteins in supernatant from imbibed grain (see Apendix-2 Table A-2.7)

#### 3.4.6 Supernatant from isolated outer fraction

To further investigate the location of the proteins identified in the supernatant from imbibed whole grain, 1D-PAGE was used to investigate proteins that leach out from the supernatant of the outer tissue fraction alone. The results showed that only polyphenol oxidase (PPO), peroxidase (POX), and oxalate oxidase (OXO) were present in the supernatant (Fig. 3.11 and Table 3.6). This suggests that PPO, POX and OXO are mainly contained in the outer tissue fraction and that the other proteins, endochitinase and histone, leach out from the inner and intermediate fractions or the germ during imbibition.

## 4. Discussion

The bran layer of the grain is essentially a barrier that protects the germ and endosperm from bacterial, fungal, viral and insect attack (Selitrennikoff, 2001; Desmond et al., 2006). It was thus not surprising to find that the majority of proteins identified in the outer and intermediate bran fractions were oxidative stress- and defence-related proteins. However, the aleurone cells, which are part of isolated bran and in our study form the major component of the 'inner fraction', are live cells and are involved in the early stages of germination. This layer should thus contain proteins related to active cellular processes as the aleurone cells are expected to satisfy their function within the germination process, which is to help breakdown the endosperm to provide energy for the growing plant (Eastwood and Laidman, 1971; Fincher, 1989). Although antifungal proteins were detected in this layer, proteins related to metabolic processes expected of living cells, such as energy metabolism and protein synthesis, were some of the more common types of proteins. Surprisingly though, the major protein identified in the inner fraction was a 7S globulin storage protein, which has not been reported as a major aleurone protein. All of these major proteins with respect to their location in each tissue fraction will be further discussed in detail later in the Discussion.

#### 4.1 Bran fractionation and properties of bran fractions

The major hurdle of this study was to collect bran tissue suitable for proteomic analysis and to separate it into distinct tissue fractions. This was overcome, allowing an efficient separation into three fractions – outer, intermediate and inner – as described earlier. The reason for separating bran into fractions was to look at the distribution of proteins across the bran tissue and to identify the location of proteins with respect to their function. For instance, if only whole bran were analysed, the results would be less informative in that the abundance of proteins in the inner fraction would have overshadowed the less abundant proteins in the outer and intermediate fractions. Furthermore, identifying the location of proteins unique to different tissue fractions is informative in that the proteins can be linked to gene expression during the early stages of bran development. The method used to remove and separate the bran into fractions has been reported (Antoine et al., 2004), however the 'freeze thaw' technique developed in this study was novel and very effective in removing the outer (epidermis and hypodermis) fraction (Antoine et al., 2004). It allowed for relatively quick and easy collection of large amounts of the outer fraction, which was required for protein extraction.

The difficulty in collecting bran and then separating it into distinct bran tissue fractions is that it is tightly bound to the endosperm and is also compact and brittle in its dry state. Achieving this separation requires the grain to be wet, in turn making the bran flexible and tougher. As a result, the grain used for the collection of tissue fractions had to be imbibed in water for two days at room temperature to allow for its removal, and also to further manually separate the bran into three distinct tissue fractions. As a result of doing this, the proteins identified in this study were necessarily representative of bran at the germination stage, especially in the inner tissue fraction which is composed of living cells. Any water-soluble proteins that may leach out during grain imbibition were collected and analysed.

Once the bran was removed from the endosperm, the manual dissection of bran into three distinct tissue fractions was essentially based on the easiest separating points of adjoining tissue. Microscopy of the dissected bran fractions showed that they were virtually free from adjoining tissue contamination (Figs. 3.1, 3.2, and 3.4). Interestingly, for the purpose of bran fractionation in milling, these points of mechanical failure might also be areas to target by using enzymes to further degrade and weaken the tissue bonds and thus improve bran fractionation (i.e., separating bran from endosperm and/or removing particular bran layers all to improve flour quality and yield) and will be discussed later in the discussion.

The analysis of proteins in the tissue fractions required a suitable protein extraction method for two-dimensional polyacrylamide gel electrophoresis (2D-PAGE). A protein extraction method developed especially for the purpose of extracting protein from olive leaf and that was suitable for 2D-PAGE was used (Wang et al., 2003). This method was effective in extracting protein from the inner fraction, however it was apparently less effective in extracting proteins from the intermediate and outer fractions.

One of the possible reasons for this may be that the proteins in the intermediate and outer tissue fractions are difficult to extract from the tissue itself because of the compacted cross-linked tissue. Protein extraction from compacted tissue fractions, such as the outer and intermediate fraction, may require a better method for grinding up tissue, to expose and release as much protein as possible into the extraction buffer.

Another possible reason for the low apparent protein levels in the bran tissue fractions could be due to the Deep Purple protein stain used to quantify proteins. The fluorescent compound in Deep Purple stain covalently binds to lysine and N-terminal residues of proteins (Coghlan et al., 2005). There is a three-to four-fold difference in the lysine content in proteins between grain components. The lysine content in the flour fraction is four times lower than the germ fraction and three times lower than the aleurone fraction (Spurway, 1998). Thus, this kind of variation in lysine content may explain the low apparent concentration of proteins in the intermediate and outer fractions as it will affect staining intensity.

Furthermore, lysine is 9.9% of the amino acid residues in bovine serum albumin (BSA). BSA serial dilutions were used to develop standard curves of stain intensity, thus underestimating quantities of lysine-poor proteins that are typical of cereal grains (Spurway, 1998; van der Meer et al., 2001).

The extracted proteins from each of the tissue fractions were separated by twodimensional electrophoresis (2-DE) and stained with Deep Purple to visualise the protein spot array (Figs. 3.5, 3.6, 3.7, 3.8. 3.9 and 3.10). The first observation from these gel images is that there was a protein array/concentration gradient going from the outer fraction toward the inner fraction, there being less protein in the outer fraction and increasing moving toward the inner fraction. Although protein concentration estimation of bran fractions has been reported by Spurway (1998), this study has shown for the first time the protein composition and distribution across the bran.

The protein spots from the 2DE gels (Figs. 3.5, 3.6, 3.7, 3.8, 3.9 and 3.10) and the protein bands from the 1D gels (Fig. 3.11) were selected and processed for identification as described in the methods. The identified proteins in each of these tissue fractions and the water-soluble proteins that leached out during grain imbibition will briefly be discussed highlighting their functions and purpose in the bran.

#### 4.2 Proteins identified in the outer fraction

As mentioned previously, the function of the bran is to act as a physical barrier to protect the nutrient-rich endosperm. The outer-most tissue fraction is thus the first line of defence in biotic stress such as fungal, microbial and insect attack on the grain. The following proteins that were identified in this fraction will be briefly discussed as to their possible roles in this protective function for wheat grain.

#### 4.2.1 Oxalate oxidase (OXO) and peroxidase (POX)

OXO is a known defence protein in plants involved in resistance to oxalate-secreting fungi (Lane et al., 1993). OXO has been reported to be commonly found in the outer surface tissues of plants (Lane et al., 1993; Lane, 2000). This study has shown that OXO is a major enzyme located in the outer bran fraction. Oxalate secreted by fungi is oxidized by OXO, thus producing  $H_2O_2$  which in turn is toxic to living cells. This phenomenon is termed the hypersensitive response (Jabs, 1999). The plant overcomes damage by the presence of peroxidase (POX) which in turn breaks down  $H_2O_2$ . POX is a stress-related enzyme, which protects against abiotic stress, mechanical damage or pathogen responses, all of which could result in the production of  $H_2O_2$  (Lane, 2000). Furthermore, the role of OXO and POX in producing  $H_2O_2$  is involved in cross linking of lignin to create a tougher impenetrable barrier in this outer layer (Gane et al., 1998; Peyron et al., 2001; Antoine et al., 2003). This is consistent with the intactness of this whole layer in our isolation method.

#### 4.2.2 Lipoxygenase (LOX)

LOX is a pathogen-induced acquired resistance (AR) protein found in plants. This protein also becomes expressed when there is mechanical damage to the plant caused by insects (Kolomiets et al., 2000). LOX has previously been reported to be mainly located in the bran and germ fractions (Rani et al., 2001). This study shows that LOX is located only in the outer fraction of the bran. LOX is a desirable protein in flour in that it causes flour bleaching and strengthening of dough in bread making (Rani et al., 2001). Moreover, its location in the outer fraction means it is unlikely to end up in white flour where these properties will be of most use commercially.

#### 4.2.3 Lipid transfer protein (LTP)

LTP protein is involved in transferring phospholipids between membranes. It is also believed to be involved in defence against fungal and bacterial pathogens (Drea et al., 2005). The LTP protein gene is expressed in the outer epidermis during bran development and its expression is greatly increased after six days post anthesis (DPA) (Selitrennikoff, 2001; Drea et al., 2005). In this study, LTP was located only in the outer fraction of the bran. It is possible that the LTP is still active in the outer fraction of the mature grain, and might contribute to pathogen defence.

#### 4.3 Proteins identified in the intermediate fraction

The next line of defence in the bran is the intermediate tissue fraction. As described in the introduction, this tissue fraction is comprised of a multilayer of compacted remnant tissues that were present during bran development (Fig. 1.4). This tissue fraction is the last barrier that protects the next tissue fraction (inner fraction) which contains live cells that are important in germination. The following proteins (discussed in more detail below) were identified in this fraction and are more specifically targeted defence type proteins than those defence proteins in the outer fraction.

#### 4.3.1 Xylanase inhibitor proteins

Fungi are common pathogens of plants. They secrete xylanase in order to break down plant cell walls for food, in turn damaging the plant (Flatman et al., 2002). Xylanase inhibitor proteins have previously been isolated from wheat (Debyser et al., 1999). This study has shown that xylanase inhibitor proteins were mostly identified in the intermediate and to a lesser extent in the inner tissue fractions. This suggests that the first point of xylanase inhibition occurs at this intermediate fraction as no xylanase inhibitor proteins were identified in the outer fraction.

#### 4.3.2 Chitinase and endochitinase

Chitinases are anti-fungal proteins found in many different plants. Their function is to break down the chitin in fungal cell walls (Fig. 4.1). Once the cell walls of the fungi are broken down, turgor pressure will cause cell lysis and death (Molano et al., 1979; Selitrennikoff, 2001; Desmond et al., 2006). This study shows that these enzymes were mainly located in the inner and intermediate tissue fractions, and not in the outer fraction. Surprisingly, chitinase was also found in the supernatant from the imbibed grain and not in the supernatant from isolated outer tissue fraction. This suggests that the chitinase possibly leaches out from the inner tissue fractions when the grain is exposed to moisture. As damp conditions are favorable to fungi, the water-soluble property of chitinase possibly allows it to diffuse out from the bran and into the fungal cell wall. These enzymes may thus prevent fungi establishing colonies on the surface of grains in damp conditions.

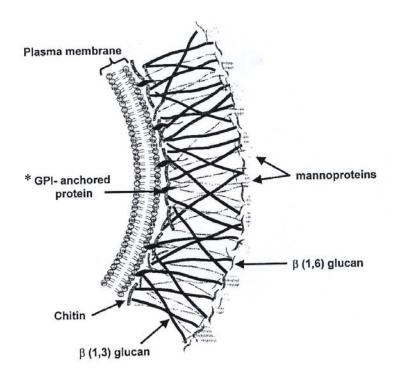


Figure 4.1 Schematic structure of a fungal cell wall (Selitrennikoff, 2001)

\* Glycophosphatidylinositol (GPI)

#### 4.3.3 α-amylase/subtilisin inhibitor proteins

 $\alpha$ -amylase and  $\alpha$ -amylase/subtilisin inhibitors were identified in the inner and intermediate tissue fractions. These proteins were also identified in the endosperm by Skylas et al. (2000), and in the germ by Mak et al. (2006b).  $\alpha$ -amylase/subtilisin inhibitor is reported to be involved in defence against microbial and insect damage. It inhibits amylases and subtilisin secreted by microbes and insects used to breakdown plant proteins and starch for food (Ohtsubo and Richardson, 1992).

#### 4.3.4 Benzothiadiazole (BTH) – clone of wheat chemically induced protein (cWCI-5)

Acquired resistance (AR) in plants is where a pathogen induces the expression of defence genes which give the plant protection from future infection. BTH induces the expression of several WCI genes, one of which is WCI-5; however, the function of this protein in defence is not yet known and it has not been reported in grain before (Gorlach et al., 1996; Schaffrath et al., 1997).

#### 4.3.5 Wheatwin1

The classing of pathogenesis-related (PR) antifungal proteins is based on their mode of action and their structural similarity (Selitrennikoff, 2001). Wheatwin1 proteins are class four pathogenesis-related (PR-4) proteins, which contain chitin-binding sites. They are active against a broad range of plant and human fungal pathogens. Their mode of action is not clearly understood, however Wheatwin1 is believed to bind to chitin in the fungal cell wall causing disruption of cell wall formation (Selitrennikoff, 2001).

#### 4.3.6 Thaumatin-like protein (TL)

Thaumatin-like protein is a class five pathogenesis-related (PR-5) antifungal protein. The mechanism of PR-5 defence proteins is not yet known, however some hypotheses are that these proteins may cause changes in fungal wall cell permeability, and some were found to have  $1,3-\beta$ -glucanase activity (Selitrennikoff, 2001).

#### 4.4 Proteins identified in the inner fraction

As described earlier, this tissue fraction contains live cells that are required for germination. It was also interesting to find that the major protein identified in this fraction was a 7S globulin storage protein not previously reported in wheat aleurone.

#### 4.4.1 7S globulin storage proteins

Storage protein is an important source of carbon and nitrogen for the developing plant during germination. 7S globulin storage proteins are a major carbon and nitrogen source in plants such as peas, nuts and legumes (Dunwell et al., 2000). This protein has previously been isolated from wheat, however its localisation has not yet been reported (Robert et al., 1985). Interestingly, the majority of protein spots in the inner fraction (around 58%) were 7S globulin storage proteins. However, this type of 7S globulin storage protein is uniquely different to the major endosperm storage proteins (gliadins) identified in the work done by Skylas et al. (2000). Since the inner fraction is only a small percentage of the whole grain, it is thus not likely to be a major contributor as a carbon and nitrogen source during germination since the gliadins in the endosperm are the major storage proteins. This suggests that these 7S globulin storage proteins may be involved in some other function and not primarily as a carbon and nitrogen source.

During grain development, there is a large accumulation of oxalate and it has been speculated that the 7S storage proteins may have a role in oxalate metabolism; for example, in binding oxalate or detoxifying oxalate (Dunwell et al., 2000). Evidence for this is in the reporting of the structural similarity of the conserved β-barrel structures found in cupins and is suggested that this may be the active site of OXO activity (Gane et al., 1998).

The 7S globulin proteins are trimers in the 50 kDa to 70 kDa molecular weight range. They form a compact structure that protects them from degradation by proteases. 7S globulins are major storage proteins found of legumes and are also a major allergen (Breiteneder and Mills, 2005). The stable structure of the 7S globulins allows them to bypass the intestinal proteases (when ingested by humans) and to be absorbed into the small intestines causing an allergic reaction (Dunwell et al., 2000).

During the two-day imbibition of the grain, the storage protein seems to have been degraded gradually, likely due to its stable compact structure mentioned previously (Dunwell et al., 2000). Degradation products can be seen in the inner tissue fraction 2D gel images (Figs. 3.9 and 3.10). In the acidic pH range (Fig. 3.9) the majority of 7S globulin storage protein fragments were in the low-molecular-weight range, thus showing degradation products. On the other hand, in the alkaline pH range (Fig. 3.10) the majority of these proteins were in the medium to high-molecular-weight range.

#### 4.4.2 Protein synthesis and metabolism

The next major groups of proteins that were identified are involved in protein synthesis, gene expression and energy production. These groups of proteins are essential for the growing plant during germination (Eastwood and Laidman, 1971; Fincher, 1989). The sub-classes of these main groups are gene expression, protein synthesis and folding, citric

acid cycle, glycolysis and gluconeogenisis, oxidative stress, pentose phosphate pathway, carbohydrate metabolism, adenosine triphosphate (ATP) sythesis and C1 metabolism.

#### 4.4.3 Voltage dependent anion channel (VDAC)

Another sub-class of protein that was identified is involved in programmed cell death (PCD). PCD occurs in the inner fraction (aleurone cells) at the beginning of germination (Eastwood and Laidman, 1971; Fincher, 1989). The voltage dependent anion channel (VDAC) protein is a mitochondrial membrane porin protein that regulates the flow of cell metabolites between the cytosol and mitochondria. PCD signaling is activated by various stimuli like hormones or a pathogen response, thus causing the VDAC protein to change its permeability to release intermembrane proteins such as cytochrome *c* and proteases, which are involved in cell-death pathways (Godbole et al., 2003).

#### *4.4.4 α-amylase*

A major hydrolytic enzyme that was not identified in the inner tissue fraction (aleurone cells) was  $\alpha$ -amylase. This is a major hydrolytic enzyme known to be expressed in aleurone cells and secreted into the endosperm during germination (Eastwood and Laidman, 1971; Fincher, 1989; Evers et al., 1999). It is likely that after two days of grain imbibition, all of the  $\alpha$ -amylase had been secreted into the endosperm. Another possibility could be that the  $\alpha$ -amylase was washed out during bran preparation.

#### 4.5 Proteins identified in supernatant from imbibed grain

Endochitinase, histone, OXO, POX, and PPO were all identified in the supernatant from imbibed grain which had leached out during the tissue extraction process. PPO and POX were the only proteins which were not identified in the other tissue fractions. OXO was identified in the outer and intermediate tissue fractions and is likely that it had leached out into the supernatant from these fractions; however, endochitinase and histone were identified in the intermediate and inner fractions respectively. This suggests that these proteins may also have leached out from the inner and intermediate tissue fractions.

#### 4.5.1 Polyphenol oxidase (PPO)

PPO has previously been reported to be found in bran and especially in the aleurone cells (Rani et al., 2001; Demeke and Morris, 2002). However, this study shows that no PPO was identified in the inner fraction (aleurone cells) or any other bran fraction and was identified only in the supernatant collected from grain and isolated outer tissue fraction imbibition. The results suggest that PPO is likely to be located in the outer tissue fraction and is easily leached out in water.

#### 4.5.2 Peroxidase (POX)

POX is a  $H_2O_2$  scavenger. It is a protective enzyme against factors such as pathogen, environmental stress and mechanical tissue damage that may initiate a response that results in oxidation reactions that produce  $H_2O_2$  (Lane, 2000; Almeselmani et al., 2006).

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#### 4.6 Potential applications

Much of this study had focused on identifying the composition and functional distribution of proteins in the bran with respect to its tissue fractions, thus giving a new insight into the function of bran as a whole. Another objective that was hoped to be achieved from this study was the application of this new knowledge to biotechnology, investigating the possibility of new methods and/or processes that will improve flour quality and yield in the milling industry. A few applications potentially exist and will briefly be discussed.

PPO and POX are oxidizing enzymes that were identified in the supernatant from imbibed grain as mentioned previously. PPO and POX are also reported to be involved in the undesirable browning of flat breads, pastas and Asian noodles (Rani et al., 2001; Demeke and Morris, 2002; Aalami et al., 2007). These proteins were identified in the outer bran fraction and in the supernatant. This indicates that these proteins leach out of the grain quite easily when exposed to water. It may be possible that these proteins diffuse and spread out from bran particle contamination in flour when water is added. Washing these proteins out from grain prior to milling may reduce browning of dough caused by these proteins.

Another possible application of the information generated from this study is the potential to use enzymes to hydrolyse bran tissue bonds in order to improve bran separation from endosperm during conditioning, as was mentioned earlier in the discussion. Treating grain with hydrolytic enzymes during the conditioning process to improve bran separation from endosperm has previously been attempted by Novozymes (PCT/DK98/00460, 1998). Their approach was to condition grain with a broad and nonspecific assortment of enzymes to degrade bran tissue bonds in the hope to improve flour yield and to minimise bran contamination in flour during milling. A Patent Cooperation Treaty (PCT) patent application was submitted by Novozymes claiming that their enzyme treatment improved flour yield and shortened conditioning time; however, the patent was not granted and further work was abandoned.

As mentioned earlier in the discussion, many of the proteins identified in the bran layers were involved in pathogen defence. Some of these proteins are hydrolytic enzyme inhibitors such as xylanase and *a*-amylase inhibitors. These inhibitor proteins could be a problem when treating grain with enzymes during conditioning as was attempted by Novozymes. A possible way to overcome this problem is to increase the concentration of enzymes in the conditioning solution; however, this approach is not cost effective when millions of tonnes of wheat are milled annually. Studies on the wheat xylanase inhibitor XIP-I has shown that recombinant xylanases derived from fungi were inhibited, whereas xylanases derived from bacteria were not inhibited (Flatman et al., 2002). These bacterially derived xylanases will potentially be more specific and effective in hydrolyzing arabinoxylans (AX's) in bran tissue. This kind of approach will require enzymes that are more specific to the target tissue material and also to be not inhibited by the enzyme inhibitors identified in this study.

Further work will be required to analyse the composition of the tissue material at the junction between bran layers to find suitable enzymes for hydrolysis of cell wall

components. This kind of bran fractionation approach will be of great value to the milling industry as one of their primary aims is to separate cleanly bran or bran fractions from endosperm during milling (Butcher and Stenvert, 1973; Moss et al., 1980; Every et al., 2002).

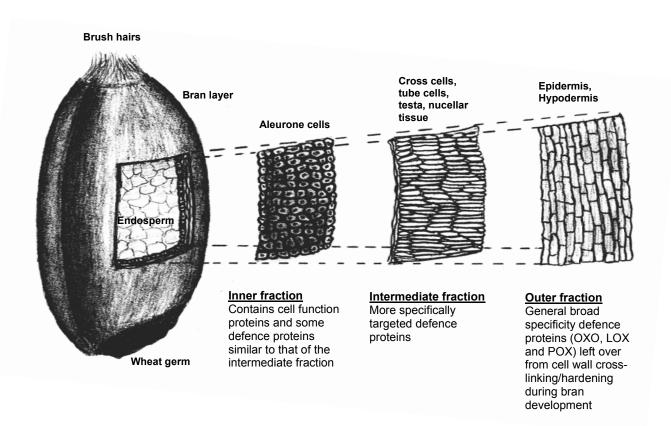
The amino acids arginine and lysine have great nutritional value in foods containing these amino acids (van der Meer et al., 2001; Flynn et al., 2002). Since the 7S globulin storage proteins are abundant in the inner fraction, the arginine and lysine content was calculated to assess the nutritional value of this protein. The arginine content in the 7S globulin storage protein is 14.9%, and thus contributes to the high arginine content previously reported in the inner (aleurone) fraction (12.3%). The inner fraction is also reported to have approximately 2.5 times higher arginine content when compared to the arginine content in the whole grain (4.6%). The lysine content however, is only 2.2% of the amino acid composition of the 7S globulin protein and thus is not likely to be a major contributor to the major lysine containing proteins in the inner fraction previously reported to be 5.9% lysine. The lysine-content in the inner fraction is also two times higher than in the whole grain (Spurway, 1998); thus, the inner fraction is important with respect to the arginine and lysine content, and introducing the inner (aleurone) fraction into flour will be nutritionally beneficial. One potential way to do this is to specifically degrade the bran tissue bonds between the inner fraction (aleurone cells) and the outer bran fractions by using enzymes as described earlier.

### 5. Conclusion and future directions

An overall summary of the broad functional groups of proteins identified in this study were the defence-and stress-related proteins found in the outer and intermediate tissue fractions and storage and metabolic proteins in the inner layers. Around 80% of proteins in the outer layers were associated with various defence-and stress-related functions. Defence-and stress-related proteins were also identified in the inner fraction, however they comprised only around 5% of the protein array. Interestingly, the major protein in the inner fraction was 7S globulin storage protein that was 57.6% of the total protein spots. Water-soluble proteins that have leached out during grain imbibition were also mostly involved in oxidative-stress response and/or have anti-fungal activity. The location of the defence-and stress-related proteins in the outer and intermediate tissues fractions is thus ideal in acting as a barrier, protecting the grain from environmental and pathogenic damage.

Consolidating this summary, a model for bran development and its role in biotic-and abiotic-defence is shown below in Fig. 5.1. This model illustrates the difference in specificity of the identified defence-related proteins contained in each tissue fraction outlined in the Discussion. Starting with the outer fraction, there were general defence-related proteins that are likely to be left over from the formation of this tissue fraction during bran development as they are associated with lignin cross-linking to form a tough outer tissue barrier (Gane et al., 1998; Peyron et al., 2001; Antoine et al., 2003). The intermediate fraction contains a much broader and more specific set of defence-related proteins. They are distributed throughout this compact multilayer tissue fraction which

also acts as a physical barrier. Finally, the inner fraction also contains some specific defence-related proteins and cell function proteins. However, the major protein (7S globulin storage protein) in this fraction is likely to be involved in defence against oxalate-secreting fungi and also to serve in protecting itself during grain development when there is potentially high oxalate levels (Dunwell et al., 2000). The model also



**Figure 5.1** Schematic diagram of a model for the development and defence protein distribution in bran showing the three tissue fractions analysed in this study and their overall role as a protective barrier for wheat grain.

shows the structural difference of each bran tissue fraction (as mentioned previously) that collectively forms a physical barrier to protect the endosperm. This physical property difference of each tissue fraction allowed it to separate from each other during tissue fraction isolation, thus suggesting that the proteins identified in this study are likely to be representative of each tissue fraction.

In conclusion, this proteomic study of bran (never before attempted to our knowledge) has given an insight into the location of protein complements with respect to the different tissue fractions. The location and functions of these proteins illustrates the function of bran as a whole. This was shown in terms of the stress- and defence-related proteins forming an outer protective barrier and the inner layer mostly containing cupin family storage proteins and the biochemical machinery necessary for germination. Lastly, protein maps generated in this study may be useful in comparing protein compositions of different wheat cultivars with respect to their farming and milling properties for the purpose of selective breeding programs.

## 6. Appendix

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A-1 Protein quantification

## A-1.1 Protein quantification of tissue fractions

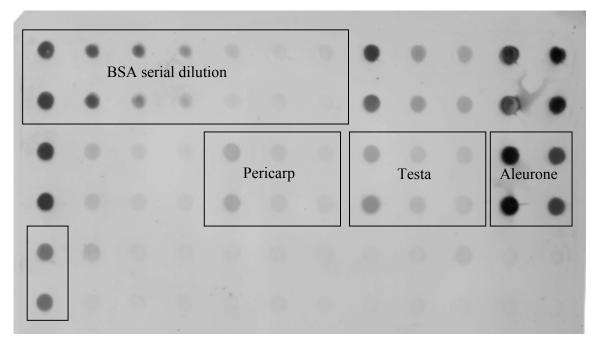


Figure A-1.1 Proteins spotted onto nitrocellulose membrane and stained with Deep

Purple

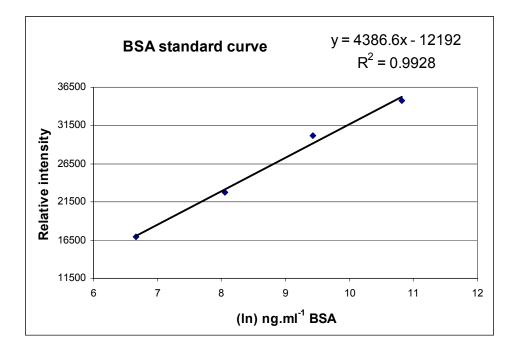


Figure A-1.2 BSA standard curve equation

BSA ng.ml <sup>-1</sup>	BSA (ln) ng.m[ <sup>1</sup>	Intensity	Intensity	Ave. intensity
50000	10.81977828	34398	35099	34748.5
12500	9.43348392	32868	27439	30153.5
3125	8.04718956	24912	20570	22741.0
781	6.66057515	17772	16123	16947.5

**Table A-1.1** BSA standard curve calculations

## A-1.2 Protein quantification of supernatant

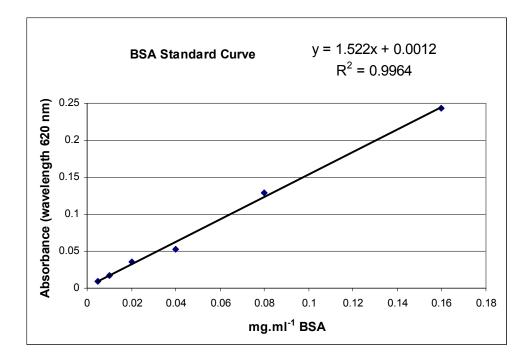


Figure A-1.3 BSA standard curve equation

 Table A-1.2 BSA standard curve calculations

BSA mg.mt <sup>-1</sup>			Average Absorbance	Blank Subtracted
0.160	0.618	0.653	0.635	0.2435
0.080	0.503	0.539	0.521	0.1290
0.040	0.429	0.460	0.444	0.0525
0.020	0.425	0.431	0.428	0.0360
0.010	0.409	0.408	0.408	0.0165
0.005	0.399	0.403	0.401	0.0090
Blank	0.399	0.385	0.392	

**Table A-1.3** Protein concentrations determined for whole grain and isolated outer

 fraction supernatant

Sample			Average Absorbance	Blank Subtracted	mg.ml <sup>1</sup>	mg.g <sup>-1</sup>
Whole grain	0.473	0.465	0.469	0.077	0.04980	NA
Outer fraction	0.408	0.433	0.421	0.029	0.01794	0.6

A-2 Protein identification tables

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
1	Oxalate oxidase precursor	70		29 / 5.2	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
2	Oxalate oxidase precursor	77		29 / 5.22	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
3	Oxalate oxidase precursor	20		30 / 5.26	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
4	Oxalate oxidase precursor	85		25 / 4.9	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
5	Oxalate oxidase precursor	84		25 / 5.0	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
6	Oxalate oxidase precursor	72		25 / 5.1	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
7	Oxalate oxidase precursor	74		25 / 5.2	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
8	Oxalate oxidase precursor	99		25 / 5.22	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
9	Oxalate oxidase precursor	70		25 / 5.25	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
10	Oxalate oxidase precursor	56		25 / 5.5	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
	Oxalate oxidase precursor	64		22 / 4.75	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
12	Oxalate oxidase precursor	73		22 / 4.8	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
13	Oxalate oxidase precursor	43		22 / 4.9	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
14	Oxalate oxidase precursor	81		24 / 5.2	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
15	Oxalate oxidase precursor	68		24 / 5.22	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
16	Oxalate oxidase precursor	53		20 / 6.4	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
17	Oxalate oxidase precursor	57		20 / 6.6	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
	Oxalate oxidase precursor	79		20 / 6.76	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0 WHEAT	

**Table A-2.1** Outer fraction (epidermis and hypodermis) proteins identified from 2D gel pH 4-7

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
1	Oxalate oxidase precursor	77		14 / 6.1	3 / 12	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
2	Oxalate oxidase precursor	45		14 / 6.3	3 / 12	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
3	Oxalate oxidase GF-2.8 precursor (EC 1.2.3.4) (Germin GF-2.8)	92		12 / 6.1	5 / 12	23.6 / 6.41	T. aestivum	GER2_WHEAT; P15290	
4	Oxalate oxidase GF-3.8 precursor (EC 1.2.3.4) (Germin GF-3.8)	98		12 / 6.3	3 / 12	23.5 / 6.9	T. aestivum	GER3_WHEAT; P26759	
5	Oxalate oxidase GF-2.8 precursor (EC 1.2.3.4) (Germin GF-2.8)	72		12 / 6.5	4 / 11	23.6 / 6.41	T. aestivum	GER2_WHEAT; P15290	
6	Oxalate oxidase GF-2.8 precursor (EC 1.2.3.4) (Germin GF-2.8)	71		6.5 / 6.1	4 / 16	23.6 / 6.41	T. aestivum	GER2_WHEAT; P15290	
7	Oxalate oxidase precursor	-4.2 <sup>1</sup>		6.5 / 6.4	1 / 6.7	23.5 / 6.9	T. aestivum	gi 46408929	
8	Oxalate oxidase GF-2.8 precursor (EC 1.2.3.4) (Germin GF-2.8)	73		6.5 / 6.5	7 / 21	23.6 / 6.41	T. aestivum	GER2_WHEAT; P15290	
9	Oxalate oxidase GF-2.8 precursor (EC 1.2.3.4) (Germin GF-2.8)	52		6.5 / 7.2	3 / 11	23.6 / 6.41	T. aestivum	GER2_WHEAT; P15290	
10	No match								
11	PLAT (Polycystin-1, Lipoxygenase, Alpha-Toxin) domain or LH2 (Lipoxygenase homology 2) domain	-7.5		6 / 9.6	1 / 5.2	26.5 / 9.3	T. aestivum	gnl CDD 28747	
12	Type 1 non-specific lipid transfer protein precursor	57		5 / 9.6	1 / 9	12.4 / 9.46	T. aestivum	Q5NE29_WHEAT	
13	No match								
14	No match								
15	No match								
16	No match								
17	Type 1 non-specific lipid transfer protein precursor	97		4 / 10.3	2 / 14	12.4 / 9.46	T. aestivum	Q5NE29_WHEAT	

 Table A-2.2
 Outer fraction (epidermis and hypodermis) proteins identified from 2D gel pH 6-11

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
1	Oxalate oxidase precursor	70		32 / 5.1	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
2	Oxalate oxidase precursor	89		31 / 5.2	1 / 7	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
3	Alpha-amylase inhibitor	141		27 / 6.8	4 / 22	19.6 / 6.77	T. aestivum	S38955	
4	Alpha-amylase inhibitor	183		27 / 6.9	4 / 22	19.6 / 6.77	T. aestivum	S38955	
5	Pathogenesis-related protein 4 (Fragment)	30		16 / 4.6	1 / 6	13.1 / 7.0	T. aestivum	Q9SQG8_WHEAT	
6	Wheat EST match	73	Chain B, Crystal Structure Of The T. aestivum Xylanase Inhibitor-I In Complex With Aspergillus Niger Xylanase-I	14 / 4.9	8 / 10	33.1 / 9.45	T. aestivum	Ta78954-3 <sup>4</sup>	GI:55669878
7	Wheat EST match	63	Xylanase inhibitor	14 / 5.1	10 / 9	47.2 /8.8	T. aestivum	Ta745253405-2	GI:23954367
8	Wheat EST match	90	Chain B, Crystal Structure Of The T. aestivum Xylanase Inhibitor-I In Complex With Aspergillus Niger Xylanase-I	15 / 5.2	9 / 17	33.1 / 9.45	T. aestivum	Ta78954-3	GI:55669878
9	No match								
10	Wheat EST match	145	Xylanase inhibitor	14 / 5.5	14 / 22	47.2 /8.8	T. aestivum	Ta745253405-2	GI:23954367
11	Wheat EST match	105	Wheatwin1	16 / 6.3	4 / 25	25.6 / 9.53	T. aestivum	Ta7944-1	GI:3135957
12	Pathogenesis-related protein 4 (Fragment)	25		17 / 6.8	1 / 10	13.1 / 7.0	T. aestivum	Q9SQG8_WHEAT	
13	Wheat EST match	33	Uncoupling protein	10 / 4.7	11 / 44	18.7 / 11.83	T. aestivum	Ta192131-2	GI:10716672
14	No match								
15	No match								

 Table A-2.3
 Intermediate fraction (nucellar tissue, testa, tube cells and cross cells) proteins identified from 2D gel pH 4-7

15 No match

16 No match

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
1	No match								
2	Actin (065316)	265		50 / 8.5	15 / 35	41.6 / 5.3	M. viride	ACT_MESVI	
3	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	144		50 / 8.8	14 / 33	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
4	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	63		50 / 9.1	11 / 40	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
5	Chain B, Crystal Structure Of Chitinase At 1.91a Resolution	128		49 / 8.5	6 / 27	26 / 8.45	H. vulgare	gi 1310889	
6	Actin (O65316)	200		49 / 9.0	16 / 34	41.6 / 5.3	M. viride	ACT_MESVI	
7	Actin (O65316)	281		47 / 9.1	14 / 32	41.6 / 5.3	M. viride	ACT_MESVI	
8	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	117		29 / 6.9	4 / 14	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
9	26 kDa endochitinase 2 precursor (EC 3.2.1.14) (CHI-26)	76		28 / 7.1	6 / 27	28.1 / 8.54	H. vulgare	CHI2_HORVU; P23951	
10	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	339		28 / 7.2	23 / 51	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
11	Secretory protein	76		27 / 7.4	7 / 32.6	24.2 / 9.32	T. aestivum	Q9SWZ5_WHEAT	
12	Chain A, Crystal Structure Of Family 11 Xylanase In Complex With Inhibitor (Xip-I)	206		30 / 7.5	11 / 33	30.3 / 8.27	T. aestivum	gi 51247633	
13	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	167		31 / 7.9	4 / 14	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
14	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	413		28 / 7.8	22 / 53	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
15	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	217		28 / 8.0	17 / 48	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	

Table A-2.4 Intermediate fraction (nucellar tissue, testa, tube cells and cross cells) proteins identified from 2D gel pH 6-11	1
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Table A-2.4	(continued)
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	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
16	Xylanase inhibitor protein I	201		26 / 8.0	11 / 29	33.2 / 8.66	T. aestivum	gi 20804336	
17	Thaumatin-like protein	85		27 / 8.3	3 / 13	23.6 / 7.85	T. aestivum	Q8S4P7_WHEAT	
18	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	137		28 / 8.4	14 / 40	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
19	26 kDa endochitinase 2 precursor (EC 3.2.1.14) (CHI-26)	100		28 / 8.5	7 / 29	28.1 / 8.54	H. vulgare	CHI2_HORVU; P23951	
20	26 kDa endochitinase 1 precursor	118		27 / 8.9	5 / 20	33.4 / 8.54	H. vulgare	gi 2506281	
21	Actin (O65316)	309		29 / 9.1	19 / 32	41.6 / 5.3	M. viride	ACT_MESVI	
22	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	287		29 / 9.3	12 / 30	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
23	Endogenous alpha-amylase/subtilisin inhibitor (WASI)	305		29 / 9.4	19 / 76	19.6 / 6.77	T. aestivum	IAAS_WHEAT; P16347	
24	No match								
25	Basic endochitinase A precursor (EC 3.2.1.14) (Rye seed chitinase-a) (RSC-a)	97		35 / 9.7	9 / 38	33.4 / 8.28	S. cereale	CHIA_SECCE; Q9FRV1	
26	26 kDa endochitinase 2 precursor (EC 3.2.1.14) (CHI-26)	80		33 / 9.7	5 / 18	28.1 / 8.83	H. vulgare	CHI2_HORVU; P23951	
27	Class II chitinase	-3.7		26 / 6.6	1 / 5.6	28.2 / 8.7	T. aestivum	gi 62465516	
28	Class II chitinase	-10.5		26 / 7.1	1 / 5.6	28.2 / 8.7	T. aestivum	gi 62465516	
29	Class II chitinase	-21.6		26 / 7.2	1 / 5.6	28.2 / 8.7	T. aestivum	gi 62465516	
30	Xylanase inhibitor protein I	164		26 / 7.3	11 / 30	33.2 / 8.66	T. aestivum	gi 20804336	
31	Plant Basic Secretory Protein	-3.1		26 / 7.4	1 / 5.6	25.4 / 4.7	T. aestivum	gi 1323750	
32	Xylanase inhibitor protein I	202		26 / 7.5	11 / 30	33.2 / 8.66	T. aestivum	gi 20804336	
33	Alpha amylase inhibitor	-1.2		26 / 7.5	1/3.9	19.6 / 6.7	T. aestivum	gi 225042	
34	Class II chitinase	-13.6		26 / 7.8	2 / 11	28.2 / 8.7	T. aestivum	gi 62465516	
35	Endogenous alpha-amylase/subtilisin inhibitor (WASI)	362		25 / 7.8	17 / 72	19.6 / 6.77	T. aestivum	IAAS_WHEAT; P16347	

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
36	Chain A, Crystal Structure Of Xylanase Inhibitor Protein (Xip-I)	-23.9		26 / 8.0	4 / 14	30.3 / 8.2	T. aestivum	gi 31615809	
37	Xylanase inhibitor protein I	124		26 / 8.2	9 / 21	33.2 / 8.66	T. aestivum	gi 20804336	
38	Xylanase inhibitor protein I	201		26 / 8.4	10 / 26	33.2 / 8.66	T. aestivum	gi 20804336	
39	Xylanase inhibitor protein I	224		27 / 8.5	12/37	33.2 / 8.66	T. aestivum	gi 20804336	
40	Xylanase inhibitor protein I	319		26 / 8.5	16 / 49	33.2 / 8.66	T. aestivum	gi 20804336	
41	26 kDa endochitinase 1 precursor	118		25 / 8.6	5 / 20	33.4 / 8.54	H. vulgare	gi 2506281	
42	Class II chitinase	-19.7		25 / 8.7	1 / 5.6	28.2 / 8.7	T. aestivum	gi 62465516	
43	26 kDa endochitinase 1 precursor	130		25 / 9.0	6 / 27	33.4 / 8.54	H. vulgare	gi 2506281	
44	Class II chitinase	-23.4		25 / 9.3	2 / 11	28.2 / 8.7	T. aestivum	gi 62465516	
45	Class II chitinase	-37.1		26 / 9.3	1 / 5.6	28.2 / 8.7	T. aestivum	gi 62465516	
46	Chain A, Crystal Structure Of Xylanase Inhibitor Protein (Xip-I)	-60.7		27 / 9.4	5 / 21	30.3 / 8.2	T. aestivum	gi 31615809	
47	No match								
48	Class II chitinase	-33.4		26 / 9.7	2 / 11	28.2 / 8.7	T. aestivum	gi 62465516	
49	Class II chitinase (EC 3.2.1.14)	72		26 / 6.9	1 / 6	28.2 / 8.66	T. aestivum	Q4Z8L7_WHEAT	
50	26 kDa endochitinase 1 precursor	106		25 / 7.1	4 / 16	33.4 / 8.54	H. vulgare	gi 2506281	
51	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	155		25 / 7.5	4 / 16	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
52	26 kDa endochitinase 1 precursor	105		25 / 7.7	4 / 16	33.4 / 8.54	H. vulgare	gi 2506281	
53	Chain A, Crystal Structure Of Xylanase Inhibitor Protein (Xip-I)	-9.6		25 / 7.8	2 / 7.7	30.3 / 8.2	T. aestivum	gi 31615809	
54	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-1 protein)	470		25 / 8.0	21 / 51	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
55	Chain B, Crystal Structure Of Chitinase At 1.91a Resolution	58		25 / 8.1	6 / 17	26 / 8.45	H. vulgare	gi 1310889	

Table A-2.4	(continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
56	Class II chitinase	-26.6		25 / 8.4	2 / 5.6	28.2 / 8.7	T. aestivum	gi 62465516	
57 58	Chain A, Crystal Structure Of Xylanase Inhibitor Protein (Xip-I) No match	-46.9		24 / 8.6	6 / 18	30.3 / 8.2	T. aestivum	gi 31615809	
59	Class II chitinase	-9		24 / 8.9	1 / 5.6	28.2 / 8.7	T. aestivum	gi 62465516	
60	Endogenous alpha-amylase/subtilisin inhibitor (WASI)	158		24 / 9.4	15 / 70	19.6 / 6.77	T. aestivum	IAAS_WHEAT; P16347	
61	26 kDa endochitinase 1 precursor	177		25 / 9.2	6 / 20	33.4 / 8.54	H. vulgare	gi 2506281	
62	26 kDa endochitinase 1 precursor	196		25 / 9.2	6 / 20	33.4 / 8.54	H. vulgare	gi 2506281	
63	Chitinase a	81		25 / 9.4	5 / 24	31.7 / 8.13	S. cereale	gi 741317	
64	Xylanase inhibitor precursor (Xylanase inhibitor TAXI-I)	140		27 / 9.4	2 / 8	40.9 / 8.18	T. aestivum	Q8H0K8_WHEAT	
65	Chain A, Crystal Structure Of Family 11 Xylanase In Complex With Inhibitor (Xip-I)	325		22 / 6.7	14 / 37	30.3 / 8.3	T. aestivum	gi 51247633	
66	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	130		21 / 7.0	3 / 10	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
67	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog) (XIP-I protein)	59		21 / 7.1	2 / 6	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
68	Endogenous alpha-amylase/subtilisin inhibitor (WASI)	167		22 / 7.4	11 / 32	19.6 / 6.8	T. aestivum	gi 123975	
69	Xylanase inhibitor XIP-III	151		24 / 7.9	13 / 24	33.3 / 7.1	T. aestivum	gi 66766322	
70	No match								
71	Xylanase inhibitor protein I	184		24 / 8.6	11 / 26	33.2 / 8.7	T. aestivum	gi 20804336	
72	Class II chitinase	120		23 / 8.6	5 / 19	28.2 / 8.7	T. aestivum	gi 62465514	
73	Class II chitinase	135		22 / 8.5	5 / 24	28.2 / 8.7	T. aestivum	gi 62465514	

Table A-2.4	(continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
74	No match								
75	Benzothiadiazole-induced protein (clone WCI-5)	104		22 / 8.7	1 / 6	25.4 / 4.65	T. aestivum	T06278	
76	26 kDa endochitinase 1 precursor	53		24 / 8.9	4 / 16	33.4 / 8.54	H. vulgare	gi 2506281	
77	Secretory protein	68		23 / 9.1	4 / 13	24.2 / 9.3	T. aestivum	gi 5669008	
78	26 kDa endochitinase 1 precursor	76		24 / 9.1	4 / 16	33.4 / 8.54	H. vulgare	gi 2506281	
79	Class II chitinase	97		22 / 9.2	4 / 14	28.2 / 8.7	T. aestivum	gi 62465514	
80	26 kDa endochitinase 1 precursor	108		24 / 9.2	4 / 16	33.4 / 8.54	H. vulgare	gi 2506281	
81	Unknown	93		24 / 9.4	3 / 14	25.4 / 4.65	T. aestivum	gi 1323750	
82	Benzothiadiazole-induced protein (clone WCI-5)	93		22 / 9.4	1 / 6	25.4 / 4.65	T. aestivum	T06278	
83	Endogenous alpha-amylase/subtilisin inhibitor (WASI)	110		19 / 6.5	7 / 32	19.6 / 6.77	T. aestivum	gi 123975	
84	Endogenous alpha-amylase/subtilisin inhibitor (WASI)	251		19 / 6.7	15 / 62	19.6 / 6.77	T. aestivum	gi 123975	
85	Chain A, Crystal Structure Of Family 11 Xylanase In Complex With Inhibitor (Xip-I)	289		19 / 6.9	14 / 33	30.3 / 8.3	T. aestivum	gi 51247633	
86	Class II chitinase	104		17 / 7.1	4 / 14	28.2 / 8.7	T. aestivum	gi 62465514	
87	Chain A, Crystal Structure Of Family 11 Xylanase In Complex With Inhibitor (Xip-I)	353		19 / 7.1	15 / 37	30.3 / 8.3	T. aestivum	gi 51247633	
88	Secretory protein	99		21 / 7.4	5 / 13	24.2 / 9.3	T. aestivum	gi 5669008	
89	Chain A, Crystal Structure Of Family 11 Xylanase In Complex With Inhibitor (Xip-I)	409		19 / 8.0	15 / 37	30.3 / 8.3	T. aestivum	gi 51247633	
90	Chain A, Crystal Structure Of Family 11 Xylanase In Complex With Inhibitor (Xip-I)	138		16 / 8.7	9 / 30	30.3 / 8.3	T. aestivum	gi 51247633	

 Table A-2.4 (continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
91	26 kDa endochitinase 1 precursor	80		15 / 9.0	4 / 16	33.4 / 8.54	H. vulgare	gi 2506281	
92	Chain A, Crystal Structure Of Family 11 Xylanase In Complex With Inhibitor (Xip-I)	236		19 / 9.1	13 / 33	30.3 / 8.3	T. aestivum	gi 51247633	
93	No match								
94	Chain A, Crystal Structure Of Family 11 Xylanase In Complex With Inhibitor (Xip-I)	141		11 / 9.4	9 / 22	30.3 / 8.3	T. aestivum	gi 51247633	
95	Pathogenesis-related protein 4 (Fragment)	67		7 / 6.5	1 / 10	13.1 / 7.0	T. aestivum	Q9SQG8_WHEAT	
96	Pathogenesis-related protein 4 (Fragment)	74		6 / 7.6	1 / 10	13.1 / 7.0	T. aestivum	Q9SQG8_WHEAT	
97	Pathogenesis-related protein 4 (Fragment)	62		6 / 7.7	1 / 10	13.1 / 7.0	T. aestivum	Q9SQG8_WHEAT	
98	No match								
99	No match								
100	Class II chitinase (EC 3.2.1.14)	82		8 / 9.5	1 / 6	28.2 / 8.66	T. aestivum	Q4Z8L7_WHEAT	
101	Class II chitinase (EC 3.2.1.14)	46		8 / 9.7	1 / 6	28.2 / 8.66	T. aestivum	Q4Z8L7_WHEAT	
102	Xylanase inhibitor protein I	91		2 / 8.7	4 / 11	33.2 / 8.7	T. aestivum	gi 20804336	
103	Chain A, Crystal Structure Of Family 11 Xylanase In Complex With Inhibitor (Xip-I)	146		4 / 9.3	5 / 12	30.3 / 8.3	T. aestivum	gi 51247633	

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
1	No match								
2	Pyruvate orthophosphate dikinase (Fragment)	42		110 / 5.4	20 / 38	32.6 / 4.97	T. aestivum	Q7XYB5_WHEAT	
3	No match								
4	Wheat EST match	80	Aconitate hydratase, cytoplasmic, putative, expressed	100 / 6.2	25 / 30	83.5 / 7.66	O. sativa	Ta18566-2 <sup>4</sup>	GI:108706066
5	No match								
6	No match								
7	HSP70	255		75 / 5.2	24 / 32	70.99 / 5.14	T. aestivum	Q9SAU8_WHEAT	
8	HSP70	294		75 / 5.25	29 / 32	70.99 / 5.14	T. aestivum	Q9SAU8_WHEAT	
9	HSP70	229		75 / 5.3	37 / 38	70.99 / 5.14	T. aestivum	Q9SAU8_WHEAT	
10	HSP70	150		75 / 5.35	29 / 29	70.99 / 5.14	T. aestivum	Q9SAU8_WHEAT	
11	Putative dnaK-type molecular chaperone	183		74 / 5.45	18 / 22	72.9 / 5.49	O. sativa	Q6Z7L1_ORYSA	
	No match	~ .			<i>(</i> / <b>2</b> )	10 1 10 50	-		
	Cyclophilin A-2 (EC 5.2.1.8) (Cyclophilin A)	21		77 / 6.15	6 / 28	18.4 / 8.52	T. aestivum	Q93XQ6_WHEAT	
14	No match								
15	No match								
16	2,3-bisphosphoglycerate-independent phosphoglycerate mutase	121		72 / 5.8	10 / 11	60.6 / 5.29	Z. mays	PMGI_MAIZE, P30792	2
17	Putative 2,3-bisphosphoglycerate- independent phosphoglycerate mutase	94		70 / 5.9	14 / 21	60.8 / 5.25	O. sativa	Q5KQH5_ORYSA	
18	Phosphoglucomutase (EC 5.4.2.2) (Fragment)	221		74 / 5.95	17 / 23	62.8 / 5.66	T. aestivum	Q8VX48_WHEAT	
19	Wheat EST match	54	Cytosolic NADP malic enzyme	72 / 6.3	12 / 42	27.4 / 8.8	O. sativa	Ta963253410-3	GI:38261493
20	Wheat EST match	122	Cytosolic NADP malic enzyme	72 / 6.35	13 / 43	27.4 / 8.8	O. sativa	Ta965253413-3	GI:38261493
21	7S Globulin storage protein	55	·	74 / 6.75	11 / 11	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
22	7S Globulin storage protein	50		74 / 6.8	14 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
23	Protein disulfide isomerase (EC 5.3.4.1) (Fragment)	118		65 / 4.95	12 / 88	10.2 / 5.43	T. aestivum	Q6JAB7_WHEAT	

**Table A-2.5** Inner bran fraction (aleurone cells) proteins identified from 2D gel pH 4-7

Table A-2.5	(continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
24	Wheat EST match	100	Putative rubisco subunit binding- protein alpha subunit precursor	57 / 5.15	4 / 67	6.1 / 4.08	O. sativa	Ta228991-3	GI:50920285
25	60 kDa chaperonin	129	r r	56 / 5.15	8 / 9	57.1 / 5.05	X. oryzae	Q2NY29_XANOR	
26	No match								
27	Chaperonin CPN60-1, (HSP60-1) mitochondrial precursor	40		57 / 5.25	18 / 30	61.2 / 5.68	Z. mays	CH61_MAIZE	
28	No match				<b>A</b> ( ) <b>(</b> )	17 0 / 5 / 4	<b>.</b> .		
29	Enolase (EC 4.2.1.11)	512		53 / 5.6	26 / 46	47.9 / 5.41	O. sativa	ENO_ORYSA	
30	Xylose isomerase (EC 5.3.1.5)	181		52 / 5.55	14 / 17	53.6 / 5.31	H. vulgare	XYLA_HORVU	
31	Enolase (EC 4.2.1.11)	469		52 / 5.6	28 / 46	47.9 / 5.41	O. sativa	ENO_ORYSA	
32	Enolase (EC 4.2.1.11)	648		52 / 5.7	32 / 52	47.9 / 5.41	O. sativa	ENO_ORYSA	
33	Atp1 protein	305		52 / 5.75	27 / 41	55.3 / 5.7	T. aestivum	Q332R4_WHEAT	
34	No match								
35	Atp1 protein	85		52 / 5.9	19 / 33	55.3 / 5.7	T. aestivum	Q332R4_WHEAT	
36	Putative 2,3-bisphosphoglycerate- independent phosphoglycerate mutase	61		67 / 6.0	19 / 21	60.8 / 5.25	O. sativa	Q5KQH5_ORYSA	
37	Atp1 protein	51		52 / 6.05	27 / 41	55.3 / 5.7	T. aestivum	Q332R4_WHEAT	
38	Wheat EST match	165	7S Globuliin storage protein	51 / 6.05	42 / 47	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
39	Em protein H5	20		49 / 6.25	7 / 60	10.05 / 5.14	T. aestivum	EM4_WHEAT	
40	Putative dihydrolipoamide dehydrogenase	42		53 / 6.4	7 / 10	52.6 / 7.21	O. sativa	Q9ASP4_ORYSA	
41	Putative dihydrolipoamide dehydrogenase	186		52 / 6.6	11 / 17	52.6 / 7.21	O. sativa	Q9ASP4_ORYSA	
42	Globosa	23		51 / 6.6	5 / 16	24.1 / 7.14	T. aestivum	Q6QPJ1_WHEAT	
43	7S Globuliin storage protein	74		50 / 6.7	14 / 17	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
44	7S Globuliin storage protein	80		50 / 6.8	13 / 14	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
45	Putative beta-N-acetylhexosaminidase (Fragment)	23		51 / 4.75	4 / 66	5.98 / 10.06	T. aestivum	Q2L3V7_WHEAT	
46	No match								
47	No match								
48	ATP synthase beta subunit	475		52 / 5.25	28 / 39	59.2 / 5.56	T. aestivum	Q41534_WHEAT	

Table A-2.5	(continued)
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	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
49	ATP synthase beta subunit	598		53 / 5.3	35 / 49	59.2 / 5.56	T. aestivum	Q41534_WHEAT	
50	Wheat EST match	43	MLo protein	52 / 5.3	3 / 10	40.5 / 8.88	T. aestivum	Ta224200-1	GI:14334167
51	UTPglucose-1-phosphate uridylyltransferase (EC 2.7.7.9)	418		51 / 5.3	23 / 37	51.6 / 5.2	H. vulgare	UGPA_HORVU	
52	No match								
53	No match								
54	No match								
55	No match								
56	No match								
57	No match								
58	No match								
59	Annexin	29		50 / 5.25	10 / 27	35.4 / 9.01	T. aestivum	Q6S9D8_WHEAT	
60	No match							· _	
61	Wheat EST match	38	12-oxo-phytodienoic acid reductase	47 / 5.35	5 / 11	38.9 / 6.33	Z. mays	Ta139439-3	GI:63021725
62	No match								
63	No match								
64	No match								
65	Phosphoglycerate kinase, cytosolic (EC 2.7.2.3)	84		42 / 5.7	11 / 33	42.1 / 5.64	T. aestivum	PGKY_WHEAT	
66	Wheat EST match	43	Subtilisin-like protease	45 / 5.7	4 / 11	39.9 / 9.32	T. aestivum	Ta250319-2	GI:86439745
67	Putative cytosolic 6-phosphogluconate dehydrogenase	160		48 / 5.8	17 / 18	53.0 / 5.92	Z. mays	O81237_MAIZE	
68	Wheat EST match	358	7S Globulin storage protein	48 / 6.5	49 / 51	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
69	7S Globulin storage protein	64		47 / 6.45	26 / 28	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
70	Formate dehydrogenase (Fragment)	181		36 / 6.7	13 / 40	28.99 / 8.61	T. aestivum	Q7X9L3_WHEAT	
71	No match								
72	No match								
73	No match								
74	No match								
75	No match								
76	Elongation factor Tu	47		43 / 5.4	16 / 23	43.1 / 5.45	X. oryzae	Q2NZX1_XANOR	

 Table A-2.5 (continued)

-	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
77	No match								
78	No match								
79	No match								
80	No match								
81	No match								
82	Phosphoglycerate kinase, cytosolic (EC 2.7.2.3)	82		38 / 5.75	14 / 40	42.1 / 5.64	T. aestivum	PGKY_WHEAT	
83	No match								
84	HSP70	276		39 / 5.8	19 / 16	70.99 / 5.14	T. aestivum	Q9SAU8_WHEAT	
85	HSP70	198		39 / 5.8	16 / 16	70.99 / 5.14	T. aestivum	Q9SAU8_WHEAT	
86	HSP70	43		39 / 6.0	12 / 16	70.99 / 5.14	T. aestivum	Q9SAU8_WHEAT	
87	HSP70	304		39 / 6.1	23 / 17	70.99 / 5.14	T. aestivum	Q9SAU8_WHEAT	
88	NADP-specific isocitrate dehydrogenase (EC 1.1.1.42)	71		39 / 6.2	12 / 21	46.1 / 6.29	O. sativa	Q9XGU7_ORYSA	
89	Cytosolic glyceraldehyde-3-phosphate dehydrogenase GAPDH (Fragment)	63		38 / 6.2	5 / 23	25.3 / 7.83	T. aestivum	Q9M4V4_WHEAT	
90	Cytosolic glyceraldehyde-3-phosphate dehydrogenase GAPDH (Fragment)	68		38 / 6.3	11 / 39	25.3 / 7.83	T. aestivum	Q9M4V4_WHEAT	
91	Cytosolic glyceraldehyde-3-phosphate dehydrogenase GAPDH (Fragment)	56		39 / 6.3	10 / 32	25.3 / 7.83	T. aestivum	Q9M4V4_WHEAT	
92	Cytosolic glyceraldehyde-3-phosphate dehydrogenase GAPDH (Fragment)	135		37 / 6.65	9 / 32	25.3 / 7.83	T. aestivum	Q9M4V4_WHEAT	
93	No match								
94	No match								
95	No match								
96	No match								
97	No match								
98	No match								
99	Glutamine synthetase isoform GS1c	181		38 / 5.45	10 / 12	39.2 / 5.41	T. aestivum	Q45NB5_WHEAT	

 Table A-2.5 (continued)

Spot	Protein name	Mascot	Protein name	Observed	Matching	Theoretical <sup>3</sup>	Species with	Swiss-Prot accession	GenBank
No.	(Mascot cereal search)	or GPM <sup>1</sup> score	(Wheat EST BLAST search) <sup>2</sup>	MW (KDa)/pI	peptides / %Coverage	MW (Da)/pI	homologous protein	number <sup>4</sup>	accession number
100	Reversibly glycosylated polypeptide	156		38 / 5.5	18 / 29	41.5 / 5.82	T. aestivum	Q9ZR33_WHEAT	
101	Phosphoglycerate kinase, cytosolic (EC 2.7.2.3)	245		38 / 5.6	14 / 27	42.1 / 5.64	T. aestivum	PGKY_WHEAT	
102	No match								
103	Cytosolic malate dehydrogenase (EC 1.1.1.37) (Fragment)	528		36 / 5.9	21 / 54	24.3 / 6.6	T. aestivum	Q6XEB8_WHEAT	
104	Wheat EST match	36	Amylogenin	38 / 5.85	4 / 13	28 / 9.79	T. aestivum	Ta2337253412-1	GI:4158230
105	Wheat EST match	67	Putative malate dehydrogenase	34 / 6.25	6 / 8	52.4 / 9.39	O. sativa	Ta2794-3	GI:50932771
106	Disease resistance protein (Fragment)	27		36 / 6.3	7 / 30	20.6 / 9.2	T. aestivum	Q3HNP9_WHEAT	
107	Cytosolic glyceraldehyde-3-phosphate dehydrogenase GAPDH (Fragment)	87		37 / 6.4	8 / 40	25.3 / 7.83	T. aestivum	Q9M4V4_WHEAT	
108	Disease resistance protein (Fragment)	35		35 / 6.7	8 / 35	20.6 / 9.2	T. aestivum	Q3HNP9_WHEAT	
109	7S Globulin storage protein	47		32 / 6.8	13 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
110	DNA-binding protein (Fragment)	24		35 / 5.1	5 / 19	20.4 / 9.15	T. aestivum	Q41541_WHEAT	
111	Putative 60S ribosomal protein L36	32		34 / 5.15	9 / 47	12.4 / 11.43	O. sativa	Q8L5X0_ORYSA	
112	No match								
113	Wheat EST match	123	7S Globulin storage protein	33 / 5.45	24 / 27	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
114	No match								
115	Wheat EST match	79	7S Globulin storage protein	32 / 5.45	30 / 35	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
116	Wheat EST match	47	7S Globulin storage protein	32 / 5.65	11 / 15	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
117	No match								
118	No match								
119	7S Globulin storage protein	125		33 / 5.85	7 / 5	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
120	7S Globulin storage protein	46		32 / 6.0	6 / 5	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
121	Cytosolic glyceraldehyde-3-phosphate dehydrogenase (Fragment)	50		33 / 6.15	5 / 15	18.2 / 6.34	T. aestivum	Q7XJJ1_WHEAT	
122	1-Cys-peroxiredoxine	22		32 / 6.25	7 / 28	23.95 / 6.08	T. aestivum	Q6W8Q2_WHEAT	

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
123	Wheat EST match	318	Glucose and ribitol dehydrogenase homolog - barley	31 / 6.55	25 / 37	48.3 / 8.17	H. vulgare	Ta631253406-3	GI:7431022
124	Wheat EST match	80	HSP70	28 / 4.8	11 / 21	40.4 / 4.92	T. aestivum	Ta236253405-2	GI:2827002
125	No match								
126	Wheat EST match	117	7S Globulin storage protein	29 / 5.3	11 / 15	60.7 / 8.24	T. aestivum	Ta2606253405-2	GI:170696
127	No match								
128	Wheat EST match	278	Putative glyoxalase I	31 / 5.5	22 / 30	46.6 / 8.19	T. aestivum	Ta52588-3	GI:7619802
129	Wheat EST match	203	7S Globulin storage protein	31 / 5.5	14 / 16	60.7 / 8.24	T. aestivum	Ta2606253405-2	GI:170696
130	No match								
131	Wheat EST match	390	7S Globulin storage protein	32 / 5.7	18 / 26	61.7 / 8.08	T. aestivum	Ta3078253410-1	GI:170696
132	Wheat EST match	184	7S Globulin storage protein	29 / 5.65	22 / 25	66.9 / 7.64	T. aestivum	Ta2606253404-2	GI:170696
133	Wheat EST match	579	7S Globulin storage protein	30 / 5.8	25 / 31	61.7 / 8.08	T. aestivum	Ta3078253410-1	GI:170696
134	Wheat EST match	117	7S Globulin storage protein	29 / 5.75	15 / 22	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
135	Wheat EST match	456	7S Globulin storage protein	29 / 5.9	19 / 19	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
136	7S Globulin storage protein	74		30 / 5.9	5 / 6	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
137	7S Globulin storage protein	47		30 / 5.95	6 / 6	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
138	Wheat EST match	641	78 Globulin storage protein	32 / 6.0	24 / 31	61.7 / 8.08	T. aestivum	Ta3078253410-1	GI:170696
139	7S Globulin storage protein	86		29 / 6.1	7 / 10	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
140	Wheat EST match	572	78 Globulin storage protein	28 / 6.1	23 / 30	61.7 / 8.08	T. aestivum	Ta3078253410-1	GI:170696
141	7S Globulin storage protein	65		29 / 6.25	6 / 5	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
142	Wheat EST match	448	Glucose and ribitol dehydrogenase homolog - barley	32 / 6.55	26 / 34	48.3 / 8.17	H. vulgare	Ta631253406-3	GI:7431022
143	No match								
144	Aldose reductase-related protein (EC 1.1.1.21)	201		31 / 6.9	14 / 31	35.8 / 6.28	A. fatua	Q43320_AVEFA	
145	Translationally controlled tumor protein	229		23 / 4.65	10 / 33	18.8 / 4.55	T. aestivum	Q8LRM8_WHEAT	
146	Aspartic proteinase	269		27 / 4.8	14 / 21	54.3 / 5.14	T. aestivum	Q401N7_WHEAT	
147	Aspartic proteinase	360		27 / 4.9	12 / 16	54.3 / 5.14	T. aestivum	Q401N7_WHEAT	
148	Heat shock protein HSP26	274		23 / 5.2	19 / 53	26.6 / 7.88	T. aestivum	Q9SBB6_WHEAT	
	Small heat shock protein Hsp23.6 precursor	19		22 / 5.25	6 / 15	23.6 / 5.25	T. aestivum	Q9ZP24_WHEAT	
150	Heat shock protein HSP26	56		24 / 5.35	4 / 15	26.5 / 9.36	T. aestivum	Q9ZSR6_WHEAT	

Table A-2.5 (continued)

Table A-2.5	(continued)
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	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
151	Triosephosphate-isomerase	284		25 / 5.6	15 / 44	26.8 / 5.38	T. aestivum	Q9FS79_WHEAT	
152	Wheat EST match	68	Cupin family protein, expressed	25 / 5.6	6 / 30	17.95 / 5.6	O. sativa	Ta132809-2	GI:108706671
153	Wheat EST match	143	7S Globulin storage protein	26.5 / 5.65	12 / 14	60.7 / 8.24	T. aestivum	Ta2606253405-2	GI:170696
154	Wheat EST match	68	Cupin family protein, expressed	25 / 5.8	8 / 55	17.95 / 5.6	O. sativa	Ta132809-2	GI:108706671
155	Wheat EST match	192	7S Globulin storage protein	27 / 5.8	10 / 11	60.7 / 8.24	T. aestivum	Ta2606253405-2	GI:170696
156	No match								
157	Wheat EST match	101	Cupin family protein, expressed	25 / 5.75	11 / 64	17.95 / 5.6	O. sativa	Ta132809-2	GI:108706671
158	Putative proteasome subunit alpha type 3	179		26 / 5.8	12 / 28	27.2 / 5.76	O. sativa	Q6F321_ORYSA	
159	Wheat EST match	45	7S Globulin storage protein	25 / 6.1	10 / 15	66.9 / 7.64	T. aestivum	Ta2606253404-2	GI:170696
160	Glutathione transferase (EC 2.5.1.18)	403		23 / 6.35	20 / 42	24.98 / 6.35	T. aestivum	Q8RW04_WHEAT	
161	Glutathione transferase (EC 2.5.1.18)	42		23 / 6.45	16 / 39	25.0 / 6.35	T. aestivum	Q8RW04_WHEAT	
162	Wheat EST match	99	7S Globulin storage protein	25 / 6.55	11 / 16	60.7 / 8.24	T. aestivum	Ta2606253405-2	GI:170696
163	Wheat EST match	348	7S Globulin storage protein	24 / 6.55	14 / 18	60.7 / 8.24	T. aestivum	Ta2606253405-2	GI:170696
164	Putative alpha 1 subunit of 20S proteasome	230		25 / 6.65	14 / 31	27.6 / 6.19	O. sativa	Q7G665_ORYSA	
165	No match								
166	Small heat shock protein Hsp23.5 precursor	73		17 / 5.2	7 / 27	23.4 / 6.22	T. aestivum	Q9ZP25_WHEAT	
167	Heat shock protein HSP26	55		18 / 5.4	17 / 58	26.6 / 7.88	T. aestivum	Q9ZSR5_WHEAT	
168	No match								
169	Wheat EST match	207	Unnamed protein product	23 / 5.65	37 / 54	36.9 / 9.06	T. aestivum	Ta13211-3	GI:21813
170	Heat shock protein HSP26	33		20 / 5.75	14 / 41	26.5 / 9.36	T. aestivum	Q9ZSR6_WHEAT	
171	Wheat EST match	122	Cupin family protein, expressed	23.5 / 5.8	6 / 31	17.95 / 5.6	O. sativa	Ta132809-2	GI:108706671
172	Wheat EST match	158	Unnamed protein product	23 / 5.95	16/32	27.9 / 7.27	T. aestivum	Ta13210-3	GI:21813
173	Wheat EST match	83	7S Globulin storage protein	17 / 6.7	24 / 21	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
174	Wheat EST match	238	Unnamed protein product	19 / 6.75	26 / 44	36.9 / 9.06	T. aestivum	Ta13211-3	GI:21813
175	Endogenous alpha-amylase/subtilisin inhibitor (WASI)	327		19 / 6.85	17 / 60	19.6 / 6.77	T. aestivum	IAAS_WHEAT	
176	Wheat EST match	363	7S Globulin storage protein	15.5 / 4.95	16 / 16	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696

Spot	Protein name	Mascot	Protein name	Observed	Matching	Theoretical <sup>3</sup>	Species with	Swiss-Prot accession	GenBank
No.	(Mascot cereal search)	or GPM <sup>1</sup> score	(Wheat EST BLAST search) <sup>2</sup>	MW (KDa)/pI	peptides / %Coverage	MW (Da)/pI	homologous protein	number <sup>4</sup>	accession number
177	Wheat EST match	387	7S Globulin storage protein	15 / 5	17 / 17	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
178	Wheat EST match	347	78 Globulin storage protein	5.2 / 5.1	14 / 11	78.9 / 8.75	T. aestivum	Ta2154253404-2	GI:170696
179	Wheat EST match	498	78 Globulin storage protein	5.1 / 5.25	18 / 17	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
180	Wheat EST match	148	78 Globulin storage protein	16 / 5.25	11 / 32	36.9 / 9.58	T. aestivum	Ta3079253404-3	GI:170696
181	Wheat EST match	638	78 Globulin storage protein	15 / 5.35	21 / 22	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
182	Wheat EST match	571	78 Globulin storage protein	15 / 5.4	14 / 16	61.7 / 8.08	T. aestivum	Ta3078253410-1	GI:170696
183	Wheat EST match	544	78 Globulin storage protein	15 / 5.45	23 / 25	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
184	Wheat EST match	600	78 Globulin storage protein	15 / 5.55	14 / 16	61.7 / 8.08	T. aestivum	Ta3078253410-1	GI:170696
185	Wheat EST match	227	78 Globulin storage protein	15.5 / 5.6	15 / 17	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
186	Wheat EST match	248	Heat shock protein 16.9B	15.5 / 5.75	13 / 27	31.2 / 7.76	T. aestivum	Ta238116-2	GI:21805
187	Wheat EST match	318	78 Globulin storage protein	16 / 5.7	11 / 31	23.1 / 5.3	T. aestivum	Ta9905-2	GI:170696
188	No match								
189	Wheat EST match	502	78 Globulin storage protein	15 / 5.8	20 / 27	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
190	Wheat EST match	359	78 Globulin storage protein	15 / 5.9	13 / 14	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
191	Wheat EST match	85	OSJNBa0027P08.9	16 / 5.9	15 / 22	36.9 / 9.06	O. sativa	Ta13211-3	GI:50924568
	16.9 kDa class I heat shock protein (Low molecular weight heat shock protein)	76		15 / 6.15	6 / 37	16.9 / 5.83	T. aestivum	HSP11_WHEAT	
193	7S Globulin storage protein	79		17 / 6.2	8 / 8	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
194	Wheat EST match	419	78 Globulin storage protein	15 / 6.25	15 / 20	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
	Heat shock protein 17.9. Pennisetum americanum	38		16 / 6.45	4 / 13	17.9 / 5.82	P. glaucum	Q40867_PENAM	
196	Wheat EST match	131	7S Globulin storage protein	16 / 6.6	13 / 35	20.5 / 8.42	T. aestivum	Ta194625-1	GI:170696
197	OJ991113_30.23 protein	28		15 / 6.9	3 / 60	4.8 / 4.53	O. sativa	Q7XSA9_ORYSA	
198	Wheat EST match	30	Putative ribophorin I homologue	13.5 / 4.6	8 / 15	21.7 / 9.4	H. vulgare	Ta127531-2	GI:2894378
199	Wheat EST match	125	7S Globulin storage protein	14.5 / 4.8	10 / 28	36.9 / 9.58	T. aestivum	Ta3079253404-3	GI:170696
200	Wheat EST match	317	7S Globulin storage protein	14 / 4.85	13 / 35	36.9 / 9.58	T. aestivum	Ta3079253404-3	GI:170696
201	Wheat EST match	259	7S Globulin storage protein	13.5 / 4.95	14 / 15	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
202	Wheat EST match	447	78 Globulin storage protein	14 / 5.0	16 / 15	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
203	Wheat EST match	436	7S Globulin storage protein	15.5 / 5.05	17 / 21	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
204	Wheat EST match	355	7S Globulin storage protein	15 / 5.15	15 / 17	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
205	Wheat EST match	369	7S Globulin storage protein	15 / 5.2	16/16	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696

Table A-2.5 (continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
206	Wheat EST match	478	7S Globuliin storage protein	15 / 5.35	16 / 17	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
207	Wheat EST match	561	7S Globuliin storage protein	15 / 5.3	23 / 25	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
208	Wheat EST match	357	7S Globuliin storage protein	15 / 5.15	13 / 14	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
209	16.9 kDa class I heat shock protein	181		15 / 5.7	11 / 47	16.9 / 5.83	T. aestivum	HSP11_WHEAT	
210	No match								
211	Wheat EST match	555	7S Globuliin storage protein	14 / 6.1	17 / 17	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
212	Wheat EST match	488	7S Globuliin storage protein	14 / 6.2	13 / 13	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
213	7S Globuliin storage protein	41		14 / 6.5	28 / 29	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
214	Wheat EST match	556	7S Globuliin storage protein	14 / 6.7	17 / 17	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
215	Wheat EST match	443	7S Globuliin storage protein	15 / 6.9	15 / 17	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
216	Wheat EST match	197	7S Globuliin storage protein	12.5 / 4.9	10 / 13	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
217	Wheat EST match	176	7S Globuliin storage protein	13 / 4.9	12 / 19	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
218	Wheat EST match	405	7S Globuliin storage protein	14 / 14.95	15 / 18	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
219	Wheat EST match	212	7S Globuliin storage protein	12 / 4.95	10 / 13	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
220	Wheat EST match	91	7S Globuliin storage protein	13 / 5.2	13 / 14	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
221	Wheat EST match	421	7S Globuliin storage protein	14 / 5.2	18 / 19	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
222	Wheat EST match	489	7S Globuliin storage protein	14.95 / 5.2	15 / 30	36.9 / 9.58	T. aestivum	Ta3079253404-3	GI:170696
223	Wheat EST match	376	7S Globuliin storage protein	14 / 5.3	16 / 17	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
224	Wheat EST match	338	7S Globuliin storage protein	14 / 5.35	19 / 21	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
225	Wheat EST match	438	7S Globuliin storage protein	14 / 5.3	18 / 21	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
226	Wheat EST match	332	7S Globuliin storage protein	14 / 5.5	16 / 17	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
227	Wheat EST match	280	Glyoxalase family protein, expressed	12 / 5.55	13 / 26	31.3 / 9.25	O. sativa	Ta886253410-2	GI:108707474
228	Wheat EST match	52	7S Globuliin storage protein	11 / 5.2	6 / 11	52.8 / 9.69	T. aestivum	Ta627253413-2	GI:170696
229	Wheat EST match	518	7S Globuliin storage protein	13 / 5.75	18 / 17	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
230	ABA inducible protein	19		12 / 5.9	6 / 27	17.5 / 5.95	T. aestivum	Q7XAP5_WHEAT	
231	Wheat EST match	120	7S Globuliin storage protein	11 / 5.9	9 / 11	52.8 / 9.69	T. aestivum	Ta627253413-2	GI:170696
232	Wheat EST match	138	Glyoxalase family protein, expressed	12 / 6.35	9 / 18	31.3 / 9.25	O. sativa	Ta886253410-2	GI:108707474
233	Wheat EST match	213	Nucleoside diphosphate kinase 1, putative, expressed	13 / 6.45	10 / 25	29.3 / 9.51	O. sativa	Ta14950-3	GI:31433540

Table A-2.5 (continued)

Spot Protein name No. (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
234 Wheat EST match	50	7S Globulin storage protein	13 / 6.6	13 / 16	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
235 PR-4 (Fragment)	47		12 / 6.8	2 / 13	13.1 / 6.28	T. aestivum	Q9SQG4_WHEAT	
236 Transposase (Fragment)	24		12.5 / 6.9	8 / 29	14.6 / 9.48	T. aestivum	Q8W1P3_WHEAT	
237 Wheat EST match	80	7S Globulin storage protein	11 / 6.9	6 / 6	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696

Table A-2.5 (continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
1	Wheat EST match	32	7S Globulin storage protein	78 / 6.7	28 / 39	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
2	7S Globulin storage protein	62		110 / 6.7	19 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
3	7S Globulin storage protein	48		75 / 6.7	16 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
4	Wheat EST match	98	7S Globulin storage protein	78 / 6.8	31 / 42	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
5	7S Globulin storage protein	69		75 / 6.9	25 / 28	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
6	Wheat EST match	140	7S Globulin storage protein	78 / 7.1	28 / 36	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
7	Wheat EST match	49	7S Globulin storage protein	78 / 7.2	33 / 45	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
8	7S Globulin storage protein	43		73 / 6.9	26 / 36	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
9	7S Globulin storage protein	37		75 / 6.9	18 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
10	7S Globulin storage protein	113		74 / 6.9	26 / 25	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
11	7S Globulin storage protein	80		75 / 7.1	23 / 27	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
12	Wheat EST match	104	7S Globulin storage protein	74 / 7.4	35 / 41	78.9 / 8.75	T. aestivum	Ta2154253404-2	GI:170696
13	7S Globulin storage protein	50		72 / 6.9	30 / 36	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
14	Wheat EST match	187	7S Globulin storage protein	72 / 6.9	36 / 49	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
15	Wheat EST match	162	7S Globulin storage protein	74 / 6.95	33 / 44	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
16	7S Globulin storage protein	56		73 / 6.95	23 / 33	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
17	7S Globulin storage protein	63		73 / 6.9	31 / 29	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
18	7S Globulin storage protein	67		74 / 6.95	28 / 29	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
19	7S Globulin storage protein	120		74 / 7.0	41 / 38	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
20	7S Globulin storage protein	98		74 / 7.15	28 / 34	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
21	7S Globulin storage protein	95		74 / 7.2	23 / 25	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
22	7S Globulin storage protein	117		75 / 7.2	30 / 39	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
23	7S Globulin storage protein	114		74 / 7.25	33 / 30	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
24	7S Globulin storage protein	102		75 / 7.55	21 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
25	7S Globulin storage protein	49		75 / 7.4	15 / 18	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
26	7S Globulin storage protein	88		73 / 7.55	25 / 26	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
27	7S Globulin storage protein	89		73 / 7.65	20 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
28	7S Globulin storage protein	77		73 / 7.8	20 / 22	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
29	Wheat EST match	135	7S Globulin storage protein	73 / 7.85	38 / 40	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696

 Table A-2.6
 Inner bran fraction (aleurone cells) proteins identified from 2D gel pH 6-11

Table A-2.6	(continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
30	Cytosolic NADP malic enzyme	74		69 / 6.1	27 / 32	64.2 / 6.5	T. aestivum	Q5JKW5_ORYSA	
31	Wheat EST match	50	Cytosolic NADP malic enzyme	71 / 6.15	18 / 54	25.8 / 8.9	O. sativa	Ta965253413-3	GI:38261493
32	Wheat EST match	169	7S Globulin storage protein	72 / 6.45	30 / 40	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
33	7S Globulin storage protein	64		72 / 6.6	26 / 31	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
34	7S Globulin storage protein	84		72 / 6.7	28 / 28	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
35	Wheat EST match	209	7S Globulin storage protein	72 / 6.75	36 / 47	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
36	7S Globulin storage protein	86		73 / 6.95	32 / 36	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
37	7S Globulin storage protein	64		72 / 6.95	27 / 29	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
38	7S Globulin storage protein	34		72 / 7.0	26 / 36	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
39	7S Globulin storage protein	122		73 / 7.05	24 / 26	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
40	7S Globulin storage protein	135		73 / 7.15	32 / 36	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
41	7S Globulin storage protein	72		72 / 7.2	31 / 29	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
42	7S Globulin storage protein	108		74 / 7.35	27 / 22	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
43	7S Globulin storage protein	102		73 / 7.4	27 / 29	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
44	7S Globulin storage protein	109		74 / 7.5	30 / 40	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
45	7S Globulin storage protein	89		73 / 7.6	25 / 28	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
46	7S Globulin storage protein	80		73 / 7.7	20 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
47	Hypothetical protein P0431E05.13	38		73 / 7.9	10 / 53	16.5 / 11.34	O. sativa	Q653N9_ORYSA	
48	7S Globulin storage protein	119		72 / 8.1	11 / 7	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
49	7S Globulin storage protein	157		72 / 8.2	17 / 15	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
50	7S Globulin storage protein	114		72 / 7.9	14 / 15	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
51	7S Globulin storage protein	76		72 / 8.3	12 / 10	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
52	7S Globulin storage protein	57		72 / 8.4	12 / 11	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
53	7S Globulin storage protein	69		72 / 8.4	12 / 10	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
54	7S Globulin storage protein	34		72 / 8.8	14 / 12	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
55	7S Globulin storage protein	172		75 / 9.5	14 / 12	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
56	7S Globulin storage protein	47		58 / 6.2	25 / 25	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
57	Putative dihydrolipoamide dehydrogenase	73		58 / 6.3	25 / 44	52.6 / 7.21	O. sativa	Q9ASP4_ORYSA	

Table A-2.6	(continued)
Table A-2.0	(continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
58	7S Globuliin storage protein	44		71 / 6.7	22 / 28	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
59	Wheat EST match	38	Unknown protein	70 / 6.8	14 / 39	23.6 / 9.18	O. sativa	Ta1159253408-3	GI:55769693
60	7S Globuliin storage protein	84		58 / 6.9	27 / 28	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
61	7S Globuliin storage protein	47		69 / 6.9	30 / 33	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
62	7S Globuliin storage protein	73		61 / 7.1	28 / 31	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
63	7S Globuliin storage protein	79		71 / 6.95	31 / 32	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
64	Wheat EST match	126	7S Globuliin storage protein	70 / 7.0	34 / 45	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
65	Wheat EST match	90	7S Globuliin storage protein	71 / 7.05	36 / 40	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
66	7S Globuliin storage protein	89		71 / 7.0	32 / 33	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
67	7S Globuliin storage protein	78		60 / 7.0	29 / 38	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
68	7S Globuliin storage protein	91		69 / 7.0	29/31	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
69	Wheat EST match	69	7S Globuliin storage protein	67 / 7.3	20/30	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
70	7S Globuliin storage protein	106		68 / 7.15	32 / 32	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
71	7S Globuliin storage protein	104		67 / 7.3	24 / 29	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
72	7S Globuliin storage protein	115		67 / 7.6	25 / 29	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
73	Putative globulin	58		68 / 7.65	9 / 13	52.1 / 6.78	T. aestivum	Q852L2_ORYSA	
74	Wheat EST match	29	7S Globuliin storage protein	56 / 6.65	29 / 18	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
75	Wheat EST match	57	7S Globuliin storage protein	56 / 6.85	32 / 22	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
76	Wheat EST match	31	7S Globuliin storage protein	57 / 6.9	29 / 40	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
77	Wheat EST match	45	7S Globuliin storage protein	60 / 7.1	25 / 29	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
78	7S Globuliin storage protein	87		58 / 7.15	26 / 36	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
79	Wheat EST match	62	7S Globuliin storage protein	60 / 7.2	30 / 32	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
80	7S Globuliin storage protein	51		67 / 7.25	23 / 26	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
81	7S Globuliin storage protein	113		59 / 7.25	24 / 26	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
82	7S Globuliin storage protein	128		50 / 7.35	34 / 36	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
83	7S Globuliin storage protein	46		59 / 7.35	23 / 32	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
84	7S Globuliin storage protein	97		59 / 7.5	29 / 34	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
85	7S Globuliin storage protein	95		67 / 7.4	22 / 25	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
86	7S Globuliin storage protein	36		70 / 7.5	14 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	

Table A-2.6	(continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
87	7S Globuliin storage protein	105		67 / 7.4	29 / 33	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
88	7S Globuliin storage protein	28		55 / 7.6	15 / 24	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
89	7S Globuliin storage protein	83		66 / 7.8	22 / 22	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
90	Putative cell death associated protein	35		59 / 7.8	11 / 34	36 / 5.26	O. sativa	Q8GSJ3_ORYSA	
91	7S Globuliin storage protein	108		67 / 7.8	18 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
92	7S Globuliin storage protein	104		62 / 7.9	23 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
93	Globulin-like protein	36		55 / 8.0	5 / 10	52 / 6.78	O. sativa	Q8L8I0_ORYSA	
94	7S Globuliin storage protein	39		72 / 8.0	11 / 12	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
95	7S Globuliin storage protein	72		70 / 8.2	13 / 11	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
96	Wheat EST match	41	Alanine aminotransferase	50 / 6.2	18 / 28	47.1 / 9.35	H. vulgare	Ta209289-1	GI:469148
97	Wheat EST match	99	7S Globuliin storage protein	52 / 6.4	28 / 36	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
98	7S Globuliin storage protein	52		53 / 6.6	28 / 32	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
99	7S Globuliin storage protein	70		53 / 6.5	21 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
100	Wheat EST match	30	7S Globuliin storage protein	53 / 6.9	22 / 56	41.3 / 9.42	T. aestivum	Ta3078253409-1	GI:170696
101	7S Globuliin storage protein	71		53 / 7.0	21 / 24	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
102	Wheat EST match	41	7S Globuliin storage protein	56 / 7.0	21 / 26	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
103	7S Globuliin storage protein	64		52 / 7.1	23 / 25	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
104	7S Globuliin storage protein	47		50 / 6.9	22 / 31	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
105	Globulin-like protein	37		57 / 7.1	13 / 21	52.0 / 6.78	O. sativa	Q8L8I0_ORYSA	
106	7S Globuliin storage protein	62		50 / 7.2	18 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
107	Wheat EST match	141	7S Globuliin storage protein	50 / 7.35	21 / 33	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
108	7S Globuliin storage protein	116		50 / 7.2	21 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
109	Globulin-like protein	51		48 / 7.2	5 / 8	52 / 6.78	O. sativa	Q8L8I0_ORYSA	
110	Wheat EST match	45	7S Globuliin storage protein	56 / 7.15	35 / 34	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
111	Wheat EST match	32	7S Globuliin storage protein	51 / 7.25	33 / 40	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
112	Wheat EST match	35	7S Globuliin storage protein	49 / 7.25	22 / 47	41.3 / 9.42	T. aestivum	Ta3078253409-1	GI:170696
113	Wheat EST match	29	7S Globuliin storage protein	49 / 7.4	20 / 47	41.3 / 9.42	T. aestivum	Ta3078253409-1	GI:170696
114	7S Globuliin storage protein	141		50 / 7.5	17 / 17	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
115	7S Globuliin storage protein	32		72 / 7.5	13 / 15	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
116	7S Globuliin storage protein	122		55 / 7.7	21 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
117	7S Globuliin storage protein	112		50 / 7.8	12 / 8	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
118	7S Globuliin storage protein	97		52 / 7.9	24 / 28	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
119	7S Globuliin storage protein	166		50 / 7.9	18 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
120	7S Globuliin storage protein	151		49 / 8.0	24 / 25	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
121	7S Globuliin storage protein	131		50 / 8.2	19 / 18	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
122	7S Globuliin storage protein	68		50 / 8.3	14 / 12	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
123	7S Globuliin storage protein	145		50 / 8.1	18 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
124	7S Globuliin storage protein	135		50 / 8.2	17 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
125	7S Globuliin storage protein	80		50 / 8.3	10 / 7	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
126	7S Globuliin storage protein	164		50 / 8.4	20 / 20	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
127	7S Globuliin storage protein	170		50 / 8.5	20 / 17	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
128	7S Globuliin storage protein	157		51 / 8.6	16 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
129	7S Globuliin storage protein	190		50 / 8.6	22 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
130	7S Globuliin storage protein	147		49 / 8.7	18 / 15	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
131	7S Globuliin storage protein	155		50 / 8.8	19 / 24	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
132	7S Globuliin storage protein	138		50 / 8.9	17 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
133	7S Globuliin storage protein	162		50 / 9.0	17 / 18	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
134	7S Globuliin storage protein	160		50 / 9.1	17 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
135	7S Globuliin storage protein	156		50 / 9.2	13 / 12	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
136	7S Globuliin storage protein	165		50 / 9.1	16 / 20	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
137	Elongation factor 1-alpha 1-alpha)	(EF- 41		50 / 9.3	9 / 14	49.1 / 9.2	T. aestivum	EF1A_WHEAT	
138	Elongation factor 1-alpha alpha)	(EF-1- 74		50 / 9.4	10 / 18	49.1 / 9.2	T. aestivum	EF1A_WHEAT	
139	Elongation factor 1-alpha alpha)	(EF-1- 60		50 / 9.5	11 / 23	49.1 / 9.2	T. aestivum	EF1A_WHEAT	
140	Formate dehydrogenase, mitochor precursor (EC 1.2.1.2)	ndrial 93		45 / 6.4	15 / 30	41.5 / 6.9	H. vulgare	FDH_HORVU	

Table A-2.6 (continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
141	Formate dehydrogenase (Fragment)	165		45 / 6.5	14 / 48	28.99 / 8.61	T. aestivum	Q7X9L3_WHEAT	
142	Citrate synthase, eukaryotic	43		47 / 6.6	13 / 27	52.3 / 7.71	O. sativa	Q2R339_ORYSA	
143	Wheat EST match	53	7S Globuliin storage protein	45 / 6.6	35 / 43	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
144	Wheat EST match	48	7S Globuliin storage protein	42 / 6.9	21 / 26	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
145	Glyceraldehyde-3-phosphate dehydrogenase, cytosolic (EC 1.2.1.12)	51		47 / 7.1	19 / 39	36.5 / 6.67	H. vulgare	G3PX_HORVU	
146	7S Globuliin storage protein	103		50 / 7.1	28 / 35	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
147	Wheat EST match	150	78 Globuliin storage protein	48 / 7.1	17 / 29	61.7 / 8.08	T. aestivum	Ta3078253410-1	GI:170696
148	7S Globuliin storage protein	72		50 / 7.25	24 / 30	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
149	7S Globuliin storage protein	62		51 / 7.4	12 / 14	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
150	7S Globuliin storage protein	94		50 / 7.3	23 / 28	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
151	7S Globuliin storage protein	84		50 / 7.4	25 / 26	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
152	7S Globuliin storage protein	136		47 / 7.5	18 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
153	7S Globuliin storage protein	115		50 / 7.6	21 / 24	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
154	7S Globuliin storage protein	79		50 / 7.7	21 / 25	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
155	7S Globuliin storage protein	106		45 / 7.6	14 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
156	7S Globuliin storage protein	141		50 / 7.8	18 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
157	7S Globuliin storage protein	102		47 / 7.9	12 / 7	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
158	7S Globuliin storage protein	112		49 / 7.8	14 / 13	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
159	7S Globuliin storage protein	89		47 / 7.9	14 / 13	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
160	7S Globuliin storage protein	125		50 / 7.7	15 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
161	Transposase	23		50 / 8	5 / 75	5.58 / 12.19	X. oryzae	Q5GYA8_XANOR	
162	Wheat EST match	112	78 Globuliin storage protein	45 / 8.0	17 / 27	61.7 / 8.08	T. aestivum	Ta3078253410-1	GI:170696
163	7S Globuliin storage protein	148		50 / 8.1	18 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
164	7S Globuliin storage protein	157		47 / 8.2	17 / 14	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
165	7S Globuliin storage protein	153		47 / 8.3	15 / 13	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
166	7S Globuliin storage protein	120		47 / 8.5	13 / 11	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
167	7S Globuliin storage protein	178		47 / 8.5	18 / 15	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
168	7S Globulin storage protein	167		47 / 8.5	16 / 12	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
169	7S Globulin storage protein	173		47 / 8.6	16 / 15	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
170	7S Globulin storage protein	128		47 / 8.8	15 / 12	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
171	7S Globulin storage protein	128		47 / 9.2	18 / 17	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
172	7S Globulin storage protein	87		47 / 9.1	21 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
173	7S Globulin storage protein	122		47 / 9.3	18 / 18	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
174	7S Globulin storage protein	143		47 / 9.4	16 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
175	7S Globulin storage protein	101		46 / 9.6	12 / 11	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
176	7S Globulin storage protein	113		47 / 9.5	11 / 8	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
177	7S Globulin storage protein	138		46 / 9.6	20 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
178	7S Globulin storage protein	95		44 / 9.6	12 / 14	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
179	Cytoplasmic aldolase	31		43 / 6.1	20 / 36	38.7 / 6.56	O. sativa	Q40676_ORYSA	
180	Enolase (EC 4.2.1.11)	86		44 / 6.2	26 / 48	47.9 / 5.41	O. sativa	ENO_ORYSA	
181	Aspartate aminotransferase precurso (EC 2.6.1.1)	r 29		43 / 6.2	11 / 16	47.6 / 6.71	P. miliaceum	Q43305_PANMI	
182	Glyceraldehyde-3-phosphate dehydrogenase, cytosolic (E0 1.2.1.12)	47 C		42 / 6.3	15 / 36	36.5 / 6.67	H. vulgare	G3PX_HORVU	
183	Glyceraldehyde-3-phosphate dehydrogenase, cytosolic (EC 1.2.1.12)	482		42 / 6.5	24 / 43	36.5 / 6.67	H. vulgare	G3PX_HORVU	
184	Putative fructose 1-,6-biphosphate aldolase (Fragment)	44		38 / 6.5	5 / 19	29.1 / 8.71	T. aestivum	Q8VWM9_WHEAT	
185	Glyceraldehyde-3-phosphate dehydrogenase, cytosolic (E 1.2.1.12)	157 C		42 / 6.8	22 / 39	36.5 / 6.67	H. vulgare	G3PX_HORVU	
186	Wheat EST match	88	7S Globulin storage protein	42 / 6.8	9 / 9	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
187	Wheat EST match	50	7S Globulin storage protein	42 / 7.1	14 / 56	23.6 / 9.99	T. aestivum	Ta136340-2	GI:170696
188	Wheat EST match	203	78 Globulin storage protein	43 / 7.2	19 / 25	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696

 Table A-2.6 (continued)

**189** Cytosolic glyceraldehyde-3-phosphate

dehydrogenase GAPDH (Fragment)

191

42 / 7.2

8 / 29

25.3 / 7.83

T. aestivum

Q9M4V4\_WHEAT

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
190	Wheat EST match	87	78 Globuliin storage protein	42 / 7.25	15 / 14	78.9 / 8.75	T. aestivum	Ta2154253404-2	GI:170696
191	Wheat EST match	53	7S Globuliin storage protein	42 / 7.3	14 / 68	23.6 / 9.99	T. aestivum	Ta136340-2	GI:170696
192	Wheat EST match	158	7S Globuliin storage protein	48 / 7.5	19 / 19	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
193	Wheat EST match	25	RNA recognition motif family protein, expressed	42 / 7.45	10 / 40	26.5 / 10.010	O. sativa	Ta212753-3	GI:108707669
194	Wheat EST match	166	7S Globuliin storage protein	43 / 7.5	22 / 23	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
195	Wheat EST match	101	7S Globuliin storage protein	43 / 7.7	13 / 14	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
196	Wheat EST match	50	7S Globuliin storage protein	42 / 7.9	9 / 30	20.5 / 8.42	T. aestivum	Ta194625-1	GI:170696
197	Wheat EST match	65	7S Globuliin storage protein	42 / 7.9	8 / 18	41.3 / 9.42	T. aestivum	Ta3078253409-1	GI:170696
198	Xylanase inhibitor (Fragment)	84		42 / 8.1	8 / 19	40.2 / 8.41	T. aestivum	Q53IQ4_WHEAT	
199	7S Globuliin storage protein	92		42 / 8.3	15 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
200	Wheat EST match	121	7S Globuliin storage protein	42 / 8.6	30 / 33	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696
201	Xylanase inhibitor precursor (Xylanase inhibitor TAXI-I)	48		42 / 8.6	4 / 6	40.9 / 8.18	T. aestivum	Q8H0K8_WHEAT	
202	Wheat EST match	91	7S Globuliin storage protein	43 / 9.1	11/9	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
203	7S Globuliin storage protein	103		43 / 9.2	11 / 9	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
204	Malate dehydrogenase	69		36 / 6.1	18 / 29	35.4 / 8.74	O. sativa	Q7FSL4_ORYSA	
205	Wheat EST match	288	O-methyltransferase	37 / 6.15	27 / 37	51.9 / 7.64	T. aestivum	Ta94253412-2	GI:4098238
206	Putative fructose 1-,6-biphosphate aldolase (Fragment)	76		37 / 6.4	8 / 33	29.1 / 8.71	T. aestivum	Q8VWM9_WHEAT	
207	Cytosolic glyceraldehyde-3-phosphate dehydrogenase (Fragment)	105		37 / 6.4	13 / 58	18.2 / 6.34	T. aestivum	Q7XJJ1_WHEAT	
208	Wheat EST match	175	Putative malate dehydrogenase	36 / 6.45	9 / 21	52.4 / 9.39	O. sativa	Ta2794-3	GI:50932771
209	Hypothetical protein P0686H11.1 (Hypothetical protein P0605H02.41)	34		37 / 6.5	7 / 74	8.92 / 5.66	O. sativa	Q6Z8U5_ORYSA	
210	Wheat EST match	160	7S Globuliin storage protein	38 / 6.9	15 / 56	23.6 / 9.99	T. aestivum	Ta136340-2	GI:170696
211	Putative glyceraldehyde-3-phosphate dehydrogenase	27		37 / 6.5	14 / 29	36.5 / 7.68	O. sativa	Q6K5G8_ORYSA	

Table A-2.6 (continued)

Table A-2.6 (c)	ontinued)
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-	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
212	Cytosolic glyceraldehyde-3-phosphate dehydrogenase (Fragment)	28		37 / 7.1	7 / 39	18.2 / 6.34	T. aestivum	Q7XJJ1_WHEAT	
213	Wheat EST match	124	7S Globuliin storage protein	42 / 7.2	15 / 21	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
214	Wheat EST match	197	7S Globuliin storage protein	41 / 7.25	13 / 50	30.0 / 7.22	T. aestivum	Ta628253407-1	GI:170696
215	Globulin-like protein	57		41 / 7.4	5 / 9	52 / 6.78	O. sativa	Q8L8I0_ORYSA	
216	Cytosolic glyceroldehyde-3-phosphate dehydrogenase GAPC4	159		39 / 7.5	15 / 34	36.4 / 6.61	Z. mays	Q43359_MAIZE	
217	Wheat EST match	289	7S Globuliin storage protein	41 / 7.7	16 / 42	30.0 / 7.22	T. aestivum	Ta628253407-1	GI:170696
218	Wheat EST match	48	Putative S-formylglutathione hydrolase	35 / 6.2	14 / 26	41.4 / 8.35	O. sativa	Ta56703-2	GI:57900400
219	Wheat EST match	271	Glucose and ribitol dehydrogenase homolog	36 / 6.45	31 / 43	48.3 / 8.17	H. vulgare	Ta631253406-3	GI:7431022
220	Wheat EST match	273	Glucose and ribitol dehydrogenase homolog	36 / 6.3	24 / 46	48.3 / 8.17	H. vulgare	Ta631253406-3	GI:7431022
221	Stress responsive protein	28		36 / 6.5	7 / 32	22.2 / 7.64	T. aestivum	Q4U0C9_WHEAT	
222	Putative aldose reductase	68		36 / 6.7	12 / 26	35.6 / 6.32	O. sativa	Q65WW3_ORYSA	
223	Caleosin 1	34		35 / 6.5	14 / 39	34.2 / 6.11	H. vulgare	Q6UFY8_HORVU	
224	Wheat EST match	145	7S Globuliin storage protein	37 / 6.7	18 / 23	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
225	Wheat EST match	116	7S Globuliin storage protein	36 / 6.95	15 / 56	23.6 / 9.99	T. aestivum	Ta136340-2	GI:170696
226	Aldose reductase-related protein (EC 1.1.1.21)	177		36 / 6.9	12 / 27	35.8 / 6.28	A. fatua	Q43320_AVEFA	
227	Putative aldose reductase	168		36 / 7	11 / 21	35.6 / 6.32	O. sativa	Q65WW3_ORYSA	
228	Wheat EST match	91	7S Globuliin storage protein	37 / 7.1	10 / 25	41.3 / 9.42	T. aestivum	Ta3078253409-1	GI:170696
229	Wheat EST match	41	7S Globuliin storage protein	38 / 7.15	12 / 61	23.6 / 9.99	T. aestivum	Ta136340-2	GI:170696
230	7S Globuliin storage protein	40		37 / 7.3	25 / 30	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
231	Wheat EST match	29	7S Globuliin storage protein	37 / 7.3	13 / 51	23.6 / 9.99	T. aestivum	Ta136340-2	GI:170696
232	7S Globuliin storage protein	74		37 / 7.45	25 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
233	Wheat EST match	401	7S Globuliin storage protein	37 / 7.6	30 / 34	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
234	7S Globuliin storage protein	81		37 / 7.7	20 / 20	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
235	7S Globuliin storage protein	66		36 / 7.75	23 / 21	72.2 / 6.8	T. aestivum	Q7DMU0 WHEAT	

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
236	Wheat EST match	82	7S Globuliin storage protein	36 / 7.75	10 / 16	39.3 / 11.17	T. aestivum	Ta5443-3	GI:170696
237	7S Globuliin storage protein	45		36 / 8.0	18 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
238	7S Globuliin storage protein	95		36 / 8.2	21 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
239	7S Globuliin storage protein	101		37 / 8.2	20 / 22	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
240	7S Globuliin storage protein	63		36.5 / 8.4	24 / 27	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
241	7S Globuliin storage protein	94		37 / 8.5	22 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
242	7S Globuliin storage protein	112		36.5 / 8.7	20 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
243	7S Globuliin storage protein	68		37 / 8.8	18 / 18	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
244	7S Globuliin storage protein	142		37 / 8.8	20 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
245	7S Globuliin storage protein	121		37 / 9.1	19 / 20	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
246	7S Globuliin storage protein	149		36.5 / 9.2	24 / 22	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
247	7S Globuliin storage protein	156		37 / 9.2	20 / 20	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
248	7S Globuliin storage protein	144		37 / 9.3	22 / 22	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
249	7S Globuliin storage protein	148		38 / 9.4	17 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
250	7S Globuliin storage protein	81		38 / 9.4	13 / 13	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
251	7S Globuliin storage protein	153		38 / 9.5	18 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
252	7S Globuliin storage protein	107		36 / 9.5	11 / 8	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
253	7S Globuliin storage protein	134		36 / 9.8	16 / 22	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
254	Wheat EST match	97	7S Globuliin storage protein	34 / 6.2	14 / 16	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
255	7S Globuliin storage protein	58		32 / 6.2	7 / 8	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
256	Wheat EST match	316	Glucose and ribitol dehydrogenase homolog	35 / 6.65	18 / 53	29.6 / 9.51	H. vulgare	Ta631253408-2	GI:7431022
256	Wheat EST match	53	Cupin family protein, expressed	31 / 6.2	10 / 17	60.7 / 8.24	O. sativa	Ta2606253405-2	GI:108708022
257	Wheat EST match	85	Glucose and ribitol dehydrogenase homolog	35 / 6.3	28 / 38	48.3 / 8.17	H. vulgare	Ta631253406-3	GI:7431022
258	7S Globuliin storage protein	53		34 / 6.4	12 / 16	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
260	Wheat EST match	191	Glucose and ribitol dehydrogenase homolog	36 / 6.65	11 / 24	26.8 / 11.79	H. vulgare	Ta631253407-3	GI:7431022
261	Putative aldose reductase	77		35 / 6.9	7 / 16	35.6 / 6.32	O. sativa	Q65WW3_ORYSA	
262	Putative aldose reductase	117		35 / 7.1	8 / 17	35.6 / 6.32	O. sativa	Q65WW3_ORYSA	

Table A-2.6 (continued)

Table A-2.6	(continued)
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	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
263	Wheat EST match	34	Glucose and ribitol dehydrogenase homolog	33 / 7.1	14 / 31	29.6 / 9.51	H. vulgare	Ta631253408-2	GI:7431022
264	Wheat EST match	34	Putative serine/threonine protein kinase	30 / 7.15	13 / 36	41.6 / 9.98	O. sativa	Ta57592-2	GI:50916410
265	Xylanase inhibitor XIP-III	57		30 / 7.6	15 / 43	33.3 / 7.14	T. aestivum	Q4W6G2_WHEAT	
266	7S Globuliin storage protein	47		30 / 7.9	17 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
267	Lipoprotein-like	105		30 / 8.0	9 / 17	28 / 7.79	O. sativa	Q94J20_ORYSA	
268	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog)	416		30 / 8.4	22 / 43	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
269	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog)	415		28 / 8.4	23 / 56	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
270	7S Globuliin storage protein	85		36 / 8.5	21 / 24	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
271	7S Globuliin storage protein	96		36 / 8.9	19 / 20	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
272	30S ribosomal protein S2	62		29 / 8.8	10 / 27	29.5 / 8.73	X. oryzae	Q2P4A5_XANOR	
273	Class II chitinase (EC 3.2.1.14)	31		35 / 8.9	4 / 10	28.2 / 8.66	T. aestivum	Q4Z8L7_WHEAT	
274	7S Globuliin storage protein	100		36 / 9.1	18 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
275	No match								
276	7S Globuliin storage protein	141		36.5 / 9.2	24 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
277	7S Globuliin storage protein	134		36 / 9.2	18 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
278	Voltage dependent anion channel (VDAC) (Fragment)	61		35 / 9.3	16 / 59	29.3 / 9.33	T. aestivum	Q41590_WHEAT	
279	7S Globuliin storage protein	147		36.5 / 9.3	20 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
280	7S Globuliin storage protein	126		37 / 9.2	21 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
281	7S Globuliin storage protein	160		37 / 9.3	21 / 20	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
282	7S Globuliin storage protein	149		37 / 9.4	21 / 17	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
283	7S Globuliin storage protein	150		37 / 9.5	21 / 20	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
284	Caleosin 1	52		26 / 6.2	16/37	34.2 / 6.11	H. vulgare	Q6UFY8_HORVU	

Table A-2.6	(continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
285	Hypothetical protein P0686H11.1 (Hypothetical protein P0605H02.41)	27		30 / 6.3	5 / 60	8.92 / 5.66	O. sativa	Q6Z8U5_ORYSA	
286	Wheat EST match	41	Cupin family protein, expressed	26 / 6.4	6 / 30	18.9 / 10.14	O. sativa	Ta189317-3	GI:108706671
287	Putative alpha 1 subunit of 20S proteasome	50		26 / 6.5	10 / 34	27.6 / 6.19	O. sativa	Q7G665_ORYSA	
288	Putative aldose reductase	42		24 / 6.5	12 / 27	35.6 / 6.32	O. sativa	Q65WW3_ORYSA	
289	Wheat EST match	80	7S Globulin storage protein	25 / 6.5	17 / 60	23.6 / 9.99	T. aestivum	Ta136340-2	GI:170696
290	Putative aldose reductase	140		24 / 6.7	6 / 16	35.6 / 6.32	O. sativa	Q65WW3_ORYSA	
291	Putative aldose reductase	100		24 / 6.9	6 / 16	35.6 / 6.32	O. sativa	Q65WW3_ORYSA	
292	Wheat EST match	28	Squamosa promoter binding protein like	1 2-24 / 7.05	9 / 39	26.2 / 9.91	O. sativa	Ta3299253405-1	GI:53791916
293	Wheat EST match	31	Cupin family protein, expressed	23 / 7.1	16 / 22	66.9 / 7.64	O. sativa	Ta2606253404-2	GI:108708022
294	Wheat EST match	31	Annexin	24 / 7.5	8 / 28	28.7 / 9.4	T. aestivum	Ta164763-3	GI:38606205
295	Wheat EST match	226	Cupin family protein, expressed	23 / 7.5	23 / 23	66.9 / 7.64	O. sativa	Ta2606253404-2	GI:108708022
296	Class II chitinase (EC 3.2.1.14)	107		24 / 7.9	8 / 30	28.2 / 8.66	T. aestivum	Q4Z8L7_WHEAT	
297	Xylanase inhibitor protein 1 precursor (Class III chitinase homolog)	273		30 / 8.1	23 / 50	33.3 / 8.66	T. aestivum	XIP1_WHEAT; Q8L5C6	
298	No match								
299	Class II chitinase (EC 3.2.1.14)	94		25 / 8.6	9 / 37	28.2 / 8.66	T. aestivum	Q4Z8L7_WHEAT	
300	Class II chitinase (EC 3.2.1.14)	99		25 / 8.8	5 / 21	28.2 / 8.66	T. aestivum	Q4Z8L7_WHEAT	
301	26 kDa endochitinase 1 precursor (EC 3.2.1.14)	123		27 / 9.1	8 / 17	33.4 / 8.54	H. vulgare	CHI1_HORVU	
302	Actin and related proteins	212		30 / 9.2	13 / 31	28.5 / 5.31	A. oryzae	Q2U9E1_ASPOR	
303	Actin and related proteins	137		27 / 9.2	12 / 31	28.5 / 5.31	A. oryzae	Q2U9E1_ASPOR	
304	Actin and related proteins	106		30 / 9.3	6 / 23	28.5 / 5.31	A. oryzae	Q2U9E1_ASPOR	
305	Class II chitinase (EC 3.2.1.14)	74		25 / 9.2	8 / 40	28.2 / 8.66	T. aestivum	Q4Z8L7_WHEAT	
306	26 kDa endochitinase 1 precursor (EC 3.2.1.14)	119		25 / 9.4	8 / 28	33.4 / 8.54	H. vulgare	CHI1_HORVU	
307	Wheat EST match	74	7S Globulin storage protein	29 / 9.5	19 / 37	39.3 / 11.17	T. aestivum	Ta5443-3	GI:170696

Table A-2.6 (	continued)
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-	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
308	Putative 40S ribosomal protein S3	76		25 / 9.5	9 / 22	25.4 / 9.63	T. aestivum	Q8L804_WHEAT	
309	1-Cys-peroxiredoxine	26		24 / 6.1	5 / 19	23.95 / 6.08	T. aestivum	Q6W8Q2_WHEAT	
310	Wheat EST match	90	7S Globulin storage protein	26 / 6.2	21 / 72	23.6 / 9.99	T. aestivum	Ta136340-2	GI:170696
311	1-Cys-peroxiredoxine	50		24 / 6.4	17 / 52	23.95 / 6.08	T. aestivum	Q6W8Q2_WHEAT	
312	Wheat EST match	28	1-Cys-peroxiredoxine	25 / 6.45	12 / 29	34.6 / 8.33	T. aestivum	Ta1386253404-1	GI:34539782
313	Prohibitin protein Wph	50		23 / 6.6	5 / 15	30.1 / 6.0	T. aestivum	Q84VJ0_WHEAT	
314	Wheat EST match	218	Cupin family protein, expressed	21 / 7.1	10 / 14	66.9 / 7.64	O. sativa	Ta2606253404-2	GI:108708022
315	Endogenous alpha-amylase/subtilisin inhibitor (WASI)	59		22 / 7.2	10 / 61	19.6 / 6.77	T. aestivum	IAAS_WHEAT	
316	Wheat EST match	138	Cupin family protein, expressed	19 / 7.5	14 / 20	66.9 / 7.64	O. sativa	Ta2606253404-2	GI:108708022
317	7S Globulin storage protein	52		21 / 8.4	21 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
318	7S Globulin storage protein	79		21 / 8.5	26 / 29	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
319	7S Globulin storage protein	60		21 / 8.7	12 / 15	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
320	Wheat EST match	73	P0481E12.28	24 / 8.7	14 / 36	37.7 / 7.75	O. sativa	Ta369-2	GI:34909516
321	Wheat EST match	68	7S Globulin storage protein	25 / 9.05	11 / 20	33.7 / 11.52	T. aestivum	Ta5446-1	GI:170696
322	Class II chitinase (EC 3.2.1.14)	102		25 / 9.2	6 / 30	28.2 / 8.66	T. aestivum	Q4Z8L7_WHEAT	
323	Basic endochitinase C precursor (EC 3.2.1.14) (Rye seed chitinase-c)	43		26 / 9.2	5 / 21	28.3 / 8.82	S. cereale	CHIC_SECCE	
324	Wheat EST match	76	7S Globulin storage protein	23 / 9.3	10 / 21	39.3 / 11.17	T. aestivum	Ta5443-3	GI:170696
325	Wheat EST match	131	7S Globulin storage protein	24 / 9.6	14/31	39.3 / 11.17	T. aestivum	Ta5443-3	GI:170696
326	Wheat EST match	108	7S Globulin storage protein	23 / 9.55	14 / 27	38.0 / 9.56	T. aestivum	Ta5441-3	GI:170696
327	Glutathione transferase (EC 2.5.1.18)	71		23 / 6.3	17 / 52	24.98 / 6.35	T. aestivum	Q8RW04_WHEAT	
328	No match								
329	Manganese superoxide dismutase (EC 1.15.1.1)	159		23 / 6.3	13 / 39	25.3 / 7.9	T. aestivum	P93606_WHEAT	
330	Manganese superoxide dismutase (EC 1.15.1.1)	217		23 / 6.4	15 / 45	25.3 / 7.9	T. aestivum	Q96185_WHEAT	

 Table A-2.6 (continued)

-	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
331	No match								
332	Wheat EST match	42	Unnamed protein product	18 / 6.2	22 / 50	27.9 / 7.27	T. aestivum	Ta13210-3	GI:21813
333	No match								
334	No match								
335	Endogenous alpha-amylase/subtilisin inhibitor (WASI)	231		17 / 6.9	23 / 82	19.6 / 6.77	T. aestivum	IAAS_WHEAT	
336	Endogenous alpha-amylase/subtilisin inhibitor (WASI)	94		15 / 7.3	19 / 76	19.6 / 6.77	T. aestivum	IAAS_WHEAT	
337	Wheat EST match	159	Cupin family protein, expressed	18 / 8.4	8 / 10	66.9 / 7.64	T. aestivum	Ta2606253404-2	GI:108708022
338	7S Globulin storage protein	50		21 / 9	12 / 14	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
339	Actin and related proteins	168		20 / 9	14 / 37	28.5 / 5.31	A. oryzae	Q2U9E1_ASPOR	
340	Wheat EST match	50	7S Globulin storage protein	19 / 9.0	10 / 20	39.3 / 11.17	T. aestivum	Ta5443-3	GI:170696
341	Wheat EST match	49	7S Globulin storage protein	20 / 9.05	14 / 15	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
342	Secretory protein	73		21 / 9.3	6 / 25	24.2 / 9.32	T. aestivum	Q9SWZ5_WHEAT	
343	7S Globulin storage protein	64		20 / 9.3	14 / 15	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
344	7S Globulin storage protein	55		20 / 9.3	11 / 12	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
345	Secretory protein	49		21/9.5	7 / 24	24.2 / 9.32	T. aestivum	Q9SWZ5_WHEAT	
346	7S Globulin storage protein	63		20 / 9.4	15 / 18	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
347	Wheat EST match	81	7S Globulin storage protein	21 / 9.5	10 / 18	40.0 / 11.75	T. aestivum	Ta9894-2	GI:170696
348	7S Globulin storage protein	47		19 / 9.4	11 / 10	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
349	7S Globulin storage protein	45		21 / 9.5	18 / 17	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
350	7S Globulin storage protein	59		19/9.5	18 / 18	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
351	7S Globulin storage protein	56		20 / 9.6	15 / 17	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
352	Wheat EST match	75	7S Globulin storage protein	18 / 9.6	13 / 29	38.0 / 9.56	T. aestivum	Ta5441-3	GI:170696
353	7S Globulin storage protein	49		20 / 9.8	8 / 4	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
354	Wheat EST match	49	7S Globulin storage protein	18 / 9.75	12 / 26	38.0 / 9.56	T. aestivum	Ta5441-3	GI:170696
355	Wheat EST match	76	7S Globulin storage protein	22 / 10.2	9 / 8	78.9 / 8.75	T. aestivum	Ta2154253404-2	GI:170696
356	Wheat EST match	151	7S Globulin storage protein	21 / 10.75	16 / 12	78.9 / 8.75	T. aestivum	Ta2154253404-2	GI:170696

<b>Table A-2.6</b> (0	continued)
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	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
357	Nucleoside diphosphate kinase (E 2.7.4.6) (Fragment)	EC 62		13 / 6.3	2 / 50	2.68 / 4.87	A. sativa	Q9S8M2_AVESA	
358	Wheat EST match	125	7S Globulin storage protein	12 / 6.2	19 / 38	38.9 / 9.17	T. aestivum	Ta3078253411-3	GI:170696
359	Wheat EST match	49	Heat shock protein 16.9B	15 / 6.4	12 / 27	25.8 / 6.43	T. aestivum	Ta238185-3	GI:21805
360	Wheat EST match	63	7S Globulin storage protein	14 / 6.4	31/39	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
361	Wheat EST match	54	7S Globulin storage protein	15 / 6.45	16 / 26	38.0 / 9.56	T. aestivum	Ta5441-3	GI:170696
362	Wheat EST match	33	7S Globulin storage protein	15 / 6.6	11 / 19	41.3 / 9.42	T. aestivum	Ta3078253409-1	GI:170696
363	7S Globulin storage protein	60		14 / 6.75	16 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
364	Wheat EST match	66	Stress-inducible membrane pore protein	15 / 6.9	8 / 12	39.3 / 10.09	B. inermis	Ta58534-1	GI:16555405
365	Wheat EST match	458	7S Globulin storage protein	12 / 6.9	18 / 20	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
366	Wheat EST match	394	7S Globulin storage protein	14 / 7.05	20 / 23	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
367	Hypothetical protein OSJNBb0094P23.22	39		15 / 7.3	4 / 92	5.6 / 11.88	O. sativa	Q6Z1K0_ORYSA	
368	No match								
369	Prohibitin protein Wph	107		16/9	6 / 15	30.1 / 6.0	T. aestivum	Q84VJ0_WHEAT	
370	OSJNba0093F12.16 protein	89		16/9.2	10 / 53	17.7 / 9.23	O. sativa	Q7XNU2_ORYSA	
371	Wheat EST match	110	7S Globulin storage protein	18 / 9.3	17 / 26	33.7 / 11.52	T. aestivum	Ta5446-1	GI:170696
372	Wheat EST match	46	7S Globulin storage protein	17 / 9.45	11 / 27	39.3 / 11.17	T. aestivum	Ta5443-3	GI:170696
373	Wheat EST match	107	7S Globulin storage protein	17 / 9.55	12 / 17	38.0 / 9.56	T. aestivum	Ta5441-3	GI:170696
374	7S Globulin storage protein	75		17 / 9.6	11 / 11	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
375	7S Globulin storage protein	67		17/9.6	17 / 17	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
376	Wheat EST match	85	7S Globulin storage protein	17 / 10.75	8 / 5	78.9 / 8.75	T. aestivum	Ta2154253404-2	GI:170696
377	Putative nucleoside diphosphate kina	ase 82		10 / 6.2	8 / 30	25.9 / 8.88	O. sativa	Q5TKF4_ORYSA	
378	Wheat EST match	188	7S Globulin storage protein	13 / 6.5	13 / 16	76.3 / 9.22	T. aestivum	Ta3078253413-3	GI:170696

Table A-2.6	(continued)

	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
379	7S Globulin storage protein	45		9 / 6.4	17 / 19	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
380	16.9 kDa class I heat shock protein	153		9 / 6.6	7 / 37	16.9 / 5.83	T. aestivum	HSP11_WHEAT	
381	R2R3MYB-domain protein (Fragment)	35		9 / 7	6 / 81	4.85 / 8.2	Z. mays	Q9SQC0_MAIZE	
382	Wheat EST match	53	7S Globulin storage protein	7 / 7.3	27 / 28	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
383	Putative nucleoside diphosphate kinase	50		9 / 7.4	9 / 24	25.9 / 8.88	O. sativa	Q5TKF4_ORYSA	
384 385	Wheat EST match No match	56	OSJNBa0027P08.9	9 / 7.5	25 / 37	36.9 / 9.06	O. sativa	Ta13211-3	GI:50924568
386	Putative 60S ribosomal protein L12	205		16 / 9.35	10 / 41	17.7 / 9.23	O. sativa	Q6Z8E0_ORYSA	
387	Wheat EST match	65	40S subunit ribosomal protein	14 / 9.5	6 / 16	19.5 / 9.84	O. sativa	Ta33749-2	GI:55775376
388	7S Globulin storage protein	64		16/9.5	10 / 11	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
389	Oleosin (Fragment)	88		14 / 9.5	4 / 35	5.79 / 8.59	H. vulgare	Q43474_HORVU	
390	7S Globulin storage protein	80		16/9.7	11 / 12	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
391	7S Globulin storage protein	66		16 / 9.8	9 / 8	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
392	7S Globulin storage protein	71		16 / 11	13 / 10	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
393	Wheat EST match	123	7S Globulin storage protein	16 / 10.75	17/33	33.7 / 11.52	T. aestivum	Ta5446-1	GI:170696
394	Wheat EST match	60	7S Globulin storage protein	13 / 10.7	10 / 19	39.3 / 11.17	T. aestivum	Ta5443-3	GI:170696
395	Wheat EST match	71	7S Globulin storage protein	9 / 6.35	10 / 15	66.9 / 6.85	T. aestivum	Ta3078253407-1	GI:170696
396	Putative nucleoside diphosphate kinase	61		8 / 6.3	7 / 23	25.9 / 8.88	O. sativa	Q5TKF4_ORYSA	
397	7S Globulin storage protein	63		7 / 6.25	14 / 18	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
398	Putative nucleoside diphosphate kinase	59		9 / 6.4	6 / 22	25.9 / 8.88	O. sativa	Q5TKF4_ORYSA	
399	7S Globulin storage protein	52		9 / 6.5	6 / 5	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
400	Wheat EST match	82	7S Globulin storage protein	8 / 6.5	6 / 13	29.7 / 9.12	T. aestivum	Ta3078253412-1	GI:170696
401	7S Globulin storage protein	82		6 / 6.7	16 / 22	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
402	78 Globulin storage protein	53		7 / 6.75	17 / 21	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	

Table A-2.6	(continued)
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	Protein name (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
403	Wheat EST match	96	7S Globulin storage protein	6 / 6.9	5 / 5	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
404	7S Globulin storage protein	73		7 / 7	7 / 8	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
405	Hypothetical protein OJ1124_E11.8.	43		9 / 7.2	9 / 32	10.9 / 9.52	O. sativa	Q6K875_ORYSA	
406	7S Globulin storage protein	77		6 / 7.25	18 / 24	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
407	Wheat EST match	62	7S Globulin storage protein	5 / 7.25	9 / 8	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
408	7S Globulin storage protein	49		7 / 7.5	19 / 23	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
409	No match								
410	Embryo globulin	67		9 / 9.1	10 / 10	72.2 / 6.8	H. vulgare	Q03678_HORVU	
411	Wheat EST match	43	7S Globulin storage protein	9 / 9.2	9 / 8	61.7 / 8.08	T. aestivum	Ta3078253410-1	GI:170696
412	Wheat EST match	90	Unknown protein	9 / 9.4	7 / 19	27.1 / /10.16	O. sativa	Ta1007253406-2	GI:52353767
413	Wheat EST match	89	7S Globulin storage protein	10 / 9.65	12 / 15	52.8 / 9.69	T. aestivum	Ta627253413-2	GI:170696
414	Wheat EST match	126	Unknown protein	9 / 9.65	6 / 13	21.99 / 9.69	O. sativa	Ta131241-1	GI:52353767
415	No match								
416	Disease-resistent-related protein	45		9 / 11	8 / 23	15.4 / 11.16	O. sativa	Q8SA80_ORYSA	
417	Wheat EST match	51	7S Globulin storage protein	10 / 10.7	11 / 19	39.3 / 11.17	T. aestivum	Ta5443-3	GI:170696
418	50S ribosomal protein L31	31		5 / 6.9	5 / 60	9.36 / 9.22	X. oryzae	Q2P6M2_XANOR	
419	7S Globulin storage protein	67		6 / 7	8 / 10	72.2 / 6.8	T. aestivum	Q7DMU0_WHEAT	
420	No match								
421	Wheat EST match	104	7S Globulin storage protein	5 / 7.0	21 / 24	52.8 / 9.69	T. aestivum	Ta627253413-2	GI:170696
422	Wheat EST match	103	7S Globulin storage protein	6 / 7.1	7 / 8	52.8 / 9.69	T. aestivum	Ta627253413-2	GI:170696
423	Wheat EST match	109	7S Globulin storage protein	5 / 7.4	13 / 15	69.9 / 8.42	T. aestivum	Ta628253405-1	GI:170696
424	Wheat EST match	119	7S Globulin storage protein	5 / 7.9	10 / 11	52.8 / 9.69	T. aestivum	Ta627253413-2	GI:170696
425	Wheat EST match	97	7S Globulin storage protein	5 / 8.05	5 / 5	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
426	No match								
427	Wheat EST match	77	7S Globulin storage protein	5 / 8.1	20 / 25	67.9 / 8.86	T. aestivum	Ta628253404-2	GI:170696
428	Wheat EST match	115	7S Globulin storage protein	5.5 / 9.2	9 / 13	52.8 / 9.69	T. aestivum	Ta627253413-2	GI:170696
429	Wheat EST match	107	78 Globulin storage protein	6/9.25	7 / 10	52.8 / 9.69	T. aestivum	Ta627253413-2	GI:170696

Tuble II 2.0 (continued)	Table A-2.6	(continued)
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	Protein name (Mascot cereal search)		Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
430	Wheat EST match		125	7S Globulin storage protein	5 / 9.4	7 / 8	52.8 / 9.69	T. aestivum	Ta627253413-2	GI:170696
	Elongation factor 1-alpha alpha)	(EF-1-	61		9 / 9.4	14 / 23	49.1 / 9.2	T. aestivum	EF1A_WHEAT	
432	Wheat EST match		109	Elongation factor 1-alpha, putative, expressed	9 / 9.5	5 / 17	29.2 / 9.84	O. sativa	Ta3484253413-3	GI:108706480
433	Transposase IS1478		49		9.5 / 9.5	10 / 35	21.5 / 11.55	X. oryzae	Q5H473_XANOR	
434	Wheat EST match		51	Histone H3	4 / 10.75	8 / 26	15.7 / 10.98	O. sativa	Ta3497253410-1	GI:34898304
	26 kDa endochitinase 1 precursor 3.2.1.14)	r (EC	121		23 / 9.2	6 / 13	33.4 / 8.54	H. vulgare	CHI1_HORVU	

opot Protein name No. (Mascot cereal search)	Mascot or GPM <sup>1</sup> score	Protein name (Wheat EST BLAST search) <sup>2</sup>	Observed MW (KDa)/pI	Matching peptides / %Coverage	Theoretical <sup>3</sup> MW (Da)/pI	Species with homologous protein	Swiss-Prot accession number <sup>4</sup>	GenBank accession number
1 Oxalate oxidase precursor	90		67 / NA	5 / 18	23.5 / 6.9	T. aestivum	Q70PK0_WHEAT	
2 Polyphenol oxidase (Fragment)	391		65 / NA	27 / 50	46.6 / 5.23	T. aestivum	Q6PLQ9_WHEAT	
3 Peroxidase (EC 1.11.1.7) (Fragment)	52		33 / NA	4 / 37	8.36 / 4.34	T. aestivum	PERX_WHEAT	
4 Endochitinase precursor	304		30 / NA	15 / 48	33.6 / 7.38	T. aestivum	Q41539_WHEAT	
5 PSBGer3 protein	73		27 / NA	4 / 16	23.4 / 6.18	T. aestivum	P93600	
6 Histone H4 variant TH011	33		14 / NA	5 / 56	11.3 / 11.48	T. aestivum	H41_WHEAT	

Table A-2.7	Identification	of proteins	in supernatant	from	imbibed grain
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1 GPM scores -log(e) values

2 Wheat EST NCBI BLAST search matching to proteins with e-values less than 0.0001

3 MW and pI are of fragment and wheat EST match sequence and not of actual complete protein

4 Accession numbers beginning with Ta are for the Wheat EST database

# A-3 Construction of Wheat EST Database for searching using XTandem and Mascot

Python Program. Requires Biopython installation see Biopython.org:

#### #!C:\Program files\Python24\python.exe

import os, sys, string

from Bio import Fasta, Translate, Alphabet, Seq

from Bio.Alphabet import IUPAC

from Bio.Seq import Seq

my\_alpha = IUPAC.unambiguous\_dna

standard\_translator = Translate.unambiguous\_dna\_by\_id[1]

parser = Fasta.RecordParser()

translated = ""

# The file sequences.txt must be in the folder in which the program is running

# Sequences.txt is the FASTA Wehaet EST-contig file

file\_to\_parse = open("sequences.txt", 'r')

iterator = Fasta.Iterator(file\_to\_parse, parser)

while 1:

```
cur_record = iterator.next()
translate_all = []
new_seq = []
```

if cur\_record is None:

break

# create reverse complement

complement\_table = {"A": "T", "T": "A", "C": "G", "G": "C", "N": "N", "R": "Y", "Y": "R", "K": "M", "M": "K", "S": "W", "W": "S", "B": "D", "D": "B", "H": "V", "V": "H"}

for letter in cur\_record.sequence:

complement\_letter = complement\_table[letter]

new\_seq.append(complement\_letter)

new\_seq.reverse()

rev\_seq = "".join(new\_seq)

# append the title to the translated list

title\_atoms = string.split(cur\_record.title)

old accession = string.split(title atoms(PCT/DK98/00460), '|')

int\_accession = string.split(old\_accession[0:1], ",")

new\_accession = "Ta" + int\_accession(PCT/DK98/00460)

title\_atoms2 = ' '.join(title\_atoms[0:])

title 1 = "> " + new accession + "-1" + " " + title atoms2 + "\n"

 $title_2 = "> " + new_accession + "-2" + " " + title_atoms2 + "\n"$ 

title 3 = "> " + new accession + "-3" + " " + title atoms2 + "\n"

 $title_4 = "> " + new_accession + "-4" + " " + title_atoms2 + "\n"$ 

title\_5 = "> " + new\_accession + "-5" + " " + title\_atoms2 + "\n"

title 6 = "> " + new accession + "-6" + " " + title atoms2 + "\n"

translate\_all.append(title\_1)

for\_seq = Seq(cur\_record.sequence, my\_alpha)

prot\_for = standard\_translator.translate(for\_seq)

for prot = ".join(prot for) + "n"

translate\_all.append(for\_prot)

translate\_all.append(title\_2)

prot\_for = standard\_translator.translate(for\_seq[1:])

for\_prot = ".join(prot\_for) + "\n"

translate\_all.append(for\_prot)

translate\_all.append(title\_3)

prot\_for = standard\_translator.translate(for\_seq[2:])

for\_prot = ".join(prot\_for) + "\n"

translate\_all.append(for\_prot)

translate\_all.append(title\_4)

reverse\_seq = Seq(rev\_seq, my\_alpha)

prot\_rev = standard\_translator.translate(reverse\_seq)

rev\_prot = ".join(prot\_rev) + "\n"

translate\_all.append(rev\_prot)

translate\_all.append(title\_5)

prot\_rev = standard\_translator.translate(reverse\_seq[1:])

rev\_prot = ".join(prot\_rev) + "\n"

translate\_all.append(rev\_prot)

translate\_all.append(title\_6)

prot\_rev = standard\_translator.translate(reverse\_seq[2:])

rev\_prot = ".join(prot\_rev) + "\n"

translate\_all.append(rev\_prot)

translated = ".join(translate\_all)

result\_file\_name = os.path.join(os.getcwd(), 'TranslatedESTs.fasta')

result\_file = open(result\_file\_name, 'a')

result\_file.write(translated)

result\_file.close()

# A-4 Wheat EST alignments for cupin domain

containing proteins

### Table A-4.1 Wheat EST alignments with cupin containing proteins

Alignment: C:\D Seaview [blocks	ocuments and -10 fontsize	i Settings\t ==10 A4] on	:jerkovi\My Fri Sep 15	Documents\: 16:52:22 20	seaview\ALL 006	_mod2.aln
	1					
Ta2154253404-2		VGDRARVTIP	TT PTT OTOTT	ER RAUER CUT	EEEDDD OOUC	
Ta9905-2	OIVENIOR PR	A ODIENTEA TTE	DDF DD <mark>919</mark> DD	FARAVSASHD	EEEDRA 30H3	
Ta3078253410-1						
Ta3078253407-1	BGAROAE	QTMAI <mark>R</mark> ATIP				LOOCAO COO
Ta3078253412-1						
Ta3078253409-1						
Ta3078253413-3		MATRGRATIP			EEEDRRGGRS	LORCVORCOO
Ta5446-1						
AAA34269		MATRAKATIP	LLFLLGTSLL	FAAAVSASHD	DEDDRR GGHS	LOOCVORCEO
Q03678 HORVU		MATRAKATIP	LLFLLGTSLL	FAAAVSASHD	DEDDRRGGHS	LOOCVORCEO
Ta194625-1						
Ta5441-3						
Ta9894-2						
Q852L2 ORYSA						
Q8L8I0 ORYSA						
Ta628253404-2						
Ta628253407-1						
Ta136340-2						
Ta627253413-2						
Ta628253405-1						
Ta132809-2						
Ta189317-3						
Ta2606253404-2						
Ta2606253405-2						
Q70PK0_WHEAT						
ATEG_07804						
-						
	61					
Ta2154253404-2	DRPRYSHARC	VOBCRGDOOO	HGRHEQEE-Q	GRGRGRHGEG	EREEEQGGGR	GRHGEGEREE
Ta9905-2						
Ta3078253410-1						
Ta3078253407-1	DRPRYSHARC	VQECREDQQQ	HGRHEOEE-O	GHSHGRHGEG	GREEEOGRGR	GRHGOGEREE
Ta3078253412-1						
Ta3078253409-1						
Ta3078253413-3	DEPENSHARC	VQECRDDQQQ	HCRUPOPP-O	GROUCE HORO	EP REP <mark>OCE</mark> CP	GREGO GEREE
Ta5446-1		· Sactor · · · · ·				
AAA34269	ERPRYSHARC	VQECRDDQQQ	HGRHEQEEEQ	an a	EREEEHGRGR	GRHGEGEREE
003678 HORVU	ERPRYSHARC	NO DO DO DO	HORHEVELEV	CRORONHODO	EREEEHGRGR	
Ta194625-1	ERFRISHERC	A BCEDRAGA	HORNEYEEEV	GROKOWHOEG	EREEEROROR	GRHGEGEREE
Ta5441-3						
Ta9894-2						
Q852L2_ORYSA	MA				NAVGWSR	
Q8L810 ORYSA	<mark>MA</mark>			SLCF-LSLAS	NAVGWSR	RGEQBEEDER
Ta628253404-2	SPT	S I IM <mark>K</mark> STVVR				RRWQEGGDDE
Ta628253407-1	<mark>SP</mark> T		SPWLALALVL	SLCLSLSFAS	WDAEDEGRGS	RRWOEGGD-E
Ta136340-2	VSSSRT	TITMKS-AVR	SPWLVLAIVL	SLCLSLSFAS	WDAEDVGRGS	RR <mark>WQ</mark> EGGD-E
Ta627253413-2						
Ta628253405-1		<mark>D</mark>	HHWLVLAIVL	SLCLSLSFAS	WDAEDVGRGS	RR <mark>WQ</mark> EGGD-E
Ta132809-2						
Ta189317-3						
Ta2606253404-2			EEGEGE	WRPBEEAKGG	GGGGG <mark>T</mark> G <mark>K</mark> GL	FLLDRVERVV
Ta2606253405-2					GGGGGAGKGL	
Q70PK0 WHEAT						
ATEG 07804						

Alignment: C:\Documents and Settings\tjerkovi\My Documents\seaview\ALL mod2.aln

	21					
Ta2154253404-2	EEGRGRGRHG	EGER			EEHGRH	EOGE GER GEG
Ta9905-2						
Ta3078253410-1						
Ta3078253407-1	EEGRGRGRHG	EGER			EEHGKH	EQGR.GRR.GEG
Ta3078253412-1						
Ta3078253409-1		<u></u>				
Ta3078253413-3	EQGRGRGRRG	EGER			DEE	
Ta5446-1						
AAA34269	EHGRGRGRHG	EGEREEERGR	GHGRHGEGER	EEERGRGRGR		GRGRGRRGEG
Q03678_HORVU	EHGRGRGRHG	BGBR BBBR GR	GHGRHGEGER	EEERGR GR GR GR	HGEGE <mark>R</mark> EEEE	GRGRGRRGEG
Ta194625-1						
Ta5441-3						
Ta9894-2						
Q852L2_ORYSA	RRHGGE					
Q8L810_ORYSA	RRHGGE					
Ta628253404-2	GRSGSG					
Ta628253407-1	RRSGE					
Ta136340-2	G <mark>RS</mark> GG					
Ta627253413-2						
Ta628253405-1	G <mark>RS</mark> GG					
Tal32809-2 Tal89317-3						
Ta169317-3 Ta2606253404-2	ESEGG-					
Ta2606253404-2 Ta2606253405-2	ESEGG					
Q70PK0 WHEAT	E9 E/9/9					
ATEG 07804						
A123_07804						
18	31					
Ta2154253404-2	B1 ERDEEOGGSR	RPYVFGPRNF	RSIIRSDHGF	VNALRPFDEV	SELLEGIENY	RVAIMEVNER
		RPYVFGPRNF	R <mark>SIIR</mark> SDHGF	VKALRPFDEV	SRLLRGIRNY	RVAIMEVNER
Ta2154253404-2		RPYVFGPRNF	RS I IRSDHGF	VKALRPFDEV	SRLLRGIRNY	RVAIMEVNPR
Ta2154253404-2 Ta9905-2		RPYVFGPRNF		VKALRPFDEV VKALRPFDEV		IMEVNPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1						IMEVNPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1						IMEVNPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1 Ta3078253413-3				VKALRPFDEV	SRLLRGIRNY	IMEVNPR RVA IMEVNPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1	ERDEEDGGSR ERDEEHGDSR	RPYVFGPRNF RPYVFGPRSF	RSIIRSDHGP RRIIRSDHGF	VKALRPFDEV VKALRPFDEV	SRLLRGIRNY SRLLRGIRNY	IMEVNPR RVA IMEVNPR RVA IMEVNPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269	ERDEEQGGSR ERDEEHGDSR		RS I IRSDHGF	VKALRPFDEV VKALRPFDEV VRALRPFDEV	SRLLRGIRNY	IMEVNPR RVA IMEVNPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU	ERDEEDGGSR ERDEEHGDSR	RPYVFGPRNF RPYVFGPRSF	RSIIRSDHGP RRIIRSDHGF	VKALRPFDEV VKALRPFDEV	SRLLRGIRNY SRLLRGIRNY	IMEVNER EVAIMEVNER EVAIMEVNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1	ERDEELGDSR HGDGR ERDEELGDSR	RPYVFGPRNF RPYVFGPRSF	RSIIRSDHGP RRIIRSDHGF	VKALRPFDEV VKALRPFDEV VRALRPFDEV	SRLLRGIRNY SRLLRGIRNY SRLLRGIRNY	IMEVNER EVAIMEVNER EVAIMEVNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AA34269 Q03678_HORVU Ta194625-1 Ta5441-3	ERDEELGDSR HGDGR ERDEELGDSR	RPYVFGPRNF RPYVFGPRSF	RSIIRSDHGP RRIIRSDHGF	VKALRPFDEV VKALRPFDEV VRALRPFDEV	SRLLRGIRNY SRLLRGIRNY SRLLRGIRNY	IMEVNER EVAIMEVNER EVAIMEVNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2	ERDEENGDSR HGDGR ERDEENGDSR ERDEENGDSR ERDEENGDSR	RPYVFGPRNF RPYVFGPRSF RPYVFGPRSF	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF	VKALRPFDEV VKALRPFDEV VRALRPFDQV VRALRPFDQV	SRLLRGIRNY SRLLRGIRNY SRLLRGIRDY SRLLRGIRDY	RVAIMEVNPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA	ERDEENGDSR ERDEENGDSR ERDEENGDSR ERDEENGDSR GG	RFYVFGPRNF RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR	VKALRPFDEV VKALRPFDEV VRALRPFDQV VRALRPFDQV FSVLERFPDE	SRLLRGIRNY SRLLRGIRNY SRLLRGIRDY SRLLRGIRDY	VAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810_ORYSA	ERDEENGDSR ERDEENGDSR ERDEENGDSR ERDEENGDSR GG GG	RPYVFGPRNF RPYVFGPRSF RPYVFGPRSF RPYVFGPRSF RPYHLGEESF	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR	VKALRPFDEV VKALRPFDEV VRALRPFDQV VRALRPFDQV FSVLERFPDE	SRLLRGIRNY SRLLRGIRNY SRLLRGIRDY SRLLRGIRDY OVVGAAVGGY	VAIMEVNPR VAIMEVNPR VAIMEVNPR VAIMEVNPR VAIMEVNPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2	ERDEELGDSR ERDEELGDSR ERDEELGDSR ERDEELGDSR GG GG SG	RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYHLGEESF RFYHLGEESF	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR RHWTRTRHGR	VKALRPFDEV VKALRPFDEV VRALRPFDQV VRALRPFDQV FSVLERFPDE FSVLERFPDE FSVLERF-DH	SRLLRGIRNY SRLLRGIRNY SRLLRGIRDY SRLLRGIRDY OVVGAAVGGY ELLRGSIGDY	VAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q852L2_ORYSA Q852L2_ORYSA Ta628253404-2 Ta628253407-1	ERDEELGDSR HGDGR ERDEELGDSR ERDEELGDSR GG GG GG GG SG 	RFYVFGFRSF RFYVFGFRSF RFYVFGFRSF RFYHLGEESF RFYHLGEESF RFYHFGEESF	REIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR RHWTRTRHGR REWAKSRHGH	VKALRPFDEV VKALRPFDEV VRALRPFDQV VRALRPFDQV FSVLERFPDE FSVLERFPDE FKVLERF-DH	SRLLRGIRNY SRLLRGIRNY SRLLRGIRDY SRLLRGIRDY OVVGAAVGGY OVVGAAVGGY ELLRGSIGDY	VAIMEVNPR VAIMEVNPR VAIMEVNPR VAIMEVNPR VAIMEVNPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253412-1 Ta3078253412-1 Ta3078253412-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2	ERDEELGDSR ERDEELGDSR ERDEELGDSR ERDEELGDSR GG GG SG	RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYHLGEESF RFYHLGEESF	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR RHWTRTRHGR	VKALRPFDEV VKALRPFDEV VRALRPFDQV VRALRPFDQV FSVLERFPDE FSVLERFPDE FSVLERF-DH	SRLLRGIRNY SRLLRGIRNY SRLLRGIRDY SRLLRGIRDY OVVGAAVGGY ELLRGSIGDY	VAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q8L810_ORYSA Ta628253404-2 Ta628253407-2 Ta628253407-2 Ta627253413-2	ERDEEQGOSR ERDEEQGDSR ERDEEQGDSR ERDEEQGDSR GG GG SG SG SG	RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYHLGEESF RFYHLGEESF RFYHFGEESF RFYHFGEESF RFYHFGEESF	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR RHWTRTRHGR REWAKSRHGH REWAKSRHGH	VKALRPFDEV VKALRPFDEV VRALRPFDOV VRALRPFDOV FSVLERFPDE FSVLERFPDE FKVLERF-DH FKVLERF-DH	SRLLRGIRMY SRLLRGIRMY SRLLRGIRDY SRLLRGIRDY OVVGAAVGGY OVVGAAVGGY ELLRGSIGDY ELLRGSIGDY	IMEVNER RVAIMEVNER RVAIMEVNER RVAIMEVNER RVAIMEVNER RVAVLEAAFR RVAVLEAAFR RVAVLEAAFR RVAVLEAAFR RVAVLEAAFR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q8L810_ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2 Ta628253405-1	ERDEELGDSR HGDGR ERDEELGDSR ERDEELGDSR GG GG GG GG SG 	RFYVFGFRSF RFYVFGFRSF RFYVFGFRSF RFYHLGEESF RFYHLGEESF RFYHFGEESF	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR RHWTRTRHGR REWAKSRHGH REWAKSRHGH	VKALRPFDEV VKALRPFDEV VRALRPFDOV VRALRPFDOV FSVLERFPDE FSVLERFPDE FSVLERFPDE FSVLERFPDE FSVLERFDH FSVLERFDH	SRLLRGIRNY SRLLRGIRNY SRLLRGIRDY SRLLRGIRDY OVVGAAVGGY OVVGAAVGGY ELLRGSIGDY	IMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta628253405-1 Ta132809-2	ERDEEDGOSR ERDEEDGOSR ERDEEDGOSR ERDEEDGOSR GG GG GG SG SG SG	RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYHLGEESF RFYHLGEESF RFYHFGEESF RFYHFGEESF RFYHFGEESF	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR RHWTRTRHGR REWAKSRHGH REWAKSRHGH	VKALRPFDEV VKALRPFDEV VRALRPFDOV VRALRPFDOV FSVLERFPDE FSVLERFPDE FSVLERFPDE FSVLERFPDE FSVLERFDH FSVLERFDH	SRLLRGIRMY SRLLRGIRMY SRLLRGIRDY SRLLRGIRDY OVVGAAVGGY OVVGAAVGGY ELLRGSIGDY ELLRGSIGDY	IMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2 Ta628253405-1 Ta132809-2 Ta189317-3	ERDEELGDSR ERDEELGDSR ERDEELGDSR ERDEELGDSR GG GG SG SG SG SG SG	RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYHLGEESF RFYHFGEESF RFYHFGEESF RFYHFGEESY RFYHFGEESY	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR RHWTRTRHGR REWAKSRHGH REWAKSRHGH REWAKSRHGH	VKALRPFDEV VKALRPFDEV VRALRPFDQV VRALRPFDQV FSVLERFPDE FSVLERFPDE FKVLERF-DH FKVLERF-DH	SRLLRGIRNY SRLLRGIRNY SRLLRGIRDY SRLLRGIRDY OVVGAAVGGY ELLRGSIGDY ELLRGSIGDY ELLRGSIGDY	VAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253412-1 Ta3078253412-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253405-1 Ta136340-2 Ta628253405-1 Ta132809-2 Ta189317-3 Ta2606253404-2	ERDEEQGGSR ERDEENGDSR ERDEEQGDSR ERDEEQGDSR GG GG SG SG SG SG SG	RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYHLGEESF RFYHLGEESF RFYHFGEESF RFYHFGEESF RFYHFGEESF RFYHFGEESF RFYHFGEESF	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR RHWTRTRHGR REWAKSRHGH REWAKSRHGH REWAKSRHGH	VKALRPFDEV VKALRPFDEV VRALRPFDQV VRALRPFDQV FSVLERFPDE FSVLERFPDE FKVLERF-DH FKVLERF-DH FKVLERF-DH	SRLLRGIRNY SRLLRGIRNY SRLLRGIRDY SRLLRGIRDY OVVGAAVGGY ELLRGSIGDY ELLRGSIGDY ELLRGSIGDY	VAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q85212 ORYSA Q8L810 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta628253405-1 Ta132809-2 Ta189317-3 Ta2606253405-2	ERDEELGDSR ERDEELGDSR ERDEELGDSR ERDEELGDSR GG GG SG SG SG SG SG	RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYHLGEESF RFYHLGEESF RFYHFGEESF RFYHFGESSF RFYHFGESSF RFYHFGESSF RFYHFGESS RFYHFGESSY RFYHFGESSY	RSIIRSDHGF RRIIQSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR RHWTRTRHGR REWAKSRHGH REWAKSRHGH REWAKSRHGH REWAKSRHGH	VKALRPFDEV VKALRPFDEV VRALRPFDQV VRALRPFDQV FSVLERFPDE FSVLERFPDE FSVLERF-DH FSVLERF-DH FSVLERF-DH	SRLLRGIRMY SRLLRGIRMY SRLLRGIRDY SRLLRGIRDY SRLLRGIRDY ELLRGSIGDY ELLRGSIGDY ELLRGSIGDY ELLRGSIGDY	RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253412-1 Ta3078253412-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253405-1 Ta136340-2 Ta628253405-1 Ta132809-2 Ta189317-3 Ta2606253404-2	ERDEEQGGSR ERDEENGDSR ERDEEQGDSR ERDEEQGDSR GG GG SG SG SG SG SG	RFYVFGPRSF RFYVFGPRSF RFYVFGPRSF RFYHLGEESF RFYHLGEESF RFYHFGEESF RFYHFGEESF RFYHFGEESF RFYHFGEESF RFYHFGEESF	RSIIRSDHGF RRIIRSDHGF RRIIQSDHGF RRIIQSDHGF RRIIQSDHGF RHWTRTRHGR RHWTRTRHGR REWAKSRHGH REWAKSRHGH REWAKSRHGH	VKALRPFDEV VKALRPFDEV VRALRPFDEV VRALRPFDQV VRALRPFDQV FSVLERFPDE FSVLERFPDE FSVLERFPDH FSVLERF-DH FSVLERF-DH FSVLERF-DH FSVLERF-DH	SRLLRGIRNY SRLLRGIRNY SRLLRGIRDY SRLLRGIRDY OVVGAAVGGY ELLRGSIGDY ELLRGSIGDY ELLRGSIGDY	VAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAIMEVNPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR RVAVLEAAPR

 Table A-4.1 Wheat EST alignments with cupin containing proteins (continued)

-	41					
Ta2154253404-2		DGVGYVAOGE	GVLTVIENCE	KRSYTVROGD	VIVAPAGS	IMHLANTDGR
Ta9905-2						
Ta3078253410-1	AFVVPGLTDA	DGVGYVAQGE	GVLTVIENGE	KR.SYTVREGD	VIVAPAGS	IMHLANTDGR
Ta3078253407-1	SEVVPGLTDA	DGVGYVAQGE	GVLTVIENGE	RRSYTVROGD	VIVAPAGS	IMHLANTDGR
Ta3078253412-1				VRSYTVROGR	MLIRCAPAGS	IMHLANTDOR
Ta3078253409-1				SAR-G-		TDGR
Ta3078253413-3	AFVVPGLTDA	DGVGYVAQGE	GVLTVIENGE	KRSYTVRQGD	VIVAPAGS	IMHLANTDGR.
Ta5446-1		YVAQGE	GVLTVIENGE	KRSYTVRQGD	VIVAPAGS	IMHLANTDGR.
AAA34269	AFVVPGFTDA	DGVGYVAQGE	GVLTVIENGE	KRSYTVKEGD	VIVAPAGS	IMHLANTDGR.
Q03678 HORVU	AFVVPGFTDA	DGVGYVAQGE	GVLTVIENGE	KRSYTVKEGD	VIVAPAGS	IMHLANTDGR
Ta194625-1						
Ta5441-3			<b>TTH</b>	ASAHTELASS	TMA	TRAR
Ta9894-2						
Q852L2_ORYSA	AFLQPSHYDA	DEVFYVKEGE	GVIVLLREGR	RESECVREGD	AMVIPAGA	IVYSANTHSS
Q8L8I0_ORYSA	AFLQPSHYDA	DEVFYVEGE	GVIVLLREGR	RESPONRED	AMVIPAGA	IVYSANTHSS
Ta628253404-2	AFLH <mark>PSHYD</mark> A	DEIAFVREGE	GVLVLL <mark>R</mark> NGK	RESECVREGD	VFVIPAGS	IVYSANTHRS
Ta628253407-1	AFLQPSHYDA	DEIAFVREGE	GVLVLLRNGK	RESECVREGD	VFVIPAGS	IVYSANTHRS
Ta136340-2	AFLOPSHHDA	DEIAFVREGE	GVLVLLRNGK	RESPOIREGD	VIVIPAGS	IVYSANTHRS
Ta627253413-2			S	R	LAS	IVYSANTHRS
Ta628253405-1	AFLOPSHHDA	DEIAFVREGE	GVLVLLRNGK	RESPOIREGD	VIVIPAGS	IVYSANTHRS
Ta132809-2						
Ta189317-3						
Ta2606253404-2	TLFVPQYIDS	NLILFVQRGD	VKVGWIHKGG	LVERQLEMGD	VLQIDAGS	TFYMVNTGRG
Ta2606253405-2	TLEVPOYIDS		VEVGWINEGG	LVENQLMMGD	VLQMDAGS	TFYMVNTGRG
Q70PK0_WHEAT	GNAVS		- VNG- HPCKP	MS KAGD	DFLFSS	KLA KAGNTST
ATEG_07804	TLASCORDD	ADLGVYLDFE	TVDNPQPIRG	ELGGTDPGPR	NYAYDRINSD	<b>KLAPPGTDS</b> G
2	01					
Ta2154253404-2	REVIANILH	TISVPGRF	QYFSAKP		SKRVLRAALK	TS-DEOLGRL
Ta9905-2						
Ta3078253410-1	RELIIANILH	TISVPGMF	OYFSAKP	LLASL	SKRVLRAALK	
Ta3078253407-1	RELVIANILH					
		TISVPGNF	OYFSARP	LLASL	SKRVLTAALK	TS-DERLERL TS-DEOLGRL
Ta3078253412-1	RKLVIAEILH	TISVPGNF TISVPGPF	QYFSAKP OYFSAKP			TS-DERLERL TS-DEQLGRL TS-DEOLGRL
Ta3078253412-1 Ta3078253409-1			QYFSAKP	<mark>LLA</mark> SL	SKRVLTAALK	TS-DEQLGRL
	RKLVIAEILH	TISVPGPF	QYFSAKP		SKRVLTAALK SKRVLTAALK	TS-DEQLGRL TS-DEQLGRL
Ta3078253409-1	RKLVIAEILH RKLVIAKILH	TISVPGPF TISVPGKF	QYFSAKP QYFSAKP	<mark>LLAS</mark> L <mark>LLAS</mark> L	SKRVLTAALK SKRVLTAALK SKRVLRAALK	TS-DEQLGRL TS-DEQLGRL TS-DEQLDRL
Ta3078253409-1 Ta3078253413-3	RKLVIAEILH RKLVIAKILH RKLVIAKILH	TISVPGPF TISVPGKF TISVPGKF	QYFSAKP QYFSAKP QYFSAKP QYFSA <mark>KP</mark>	LLASL LLASL LLASL	SKRVLTAALK SKRVLTAALK SKRVLRAALK SKRVLTAALK	TS-DEOLGRL TS-DEOLGRL TS-DEOLDRL TS-DEOLDRL TS-DERLGSL
Ta3078253409-1 Ta3078253413-3 Ta5446-1	RKLVIAEILH RKLVIAKILH RKLVIAKILH RKLVIAKILH	TISVPGEF TISVPGKF TISVPGKF TISVPGKF	QYFSAKP QYFSAKP QYFSAKP QYFSAKP	LLASL LLASL LLASL LLASL	SKRVLTAALK SKRVLTAALK SKRVLRAALK SKRVLTAALK SKRVLTAALK	TS-DEQLGRL TS-DEQLGRL TS-DEQLDRL TS-DERLGSL TS-DERLGSL
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269	RKLVIAEILH RKLVIAKILH RKLVIANILH RKLVIANILH RKLVIANILH	TISVPGFF TISVPGKF TISVPGKF TISVPGKF TISVPGKF	QYFSAKP QYFSAKP QYFSAKP QYFSAKP QFLSV <mark>KP</mark>	LLASL LLASL LLASL LLASL LLASL	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK	TS-DEOLGRL TS-DEOLGRL TS-DEOLDRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU	RKLVIAEILH RKLVIAKILH RKLVIAKILH RKLVIAKILH RKLVIAKILH RKLVIAKILH	TISVPGFF TISVPGKF TISVPGKF TISVPGKF TISVPGKF	QYFSAKP QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP	LLASL LLASL LLASL LLASL LLASL	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK	TS-DEOLGRL TS-DEOLGRL TS-DEOLDRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1	RKLVIAEILH RKLVIAKILH RKLVIAKILH RKLVIAKILH RKLVIAKILH RKLVIAKILH	TISVPGEP TISVPGKP TISVPGKP TISVPGKP TISVPGKP TISVPGKP	QYFSAKP QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP	LLASL LLASL LLASL LLASL LLASL	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK	TS-DEQLGRL TS-DEQLGRL TS-DEQLDRL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1 Ta5441-3	RKLVIAEILH RKLVIAKILH RKLVIAKILH RKLVIAKILH RKLVIAKILH RKLVIAKILH	TISVPGEP TISVPGKP TISVPGKP TISVPGKP TISVPGKP TISVPGKP	QYFSAKP QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP VSASHDE	LLASL LLASL LLASL LLASL LLASL EEDRR	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK	TS-DEQLGRL TS-DEQLGRL TS-DEQLDRL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL RC-HQDRPRY
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810_ORYSA	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG	TISVPGPF TISVPGKF TISVPGKF TISVPGKF TISVPGKF TISVPGKF TSLLFAAA	QYFSAKP QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP CFLSVKP GDP EEYFPVGG EEYFPVGG	LLASL LLASL LLASL LLASL LLASL LLASL 	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAAFK SKRVLRAAFK GGRSLQQCVQ GRHGEGEREE	TS-DEQLGRL TS-DEQLGRL TS-DEQLDRL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL RC-HQDRFRY HG-KHEQGR- TR-REELEKV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN	TISVPGRP TISVPGRP TISVPGRP TISVPGRP TISVPGRP TISVPGRP TISLLFAAA PVSTPGHF	QYFSAKP QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP VSASHDE GDP EEYFPVGG	LLASL LLASL LLASL LLASL LLASL EEDRR VVPGR DRPESFFSAF	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK GGRSLOOCVO GRHGEGEREE SDDVLQAAFN SDDVLQAAFN	TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL RC-HQDRPRY HG-KHEQGR- TR-REELEKV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q85212 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN	TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TSLLFAAA PVSTPGHF PVSTPGHF PVSTPGSF PVSTPGSF	QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP GDP EEYFPVGG EEYFPVGG QEFSPIGF3G QEFSPIGF3G	LLASL LLASL LLASL LLASL EEDRR VVPGR DRPESFFSAF DRPESFFSAF	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAAFK SKRVLRAAFK GGRSLOOCVO GRHGEGEREE SDDVLQAAFN SDDVLQAAFN	TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL SC-HQDRPRY HG-KHEQGR- TR-REELEKV TR-REELEKV TRQREDVDRV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253404-2 Ta628253407-1 Ta136340-2	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN	TISVPGPF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TSLLFAAA PVSTPGHF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF	QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP GDP EEYFPVGG EEYFPVGG QEFSPIGFGG QEFSPIGFGG QEFSPIGFGG	LLASL LLASL LLASL LLASL LLASL LLASL LLASL DRPESFFSAF DRPESFFSAF DRPESFFSAF EQPOSFFSVF	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK GGRSLQQCVQ GRHGEGEREE SDDVLQAAFN SDDVLQAAFN SDEVIQAAFN	TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL RC-HQDRPRY HG-KHEQGR- TR-REELEKV TR-REELEKV TR-REELEKV TRQREDVDRV SRRR-APAV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q8L810_ORYSA Ta628253404-2 Ta628253404-2 Ta6340-2 Ta627253413-2	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLN KWFRVVMLN	TISVPGPF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TSLLFAAA PVSTPGHF PVSTPGFF PVSTPGSF PVSTPGSF PVSTPGRF PVSTPGSF	QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP GDP GDP EEYFPVGG QEFSPIGFGG QEFSPIGFGG QEFSPIGFGG	LLASL LLASL LLASL LLASL LLASL LLASL LLASL LLASL DRPESFFSAF DRPESFFSAF EQPOSFFSVF EQPOSFFSVF	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAAFK GGRSLQQCVQ GRHGEGEREE SDDVLQAAFN SDEVIQAAFN SDEVIQAAFN	TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL RC-HQDRFRY HG-KHEQGR- TR-REELEKV TR-REELEKV TRQREDVDRV TRQREDVDRV TRQREDVDRV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q8L810_ORYSA Q8L810_ORYSA Ta628253404-2 Ta628253404-2 Ta628253405-1	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN	TISVPGPF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TSLLFAAA PVSTPGHF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF	QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP GDP EEYFPVGG EEYFPVGG QEFSPIGFGG QEFSPIGFGG QEFSPIGFGG	LLASL LLASL LLASL LLASL LLASL LLASL LLASL DRPESFFSAF DRPESFFSAF DRPESFFSAF EQPOSFFSVF	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAAFK GGESLQQCVQ GRHGEGEREE SDDVLQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAALN	TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL RC-HQDRFRY HG-KHEQGR- TR-REELEKV TR-REELEKV TRQREDVDRV SRRR-APAV TRQREDVDRV TRQREDVDRV TR-REDVDRV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q8L810_ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2 Ta628253405-1 Ta132809-2	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLN KWFRVVMLN	TISVPGPF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TSLLFAAA PVSTPGHF PVSTPGFF PVSTPGSF PVSTPGSF PVSTPGRF PVSTPGSF	QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP GDP GDP EEYFPVGG QEFSPIGFGG QEFSPIGFGG QEFSPIGFGG	LLASL LLASL LLASL LLASL LLASL LLASL LLASL LLASL DRPESFFSAF DRPESFFSAF EQPOSFFSVF EQPOSFFSVF	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK GGRSLQQCVQ GRHGEGEREE SDDVLQAAFN SDDVLQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN	TS-DEQLGRL TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL RC-HQDRPRY HG-KHEQGR- TR-REELEKV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2 Ta628253405-1 Ta132809-2 Ta189317-3	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLN KWFRVVMAPQ KWLRVVMPIN	TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF PVSTPGRF PVSTPGRF PVSTPGRF PVSTPGRF PVSTPGRF	QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP CFLSVKP CFLSVKP CFLSVKP CFLSVFVGG QEFSPIGFGG QEFSPIGFGG QEFSPIGFGG QEFFLIGSGD	LLASL LLASL LLASL LLASL LLASL LLASL LLASL LLASL DRPESFFSAF DRPESFFSAF DRPESFFSAF DRPESFFSAF DRPESFFSAF DRPESFFSAF DRPESFFSVF EQPQSFFSVF	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK SKRVLRAAFK SKRVLRAAFK SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN	TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL TS-DERLERL TS-DERLERL TS-REELERV TR-REELERV TR-REELERV TR-REELERV TR-REDVDRV SRRRAPAV TR-REDVDRV TR-REDVDRV TR-REDVDRV TR-REDVDRV TR-REDVDRV TR-REDVDRV TR-REDVDRV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253404-2 Ta628253405-1 Ta136340-2 Ta628253405-1 Ta132809-2 Ta189317-3 Ta2606253404-2	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLN KWFRVVMLN KWFRVVMSIN	TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TSLLFAAA PVSTPGHF PVSTPGHF PVSTPGRF PVSTPGRF PVSTPGRF PVSTPGRF PVSTPGRF PVSTPGRF	QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP GDP EEYFPVGG EEYFPVGG EEYFPVGG QEFSPIGFGG QEFSPIGFGG QEFSPIGFGG QEFFLIGSGD	LLASL LLASL LLASL LLASL LLASL LLASL LLASL LLASL DEPESFEAF DEPESFEAF DEPESFEAF DEPESFESFE DEPESFESFE DEPESFESFE DEPESFESFE DEPESFESFE	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK SKRVLRAAFK SKRVLRAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN	TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL TS-DERLERL TS-DERLERL TR-REELEKV TR-REELEKV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TR-REDVDRV TR-REDVDRV TR-REDVDRV TR-REDVDRV TR-REDVDRV TR-REDVDRV TR-REDVDRV TR-REDVDRV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2 Ta628253405-1 Ta132809-2 Ta132809-2 Ta189317-3 Ta2606253404-2 Ta2606253405-2	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLN KWFRVVMLN KWFRVVMLN KWFRVVMLSN KWLRVVMFIN	TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF PVSTPGHF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF	QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP GDP	LLASL LLASL LLASL LLASL LLASL LLASL LLASL LLASL DRPESFFSAF DRPESFFSAF DRPESFFSAF EQPQSFFSVF EQPQSFFSVF EQPQSFFSVF EQPQSFFSVF	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK GGRSLQQCVQ GRHGEGEREE SDDVLQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN	TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL TS-DERLERL TS-REELEKV TR-REELEKV TRQREDVDRV SRRR-APAV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TR-REDVDRV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q8L810_ORYSA Q8L810_ORYSA Ta628253404-2 Ta628253405-1 Ta136340-2 Ta628253405-1 Ta132809-2 Ta189317-3 Ta2606253404-2 Ta2606253405-2 Q70FK0_WHEAT	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLN KWFRVVMLN KWFRVVMLN KWFRVVMLN KWFRVVMPIN	TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TSLLFAAA PVSTPGHF PVSTPGFFF PVSTPGFFF PVSTPGFFF PVSTPGFFF PVSTPGFFF PVSTPGFFF PVSTPGFFF	QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP GDP	LLASL LLASL	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK GGRSLQQCVQ GRHGEGEREE SDDVLQAAFN SDDVLQAAFN SDEVIQAAFN	TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL TS-DERLERL TS-REELEKV TR-REELEKV TR-REELEKV TRQREDVDRV SRRAPAV TRQREDVDRV TR-REDVDRV
Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2 Ta628253405-1 Ta132809-2 Ta132809-2 Ta189317-3 Ta2606253404-2 Ta2606253405-2	RKLVIAEILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH RKLVIARILH VTIPLLFLLG KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLLN KWFRVVMLN KWFRVVMLN KWFRVVMLN KWFRVVMLSN KWLRVVMFIN	TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF TISVPGRF PVSTPGHF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF PVSTPGFF	QYFSAKP QYFSAKP QYFSAKP QFLSVKP QFLSVKP QFLSVKP GDP	LLASL LLASL LLASL LLASL LLASL LLASL LLASL LLASL DRPESFFSAF DRPESFFSAF DRPESFFSAF EQPQSFFSVF EQPQSFFSVF EQPQSFFSVF EQPQSFFSVF	SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLTAALK SKRVLRAAFK SKRVLRAAFK GGRSLQQCVQ GRHGEGEREE SDDVLQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN SDEVIQAAFN	TS-DEQLGRL TS-DEQLGRL TS-DERLGSL TS-DERLGSL TS-DERLGSL TS-DERLERL TS-DERLERL TS-DERLERL TS-REELEKV TR-REELEKV TRQREDVDRV SRRR-APAV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TRQREDVDRV TR-REDVDRV

 Table A-4.1 Wheat EST alignments with cupin containing proteins (continued)

,	61					
Ta2154253404-2		SRSISIIRAS	EBOLRELSRO	ASEGGOGHHW	PLPPFRGDSR	DTFNLLEORP
Ta9905-2						
Ta3078253410-1	LDPROGOENT	GGSMS IVRAS	EEQLHELSEQ	ASEGSOGHHW	PLPPFRGDSR	DTYNLLEORP
Ta3078253407-1	LFGRRQGQEE	ESSISIVEAS	EEQLRELRRQ	AS EGGO GHHW	PLPPFRGDSR	D'TFNLLEQRP
Ta3078253412-1	LFGRRQGQEE	ESSISIVEAS	EEQLRELRRQ	AS EGGO GHHW	PLPPFRGDSR	D'TFNLLEQRP
Ta3078253409-1	LFGRRQGQEE	EESISIVRAS	EEQLRELRRE	AS EGGO GHHW	PLPPFRGDSR	DTYNLLEORP
Ta3078253413-3	LGSROGKEEE	EKSISIVRAS	EEQLRELRRQ	ASEGDOGHHW	PLPPFRGDSR	D'TFNLLEORP
Ta5446-1	LGSRQE					
AAA34269	FNQRQ-GQEK	TRSVSIVRAS	EEQLRELRRE	AAEGGOGHRW	PLPPFRGDSR	DTFNLLEORP
Q03678_HORVU	FNORQ-GOEK	TRSVSIVRAS	ee <mark>olr</mark> elrre	AABGGOGHEW	PLPPFRGDSR	DTFNLLEORP
Ta194625-1	AR			GRGHGR	<mark>H</mark> GEGG	<b>R</b>
Ta5441-3	SHARC	VQECR	DE <mark>QQQH</mark> GRHE	QEEQGRGHGR	<mark>H</mark> GEGG	R
Ta9894-2	GRR				GEGE	R
Q852L2_ORYSA	FEROREG	GEITTAP	EEQIRELSKS	CSRGGGG	GSGSE-WEIK	PS-SLTGRSP
Q8L810_ORYSA	FEROREG	GEITTAP	EEQIRELSKS	CSRGGGG	GSGSE-WEIK	PP-SLTGKSP
Ta628253404-2	FORKSRGE	GQISEGS	EEQIRELSRS	CSRGGRGGGG	GSGSEKEDIQ	PR-SLTGERP
Ta628253407-1	FETKSRGQ	GQISBGS	EEQIRELSES	CSR GGR GGGG	GSG	
Ta136340-2	LLER					
Ta627253413-2	FORKSRGE	GPISEGS	EEQIRELSRS	CSRGGRGGGG	GSGSEKEDIQ	PR-SLTGEKP
Ta628253405-1	FESKSBGE	GEIYEAS	BEQIRBLSES	Cargerage	- SGSEKEDIQ	PR-SLTGEKP
Tal32809-2 Tal89317-3	FTSNAINC					
Ta2606253404-2	LPVKPLGPFV	SYTTESGG	KEHGOGDKRD	VGENGRESEP	WRPVGRGNDD	
Ta2606253405-2	LPVRTGGPFV	SYTTESGEGG	KEHGOGD	VGENGRESEP	WRFVGRGDDD	DD
Q70PK0 WHEAT	LLVGILGS		LDSGNKLYSR	VVRAGETFLI	PRGLMH	
ATEG 07804	CRICAIN		- ENGETFIDD	VT-EGDVWFF	PPGVPHS IOA	
ALDO_0/004						
—						
- 4	21					
4 Ta2154253404-2	21 MIANRHGRLY	QADARSFHAL	AQHDV <mark>R</mark> VAVA	NITEGSMTAP	YLNTQSF <mark>K</mark> LA	VVLEG-EGEV
-		QADARSFHAL	AQHDVRVAVA AQHDV-ASPG	NITPGSMTAP PTSREALDEA	VLNTQSFKLA	VVLEG- EGEV VVLEG- EGEV
Ta2154253404-2	<b>NIANR</b> HGRLY	HAS EADARSFHAL				
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1	KIANRHGRLY RIANRHGRLY KIANRHGRLF	EADARSFHAL EADARSFHAL	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA	PTSRRALDRA NITPGSMTAP NITPGSMTAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253407-1	KIANRHGRLY RIANRHGRLY KIANRHGRLF KIANRHGRLF	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA	PTSREALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA	VVLBG-BGEV VVLBG-BGEV VVLBG-BGEV VVLBG-QGEV
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1	KIANRHGRLY RIANRHGRLY KIANRHGRLF KIANRHGRLF KIANRHGRLY	HAS EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA	PTSERALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-OGEV VVLEG-EGEV
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-3	KIANRHGRLY RIANRHGRLY KIANRHGRLF KIANRHGRLF	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA	PTSREALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA	VVLBG-BGEV VVLBG-BGEV VVLBG-BGEV VVLBG-QGEV
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253412-1 Ta3078253412-3 Ta5446-1	KIANRHGRLY RIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY	HAS EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL	ACHDV-ASPG ACHDVRVAVA ACHDVRVAVA ACHDVRVAVA ACHDVRVAVA ACHDVRVAVA	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269	KIANRHGRLY RIANRHGRLF KIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL	AQHDV ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-QGEV VVLEG-EGEV VVLEG-EGEV
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU	KIANRHGRLY RIANRHGRLF KIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY	HAS EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1	KIANRHGRLY RIANRHGRLY KIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEDQGR	HAS EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL GRGRHGQ	ACHDV-ASPG ACHDVRVAVA ACHDVRVAVA ACHDVRVAVA ACHDVRVAVA ACHDVRVAVA ACHDVRVAVA ACHDVRVAVA ANCDVRVAVA GEREEEQGRG	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3	KIANRHGRLY RIANRHGRLY KIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEDQGR- - EEDQGR-	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL GRGRHGQ 	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA GEREEEQGRG GEREEEQGRG	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGOGER- GGRHGEGER-	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA EEEDGRG	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV RGRHG-EGER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2	KIANRHGRLY RIANRHGRLY KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEEQGR - DEEHGD	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL GRGRHGQ 	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA ANQDVRVAVA GEREEQGRG GEREEEGGRG GEREEEGGRG GERNFRSIIR	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGQGER- SCHGEGER- S-DHGFVKAL	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA EEDQGRG RFFDEVSRLL	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGER RGRHG-QEER RGRHG-QEER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9494-2 Q852L2_ORYSA	KIANRHGRLY RIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEEQGR - EEEQGR - DEEHGD YFSNNHGRLF	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL 	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA ANQDVRVAVA GEREEEQGRG GEREEEGGRG GEREEEGGRG GEREEEGGRG GEREEEGGRG	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGGGER- RGRHGGGER- S-DHGFVKAL NITRGSMIAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA 	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV RGRHG-EGER RGRHG-EGER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta984-2 Q852L2 ORYSA Q8L810 ORYSA	KIANRHGRLY RIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY CEEDQGR - EEEQGR - DEEHGD YFSNNHGRLF YFSNNHGRLF	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL 	AQHDV ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGOGER- RGRHGDGER- S-DHGFVKAL NITRGSMIAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA 	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV RGRHG-QEER RGRHG-QEER RGRHG-EGER S
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8120 ORYSA Ta628253404-2	KIANRHGRLY RIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEEQGR - EEEQGR - DEEHGD YFSNNHGRLF	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL 	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA ANQDVRVAVA GEREEEQGRG GEREEEGGRG GEREEEGGRG GEREEEGGRG GEREEEGGRG	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGGGER- RGRHGGGER- S-DHGFVKAL NITRGSMIAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA 	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV RGRHG-EGER RGRHG-EGER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q852L2_ORYSA Q852L2_ORYSA Ta628253404-2 Ta628253407-1	KIANRHGRLY RIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY CEEDQGR - EEEQGR - DEEHGD YFSNNHGRLF YFSNNHGRLF	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL 	AQHDV ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGOGER- RGRHGDGER- S-DHGFVKAL NITRGSMIAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA 	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV RGRHG-QEER RGRHG-QEER RGRHG-EGER S
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L010 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2	KIANRHGRLY RIANRHGRLY KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEEQGR - DEEHGD YFSNNHGKLF YFSNNHGKLF RYSNNHGRFH	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL 	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GERNFRSIIR KKLDLQIGLA KKLDLQIGLA	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGQGER- S-DHGFVMAL NITRGSMIAP NITRGSMIAP NITRGSMIAL	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA 	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGER RGRHG-EGER RGRHG-EGER RGRHG-EGER RUUQG-SGYP VVLQG-SGYP
Ta2154253404-2 Ta9905-2 Ta3078253407-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2	KIANRHGRLY RIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY CEEDQGR - EEEQGR - DEEHGD YFSNNHGRLF YFSNNHGRLF	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL - GRGRHGQ - GRGRHGQ - GRGRHGQ - SRRFYVF ELTGDECRHL QITGDQCHHL	AQHDV ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGOGER- RGRHGDGER- S-DHGFVKAL NITRGSMIAP	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA 	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV RGRHG-QEER RGRHG-QEER RGRHG-QEER RGRHG-GER VVLQG-SGYP VVLQG-SGYP
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L010 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2	KIANRHGRLY RIANRHGRLY KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEEQGR - DEEHGD YFSNNHGKLF YFSNNHGKLF RYSNNHGRFH	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL 	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA GEREEQGRG GEREEQGRG GEREEQGRG GERNFRSIIR KKLDLQIGLA KKLDLQIGLA RKLDMDVTLV	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGQGER- S-DHGFVKAL NITRGSMIAP NITRGSMIAP NITRGSMTAL	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA 	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV RGRHG-EGER RGRHG-EGER S
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2 Ta628253405-1	KIANRHGRLY RIANRHGRLY KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEEQGR - DEEHGD YFSNNHGKLF YFSNNHGKLF RYSNNHGRFH	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL - GRGRHGQ - GRGRHGQ	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GERE GERE	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGGGER- S-DHGFVKAL NITRGSMIAP NITRGSMIAP NITRGSMTAL	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA 	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV RGRHG-EGER RGRHG-EGER S
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253412-1 Ta3078253412-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta628253405-1 Ta132809-2	KIANRHGRLY RIANRHGRLF KIANRHGRLF KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY CEEDOGR - EEEOGR - DEEHGD - DEEHGD YFSNNHGRLF YFSNNHGRLF RYSNNHGRFH RYSNNHGRFH	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL CADARSFHAL	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA ANQDVRVAVA GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GEREEEQGRG GERE GERE	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGOGER- S-DHGFVBAL NITRGSMIAP NITRGSMIAP NITRGSMTAL NITRGSMTAL NITRGSMTAL	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA OLNTQSFKLA PLNTQSFKLA PLNTQSFKLA NYNTRATKLA NYNTRATKLA NYNTRATKLA NYNTRATKLA NYNTRATKLA NYNTRATKLA NYNTRATKLA NYNTRATKLA	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV RGRHG-EGER RGRHG-EGER S
Ta2154253404-2 Ta9905-2 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q85212 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2 Ta628253405-1 Ta132809-2 Ta189317-3	KIANRHGRLY RIANRHGRLY KIANRHGRLF KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEEQGR - DEEHGD YFSNNHGKLF YFSNNHGKLF RYSNNHGRFH	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL CADARSFHAL	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA GEREEEQGRG GEREEEQGRG GEREEEQGRG GFRNFRSIIR KKLDLQIGLA KKLDLQIGLA KKLDLQIGLA KKLDMDVTLV RKLDMDVTLV -KSEAYSSVS -TSEAYSSVS	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGOGER- RGRHGEOGER- S-DHGFVRAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA 	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGER RGRHG-EGER RGHQ VVLQG-SGYF IVVEGRDGYF IVVEGRDGYF ILRQGFGVSA
Ta2154253404-2 Ta9905-2 Ta3078253407-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta5446-1 AA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q81810_ORYSA Ta628253404-2 Ta628253404-2 Ta628253405-1 Ta132809-2 Ta189317-3 Ta2606253404-2	KIANRHGRLY RIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEDQGR - DEEHGD YFSNNHGRLP YFSNNHGRLP RYSNKHGRPH RYSNKHGRPH RYSNKHGRPH	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL CADARSFHAL	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA GEREEEQGRG GEREEQGERG KKLDMDVTLV RKLDMDVTLV FKLDMDVTLV FKLDMDVTLV FKLDMDVTLV FKLDMDVTLV FKLDMDVTLV FKLDMDVTLV FKLDMDVTLV	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGOGER- RGRHGEGER- S-DHGFVKAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA EEEQGRG EEEQGRG RPFDEVSRLL NYNTRATKLA RYTTRSTRIY RYTTRSTRIY RYTTRSTRIY RYTTRSTRIY FYNLFDHEP	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLQG-SGYP VVLQG-SGYP IVVEGRDGYF VVVEGRDGYF VVVEGRDGYF ILRQGFGVSA SFRNTYGKSI
Ta2154253404-2 Ta9905-2 Ta3078253407-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253405-1 Ta136340-2 Ta628253405-1 Ta132809-2 Ta189317-3 Ta2606253404-2 Ta2606253405-2	KIANRHGRLY RIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY KIANRHGRLY - EEDQGR - DEEHGD YFSNNHGRLP YFSNNHGRLP RYSNKHGRPH RYSNKHGRPH RYSNKHGRPH	EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL EADARSFHAL CADARSFHAL	AQHDV-ASPG AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA AQHDVRVAVA GEREEQGRG GEREEQGRG GEREEQGRG GEREEQGRG GEREEGGRG GERNFRSIIR KKLDLQIGLA KKLDLQIGLA KKLDMDVTLV RKLDMDVTLV RKLDMDVTLV SEAYSSVS -TSEAYSSVS MSRFIGGELN	PTSRRALDRA NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP NITPGSMTAP HGRHGQGER- SCHGFVGAL NITRGSMIAP NITRGSMIAP NITRGSMIAP NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL NITRGSMTAL	VLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA YLNTQSFKLA EEEQGRG PFDEVSRLL NYNTRATKLA RYTTRSTRIY RYTTRSTRIY FEVK EPYNLFDHEP	VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV VVLEG-EGEV SGRHG-QEER SGRHG-QEER SGRHG-EGER VVLQG-SGYP IVVEGRDGYF IVVEGRDGYF ILRQGFGVSA SFRNTYGWSI SFRNTYGWSI

 Table A-4.1 Wheat EST alignments with cupin containing proteins (continued)

4	81					
Ta2154253404-2	EIVCPHLGRD	SERREHGK	- GRWS EEEED	DRRQQ		RRHGSGSESE
Ta9905-2	EIVCPHLGRD	SERREHGK	- GRWS EEEED	DRROO		RRHGSGSESE
Ta3078253410-1	QIVCPHLGRD	SERREQGK		DQRQQ		RRRG <mark>S</mark> G <mark>SES</mark> E
Ta3078253407-1	QIVCPHLGQD	SERE-HEHGK		DOROO		RRRGSGSESE
Ta3078253412-1	QIVCPHLGOD	SERE-HEHGK	- GRRS EEEED	DORCO		R
Ta3078253409-1	QIVCPHLARD	SERHKOGK		AQRQQ		RRRGSGSESE
Ta3078253413-3 Ta5446-1	EIVCPHLGRD	SERRECEHGK	GRWRSEEEED	DRRQQ		RRRGSGSESE
AAA34269	QIVCPHL/GRE	SESE-REHCK	- GRRREEEED	DOBCO		RRRGSESESE
003678 HORVU	OIVCPHLGRE	SESE-REHGK	- GRRREEEED	DOBOO		RRRGSESESE
Ta194625-1	EEHGRRE	OEEOGOGR	- GRRGEGERD	EEHGD		SRR
Ta5441-3	EEHGKHE	QGR	- GRRGEGERD	EE <mark>H</mark> GD		SRR
Ta9894-2	ELPRRHHGSE	PGL	VRRAGTHGRG	RRR <mark>LR</mark>		RSRRGGAD
Q852L2_ORYSA	EMACPHVS GG	G <mark>SSE-RRE</mark>	REREHG	RRREE		- EQGEEE
Q8L8I0_ORYSA	EMACPHVSGG	GSSE-RRE	REREHG	RRREE		- EQGEEE
Ta628253404-2	EMACPHVSSS	GRSE-RREHE	<mark>Q BR BR BH</mark> GHG	<u>rrseer</u> goeh		- GRRS BEEDH
Ta628253407-1 Ta136340-2						
Ta627253413-2	EMACPHVSSF	GRSE-RREHE	OERERENGHG	RRSEEREREH	GOGRRSEERK	DECGROEEEO
Ta628253405-1	EMACPHISSS	GRSE-RREHE	GEREREHGOG	RRSEEREREQ		OBOGROEEEO
Ta132809-2	EVVEAIOSAN	SPOSI	ITYNPDOEKE			
Ta189317-3	EVVEAIRSAK	SPOSI	ITYNPDQKKE	EKS		
Ta2606253404-2	SVDKHDYHPL	DHSDIGVYLV	NLTAGSMMAP	HVNPR		ATEYGVVLAG
Ta2606253405-2	SVDKHDYEPL	DHSDIGVYLV	NLTAGSMMAP	HVNPR		ATEYGVVLAG
Q70PK0_WHEAT	PIPTPVL	TRALEVEAGV	VEL			
ATEG_07804	PGTPAPKDIE	BONVTTAAGV	VPLEDSYSYH	FSEQPA		HQVAGGSVEI
5	41					
-	41 SEEEODOO	RYETVRARVS	RGSAFVVPPG	HPVVEIS-SS	OGSSNLOVVC	FEINAERNER
5 Ta2154253404-2 Ta9905-2	41 SEEEODOO SEEEODOO	RYETVRARVS RYETVRARVS	RG <mark>SAFVV</mark> PPG RGSAFVVPPG	H <mark>PVVE</mark> IS-SS HPVVEIS-SS	QGSSNLQVVC QGSSNLQVVC	FEINAERNER FEINAERNER
Ta2154253404-2	SEEEQDQQ					
Ta2154253404-2 Ta9905-2	SEEEODQQ SEEEODQQ	RYETVRARVS	RG <mark>SAFVV</mark> PPG	HPVVEIS-SS	<b>QGSSNLQVV</b> C	FEINAERNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1	SEEEQDQQ SEEEQDQQ SESEEQQDQQ EEQDQQ	RYETVRARVS RYQTIRARVS RYETVRARVS	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG	H <mark>PVVEIS-SS</mark> HPVVEIA-SS HPVVEIA-SS	OGSSNLOVVC OGSSNLOVVC RGSSNLOVVC	FEINAERNER FEINAERNER FEINAERNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1	SE EEQDQQ SE EEQDQQ SESEEQQDQQ EEQDQQ SS- EEEQDQQ	RYETVRARVS RYQTIRARVS RYETVRARVS	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS	QGSSNLQVVC QGSSNLQVVC RGSSNLQVVC RGSSNLQVVC	FEINAERNER FEINAERNER FEINAERNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3	SEEEQDQQ SEEEQDQQ SESEEQQDQQ EEQDQQ	RYETVRARVS RYQTIRARVS RYETVRARVS	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG	H <mark>PVVEIS-SS</mark> HPVVEIA-SS HPVVEIA-SS	GSSNLQVVC GSSNLQVVC RGSSNLQVVC RGSSNLQVVC RGSSNLQVVC	FEINAERNER FEINAERNER FEINAERNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1	SEEE0D00 SE-EE0D00 SESED0000 EE0D00 SS-EE0D00 EE	RYETVRARVS RYQTIRARVS RYETVRARVS RYETVRARVS RYETVRARVS	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS	QGSSNLQVVC QGSSNLQVVC RGSSNLQVVC RGSSNLQVVC RGSSNLQVVC	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269	SEEE0D00 SE-EE0D00 SESEB00D00 EE0D00 SS-EEE0D00 EE0D00 EEE000	RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS	QGSSNLQVVC QGSSNLQVVC RGSSNLQVVC RGSSNLQVVC RGSSNLQVVC	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU	SEEE0D00 SE-EE0D00 SESED0000 EE0D00 SS-EE0D000 EE	RYETVRARVS RYQTIRARVS RYETVRARVS RYETVRARVS RYETVRARVS	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS	OGSSNLOVVC OGSSNLOVVC RGSSNLOVVC RGSSNLOVVC RGSSNLOVVC OGSSNLOVVC	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269	SEEE0D00 SE-EE0D00 SESEB00D00 EE0D00 EE0D00 EEE00 EEE00	RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS	QGSSNLQVVC QGSSNLQVVC RGSSNLQVVC RGSSNLQVVC RGSSNLQVVC	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1	SEEE0D00 SE-EE0D00 SESEB00D00 EE0D00 EE0D00 EE0D00 EEE00 EEE00	RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GPRSFRS I IR	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVKAL	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL	CGSSNLOVVC CGSSNLOVVC RGSSNLOVVC RGSSNLOVVC CGSSNLOVVC CGSSNLOVVC RGIRNYRVAI	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA	SEEEQDQQ SESEDQQDQQ SESEDQQDQQ EEQDQQ EEQDQQ EEEEQQ EEEEQQ EEEEQQ 	RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GPRSFRS IIR GPRNFRS IIR RREAVLLROA RYHKVRAQVR	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVKAL- SDHGFVKAL- RR- EESVIVIPAS	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL RPFDEVSRLL HPATIVA-	QGSSNLQVVC QGSSNLQVVC RGSSNLQVVC RGSSNLQVVC QGSSNLQVVC QGSSNLQVVC QGSSNLQVVC RGIRNYRVAI RGIRNYRVAI RGIRNYRVAI RGAG	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV MEVNPRAFVV HHAPGQHRRP FFVGANHDEK
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta984-2 Q852L2 ORYSA Q8L810_ORYSA	SEEEQDQQ SE-EEQDQQQ SESEDQDQQ EEQDQQ EEQDQQ EEQDQQ EEEQQQ EEEQQQ EEEQQQ EEEQQQ FYVP 	RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GPRSFRSIIR GPRNFRSIIR RREAVLIROA RYENVRAQVR	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVRAL- SDHGFVRAL- RE- EESVIVIPAS EGSVIVIPAS	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL HPATIVA	QGSSNLQVVC QGSSNLQVVC RGSSNLQVVC RGSSNLQVVC QGSSNLQVVC QGSSNLQVVC QGSSNLQVVC QGSSNLQVVC RGIRNYRVAI RGIRNYRVAI RGAG-GV SEGESLAVVC	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV MEVNPRAFVV HHAPGOHRRP FFVGANHDEK
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2	SEEEQDQQ SESEDQQDQQ SESEDQQDQQ EEQDQQ EEQDQQ EEEEQQ EEEEQQ EEEEQQ 	RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GPRSFRS IIR GPRNFRS IIR RREAVLLROA RYHKVRAQVR	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVRAL- SDHGFVRAL- RE- EESVIVIPAS EGSVIVIPAS	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL RPFDEVSRLL HPATIVA-	QGSSNLQVVC QGSSNLQVVC RGSSNLQVVC RGSSNLQVVC QGSSNLQVVC QGSSNLQVVC QGSSNLQVVC RGIRNYRVAI RGIRNYRVAI RGIRNYRVAI RGAG	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV MEVNPRAFVV HHAPGQHRRP FFVGANHDEK
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q852L2_ORYSA Q852L2_ORYSA Ta628253404-2 Ta628253407-1	SEEEQDQQ SE-EEQDQQQ SESEDQDQQ EEQDQQ EEQDQQ EEQDQQ EEEQQQ EEEQQQ EEEQQQ EEEQQQ FYVP 	RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GPRSFRSIIR GPRNFRSIIR RREAVLIROA RYENVRAQVR	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVRAL- SDHGFVRAL- RE- EESVIVIPAS EGSVIVIPAS	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL HPATIVA	QGSSNLQVVC QGSSNLQVVC RGSSNLQVVC RGSSNLQVVC QGSSNLQVVC QGSSNLQVVC QGSSNLQVVC QGSSNLQVVC RGIRNYRVAI RGIRNYRVAI RGAG-GV SEGESLAVVC	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV MEVNPRAFVV HHAPGOHRRP FFVGANHDEK
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q8L810_ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2	SEEE0D00 SE-EE0D00 SESED0000 SS-EE00D00 EE0D00 EE0D00 EEE000 EEE000 EEE000 	RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GPRSFRSIIR GPRNFRSIIR RREAVLLROA RYHKVRAQVR RYHKVRAQVR GYRQVRAQIK	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVKAL- SDHGFVKAL- RR EGSVIVIPAS EGSVIVIPAS VGSVIVLPAG	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL HPATIVA	QGSSNLQVVC QGSSNLQVVC RGSSNLQVVC RGSSNLQVVC QGSSNLQVVC QGSSNLQVVC QGSSNLQVVC RGIRNYRVAI RGIRNYRVAI RGIRNYRVAI RGAG-GV SEGESLAVVC GNEGNLALLS	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV HHAPGQHRRP FFVGANHDEK FFVGANHDEK FFVGANHDEK
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2		RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GPROFESIE GPROFESIE RREAVLLEOA RYHEVRAOVR GYROVRAOIK	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVKAL-	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL HPATIVA	GSSNLQVVC GSSNLQVVC RGSSNLQVVC RGSSNLQVVC GSSNLQVVC GSSNLQVVC GSSNLQVVC RGIRNYRVAI RGIRNYRVAI RGIRNYRVAI RGAG-GV SEGESLAVVC SEGESLAVVC GNEGNLALLS	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV HHAPGOHRR FFVGANHDEK FFVGANHDEK FFVGANNDEE
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q8L810_ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2	SEEE0D00 SE-EE0D00 SESED0000 SS-EE00D00 EE0D00 EE0D00 EEE000 EEE000 EEE000 	RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GPRSFRSIIR GPRNFRSIIR RREAVLLROA RYHKVRAQVR RYHKVRAQVR GYRQVRAQIK	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVKAL-	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL HPATIVA	QGSSNLQVVC QGSSNLQVVC RGSSNLQVVC RGSSNLQVVC QGSSNLQVVC QGSSNLQVVC QGSSNLQVVC RGIRNYRVAI RGIRNYRVAI RGIRNYRVAI RGAG-GV SEGESLAVVC GNEGNLALLS	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV HHAPGQHRRP FFVGANHDEK FFVGANHDEK FFVGANHDEK
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1 Ta3078253409-1 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q8L810_ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta627253413-2 Ta628253405-1		RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GPROFESIE GPROFESIE RREAVLLEOA RYHEVRAOVR GYROVRAOIK	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVKAL-	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL HPATIVA	GSSNLQVVC GSSNLQVVC RGSSNLQVVC RGSSNLQVVC GSSNLQVVC GSSNLQVVC GSSNLQVVC RGIRNYRVAI RGIRNYRVAI RGIRNYRVAI RGAG-GV SEGESLAVVC SEGESLAVVC GNEGNLALLS	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV HHAPGOHREP FFVGANHDEK FFVGANHDEK FGVGANNDEE FGVGANNDEE
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253407-1 Ta136340-2 Ta628253405-1 Ta132809-2		RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GPROFESIE GPROFESIE RREAVLLEOA RYHEVRAOVR GYROVRAOIK	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVKAL-	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL 	GSSNLQVVC GSSNLQVVC RGSSNLQVVC RGSSNLQVVC GSSNLQVVC GSSNLQVVC GSSNLQVVC RGIRNYRVAI RGIRNYRVAI RGIRNYRVAI RGAG-GV SEGESLAVVC SEGESLAVVC GNEGNLALLS	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV HHAPGOHREP FFVGANHDEK FFVGANHDEK FGVGANNDEE FGVGANNDEE
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2_ORYSA Q852L2_ORYSA Q852L2_ORYSA Q852L2_ORYSA Za628253404-2 Ta628253405-1 Ta132809-2 Ta182817-3 Ta2606253404-2 Ta2606253405-2	SEEEODOO SE-EEODOO SESEDOODOO SESEDOODOO EEODOO EEODOO EEOOO EEEOOO EEEOOO EEEOOO FYVF 	RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GYRSFRSIIR GYRSFRSIIR RYETVRARVS GYRSFRSIIR RYHKVRAQVR GYRQVRAQIK GYRQVRAQIK GYRQVRAQIK GYRQVRAQIK RYQR	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVKAL- SDHGFVKAL- SDHGFVKAL- RE- SDHGFVKAL- RE- VGSVIVIPAS VGSVIVLPAG VGSVIVLPAG	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL 	GSSNLQVVC GSSNLQVVC RGSSNLQVVC RGSSNLQVVC SEGSSNLQVVC SEGSSNLQVVC SEGSSNLQVVC SEGSSNLQVVC SEGSSNLQVVC SEGSSNLQVVC SEGSSNLQVVC SEGSSNLQVVC SEGSSNLQVC SEGSSNLQVVC SEGSSNLQVVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVVC SEGSSNLQVC	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV HHAPGOHRRP FFVGANHDEK FFVGANHDEK FGVGANNDEE FGVGANNDEE
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810 ORYSA Ta628253404-2 Ta628253404-2 Ta628253407-1 Ta136340-2 Ta628253405-1 Ta132809-2 Ta189317-3 Ta2606253404-2		RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS RYETVRARVS GYRSFRSIIR GPRSFRSIIR GPRNFRSIIR RREAVLUROA RYHKVRAQVR GYRQVRAQIK GYRQVRAQIK GYRQVRAQIK GYRQVRAQIK GYRQVRAQIK GYRQVRAQIK GYRQVRAQIK GYRQVRAQIK	RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG RGSAFVVPPG SDHGFVKAL- SDHGFVKAL- SDHGFVKAL- SDHGFVKAL- RE- EGSVIVIPAS VGSVIVLPAG VGSVIVLPAG VGSVIVLPAG	HPVVEIS-SS HPVVEIA-SS HPVVEIA-SS HPVVEIA-SS HPVVEIS-SS HPVVEIS-SS RPFDEVSRLL RPFDEVSRLL HPATIVA	GSSNLQVVC GSSNLQVVC RGSSNLQVVC RGSSNLQVVC SEGSLAVVC SEGSSLQVVC SEGSSLQVC SEGSSLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNLQVC SEGSSNC SEGSSNLQVC SEGSSNC SEGSSNLQVC SEGSSNC	FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER FEINAERNER MEVNPRAFVV MEVNPRAFVV HHAPGOHRRP FFVGANHDEK FFVGANHDEK FGVGANNDEE FGVGANNDEE

 Table A-4.1 Wheat EST alignments with cupin containing proteins (continued)

6	01					
Ta2154253404-2	VWLAGRNNVI		TFGRPAREVQ	EVFRAKDOOD	EG <mark>FVA</mark> GPEQQ	SHEQEQERG-
Ta9905-2	VWLAGRNNV I		TFGRPAREVQ	EVFRANDOOD	EGFVAGPEQQ	
Ta3078253410-1 Ta3078253407-1	VWLAGRNNVI VWLAGRNNVI		TFGRPAREVQ TFGRPAREVO	EVFRANDQQD EVFRANDQQD	EGFVAGPEQQ EGFVAGPEOO	
Ta3078253407-1 Ta3078253412-1	WILR SEMAVI	GRUDNEAVED	IF GREARENVY	EVERAL VVD	EVOL ANOT EVOL	EVERGURKR
Ta3078253409-1	VWLAGENNVI	GKLDNPAQEL	TEGREAREVO	EVFRANDOOG	RR-ASSPDPS	SETESAGTA-
Ta3078253413-3	VWLAGRNNVI	AKLDDPAQEL	AFGRPAREVQ	EVFRANDQQD	EG <mark>FVA</mark> GPE <mark>QQ</mark>	QEHERG-
Ta5446-1						
AAA34269 003678 HORVU	VWLAGRNNVI VWLAGRNNVI	GKLGSPAQEL GKLGSPAQEL	TFGRPAREVQ TFGRPAREVQ	EVFRAQDQ-D EVFRAQDQ-D	EGFVAGPEQQ EGFVAGPEQQ	SREQEQEQER SREQEOEOER
Ta194625-1	PGLTDADG-V	GYVAOGEGVL	TVIENGEK	RSYTVRE		
Ta5441-3	PGLTDADG-V	GYVAQGEGVL	TVIENGER	RSYTVROGDV	IVAPAGS IMH	LANTDGREEL
Ta9894-2	EEAGHRODPP	HHLRAROVEV	FLGQASSREF	EQTRAESGSO	DLG	
Q852L2_ORYSA	VFLAGENSPL	ROLDDPAKKL	VFGGSAAREA	DRVLAAQPEQ	ILLRGPHGRG ILLRSPHGRG	
Q8L8I0_ORYSA Ta628253404-2	VFLAGENSPL VFVTGGNSVL	ROLDDPAKKL KOLDEAAKAL	VFGGSAAREA AFPOOARELA	DRVLAAQPEQ DRVIRAOPES	VFVPGPOOOR	
Ta628253407-1						
Ta136340-2						
Ta627253413-2	VFVTGGNSVL					
Ta628253405-1 Ta132809-2	VFVTGGNSVL EERERREEKE	KOLDEAAKAL RKOREEEERA	SFPQQARELA RREKERKERE			RVADM
Ta189317-3	NWTEEIF	DALWGDESPL		CCCHHCK CVC		
Ta2606253404-2	QFLTGPTSVL	RMML/GPELAA	GL/GVPEKE	LKDVMEAQKV	AVIEPPLPEK	EKGGKEREPF
Ta2606253405-2	QFLTGPTSVF		GL/GVPENEKE	LREVVEAOTV		
Q70PK0 WHEAT ATEG 07804			DEELMFLEVL			
AIE/3 0/804	THOD VOIL FV	STORITERIO	DEPORT DEVE	CHROLIDIHR	OCUTODIC DC	TANDIDEDEE
=						
-	61		_			
Ta2154253404-2	DRRRGDRGRG	DDAV <mark>GAFLR</mark> M				
Ta2154253404-2 Ta9905-2	DRRRGDRGRG DRRRGD					
Ta2154253404-2	DRRRGDRGRG DRRRGD DRRRGDRGRG		ATAAL			
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1	DRRRGDRGRG DRRRGD DRRRGDRGRG GDR	DEAVEAFL <mark>R</mark> M	ATAAL			
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1	DRRRGDRGRG DRRRGD DRRRGDRGRG GDR	DEAVEAFLRM	ATAAL			
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1 Ta3078253409-1	DRRRGDRGRG DRRRGD DRRRGDRGRG GDR AVVTAAAAATK DRRRGDRGRG	DEAVEAFL <mark>R</mark> M	ATAAL			
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1	DRRRGDRGRG DRRRGD	DEAVEAFLEM PWEPS- DEAVEAFLEM	ATAAL			
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU	DRRRGDRGRG DRRRGD- DRRRGDRGRG GDR- AVVTAAAATK DRRRGDRGRG HRRRGDRGRG HRRRGDRGRG	DEAVEAFLEM PWEPS DEAVEAFLEM DEAVETFLEM DEAVETFLEM	ATAAL ATAAL ATGAI ATGAI			
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1	DRRRGDRGRG DRRRGD DRRRGDRGRG GDR AVVTAAAATK DRRRGDRGRG HRRRGDRGRG HRRRGDRGRG	DEAVEAFLRM PWRPS- DEAVEAFLRM DEAVETFLRM DEAVETFLRM	ATAAL ATAAL ATGAI ATGAI			
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678_HORVU	DRRRGDRGRG DRRRGD- DRRRGDRGRG GDR- AVVTAAAATK DRRRGDRGRG HRRRGDRGRG HRRRGDRGRG	DEAVEAFLRM PWRPS- DEAVEAFLRM DEAVETFLRM DEAVETFLRM	ATAAL ATAAL ATGAI ATGAI			
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253412-1 Ta3078253409-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3	DRRRGDRGRG DRRRGD DRRRGDRGRG GDR AVVTAAAATK DRRRGDRGRG HRRRGDRGRG HRRRGDRGRG	DEAVEAFLRM PWRPS- DEAVEAFLRM DEAVETFLRM DEAVETFLRM	ATAAL ATAAL ATGAI ATGAI			
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253412-1 Ta3078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q852L2 ORYSA Q8L810_ORYSA	DRRRGDRGRG DRRRGD DRRRGDRGRG GDR AVVTAAAATK DRRRGDRGRG HRRRGDRGRG HRRRGDRGRG	DEAVEAFLRM PWRPS- DEAVEAFLRM DEAVETFLRM DEAVETFLRM	ATAAL ATAAL ATGAI ATGAI			
Ta2154253404-2 Ta9905-2 Ta3078253410-1 Ta3078253407-1 Ta3078253409-1 Ta3078253409-1 Ta5078253413-3 Ta5446-1 AAA34269 Q03678 HORVU Ta194625-1 Ta5441-3 Ta9894-2 Q85212 ORYSA Q8L810 ORYSA Ta628253404-2	DRRRGDRGRG DRRRGD DRRRGDRGRG GDR AVVTAAAATK DRRRGDRGRG HRRRGDRGRG HRRRGDRGRG	DEAVEAFLRM PWRPS- DEAVEAFLRM DEAVETFLRM DEAVETFLRM	ATAAL ATAAL ATGAI ATGAI			
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 Table A-4.1 Wheat EST alignments with cupin containing proteins (continued)

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Plant genome database

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