Structural characterisation of yeast Lsm protein complexes

Jens Moll

Master of Science (Biotechnology)

A thesis presented to Macquarie University in fulfillment of the requirements for the degree of Doctor of Philosophy

Department of Chemistry & Biomolecular Sciences,

Macquarie University, Sydney NSW

Australia

August 2011

Content

Acknowledgements		vii	
<u>Decla</u>	aration	viii	
<u>Abstı</u>	ract	ix	
<u>List o</u>	of abbreviations	X	
1 Inti	roduction	1	
1.1	The Lsm proteins: Ring architectures for RNA capture	1	
1.1.1	Introduction: life cycle of mRNA	1	
1.1.2	Functional roles for Lsm proteins	3	
1.1.3	Specific functions of bacterial Hfq	7	
1.1.4	Lsm proteins in human disease and viral replication	8	
1.1.5	Phylogeny of Lsm protein sequences	Ģ	
1.1.6	Structures of Lsm protein ring complexes	13	
1.2	Scope of the thesis	21	
<u>2 Ma</u>	terials and Methods	27	
2.1	Materials	27	
2.1.1	Reagents	27	
2.1.2	Growth media and buffers	27	
2.1.3	Bacterial strains plasmids and RNA	30	
2.2	Methods	33	
2.2.1	Preservation of bacterial strains	33	
2.2.2	Cloning of Lsm polyproteins	33	
2.2.3	Plasmid isolation from E. coli	34	
2.2.4	Preparation of chemically-competent cells	34	
2.2.5	Transformation for plasmid propagation	35	
2.2.6	Transformation for protein expression	36	
2.2.7	Expression of Lsm polyproteins	36	
2.2.7.	1 Small-scale expression trials	36	
2.2.7.3	2 Large scale protein expression	37	

2.2.7.3 Protein expression for L-selenomethionine incorporation	38
2.2.8 Protein extraction	38
2.2.9 Purification of recombinant Lsm polyproteins	39
2.2.9.1 Chromatography equipment, media and columns	39
2.2.9.2 IMAC purification of Lsm polyproteins	39
2.2.9.3 Preparative size exclusion chromatography (SEC)	40
2.2.10 Protein concentration and storage	40
.2.11 Protein analysis	42
.2.11.1 Protein electrophoresis	42
.2.11.2 Analytical size exclusion chromatography	43
.2.11.3 SEC coupled to multi angle laser light scattering	44
SEC-MALLS)	44
.2.11.4 Small angle X-ray scattering (SAXS)	47
.3 Lsm-RNA interaction studies	49
.3.1 Surface plasmon resonance (SPR)	49
.3.2 Isothermal titration calorimetry (ITC)	50
.4 Crystallographic studies of Lsm polyproteins	51
.4.1 Crystallisation of Lsm polyproteins	51
4.2 Crystallographic data collection	52
.4.3 Crystallographic data processing	54
.4.4 Phasing of diffraction data	56
Solution behaviour of Lsm polyproteins	<u>58</u>
.1 Preparation of recombinant Lsm polyprotein complexes	61
.1 Quaternary structures of Lsm polyproteins in solution	65
.2.1 Polyprotein Lsm[4+1]	65
2.2 Polyprotein Lsm[4+1ext]	72
.2.3 Polyprotein Lsm[2+3]	74
.3 Conclusion	79
Biophysical characterisation of Lsm complexes and their RNA int	teractions 81
.1 Small Angle X-ray Scattering (SAXS)	83
1.1.1 Theory of Small Angle X-ray Scattering	83
.1.2 Scattering by Lsm polyprotein complexes	87
.1.2 Scattering by Esin polyprotein complexes .1.3 Solution shape of Lsm[2+3] ₄ with RNA	96
11.5 Solution shape of LSin($2+3$) ₄ with KNA	90

4.2	Surface plasmon resonance studies using immoblised RNA	101
4.2.1	Theory of surface plasmon resonance	101
4.2.2	SPR of Lsm polyproteins	105
4.3	Probing Lsm-RNA interactions by isothermal titration calorimetry (ITC)	113
4.3.1	Isothermal titration calorimetry theory	113
4.3.2	ITC experiments with Lsm polyproteins and RNA	114
4.4	Conclusion	120
<u>5 Cr</u>	ystallographic studies of Lsm polyproteins	125
5.1	Crystallisation screening	126
5.2	More extensive crystallisation strategies	131
5.2.1	Co-crystallisation with RNA	131
5.2.2	In situ proteolysis	131
5.2.3	Systematic screens	134
5.2.4	Grid screens	136
5.2.5	Searches of additives to assist crystal quality	141
5.2.6	Microseeding	144
5.2.7	Heavy atom derivatives of Lsm[4+1] ₄	146
5.2.7	.1 SeMet-Lsm[4+1] ₄	146
5.2.7	.2 Iodide derivatives of Lsm[4+1] ₄	149
5.3	Crystallographic studies	149
5.3.1	Diffraction from Lsm[4+1] ₄ crystals	149
5.3.2	Processing of acquired diffraction data	153
5.3.3	Molecular replacement procedures	158
<u>6 Co</u>	onclusion	167
6.1	Solution behaviour of Lsm polyproteins	168
6.2	Evidence for ring structures in solution	170
6.3	RNA binding by Lsm polyproteins	172
6.4	Crystallisation of Lsm polyproteins	174
6.5	Future perspectives and outlook	176
7 Re	ferences	179

Acknowledgements

A lot of hard work and effort went into the preparation of this thesis and many people guided and supported me along the way and deserve to be thanked. First of all, I thank my supervisor Dr. Bridget Mabbutt for giving me the opportunity to work on this exciting project and for her guidance and support throughout my thesis.

Thanks also goes to Dr. Robert Willows for advice on SPR experiments, Dr. Richard Hutton for ITC training, Dr. Grant Pearce who offered me beamtime for SAXS data collection, and Dr. Paul Curmi, Dr. Louise Brown and Dr. Paul Haynes for helpful discussions and inputs.

Special thanks to Dr. Stephen Harrop for sitting with me through countless hours of data collection at the Australian Synchrotron and for his advice on protein crystallisation experiments and crystallographic data processing.

All current and past members of the protein structure group deserve to be thanked, in particular Meghna for providing an excellent starting point for my own work, and for passing on her technical skills, Bhumika, Chandrika, Fran, Jan and Vani for their support, help, friendship, discussions and for keeping me company in the lab.

I am also grateful for the support I experienced from my family an friends and especially my fiancée Kerstin. Thanks for putting up with me and for making me laugh even in times of stress in the final stages.

Declaration

Where appropriate, work done in collaboration with other groups or individuals has been acknowledged. Outside these contributions, the material in this thesis is entirely my own work and to the best of my knowledge original. No part of this thesis has been submitted for a higher degree to any other university or institution. I consent to this thesis being made available for photocopy and loan.

Jens Moll

Department of Chemistry and Biomolecular Sciences

Macquarie University

Sydney NSW 2109

Australia

August 2010

Abstract

Lsm proteins are a family of RNA chaperones present in all kingdoms of life. Members of this protein family organise into ring-shaped quaternary structures and engage in the processing, sorting and regulation of a variety of RNA species. In archaea and bacteria, homomeric complexes of six or seven proteins are functional, whilst discrete heteromeric complexes of seven distinct Lsm proteins occur in eukaryotes. Eukaryotic Lsm assemblies modulate according to cellular localisation and RNA target, demonstrating that specific functionalities may exist for individual Lsm proteins.

In this study, I utilised Lsm polyproteins to pursue structural and functional studies of mixed Lsm rings, as well as to probe their quaternary dynamics. My work focused on two polyprotein forms, fusions of yeast Lsm[2+3] and Lsm[4+1]. Size exclusion chromatography in conjunction with static light scattering detects the formation of stable tetra- and octameric complexes, suggesting the formation of single and stacked tetrameric rings. Elevated populations of octamers are favoured at low ionic strength, indicating electrostatically-mediated packing of Lsm tetramers. A ring morphology for both tetrameric and octameric assemblies is confirmed by small angle X-ray scattering, estimating toroid dimensions to be 75 Å x 50 Å.

The simplified Lsm polyprotein complexes provide excellent probes of specific Lsm affinities for RNA sequences. Differential affinities of Lsm polyprotein towards Urich G_5U_{10} and U_{10} oligonucleotides are detected by surface plasmon resonance and isothermal titration calorimetry. The highest affinity for these oligonucleotides are

observed for Lsm[2+3] (K_D = 34 ± 15 nM), possibly due to specific basic residues within the linker used to fuse Lsm2 and Lsm3 domains. Isolated Lsm polyprotein complexes were subjected to crystallographic studies, resulting in regular crystalline forms of Lsm[4+1] in three distinct morphologies. Subsequently, native datasets were collected and processed to 3 Å resolution. Extensive phasing attempts using molecular replacement have been made, but have not so far yielded a solution. This data will serve to solve the first crystal structure of a heteromeric Lsm protein complex at atomic resolution upon collection of a suitable heavy atom dataset, however, despite extensive screening, only weakly diffracting crystals were obtained from L-selenomethionine derivatives of the protein to date.

The results obtained from simplified Lsm complexes aid the understanding of natural Lsm assemblies *in vivo*. It is the dynamic reorganisation of Lsm complexes that likely contributes to the hurdle of obtaining quality diffracting crystals for Lsm complexes. My biophysical studies have, however, confirmed the likelihood of ringshaped morphology of mixed Lsm rings *in vivo*, as well as differential affinities for RNA by their separate Lsm components.

List of abbreviations

List is based on the abbreviations accepted by JBC

Å angstrom

A₂₈₀ absorbance at 280 nm

A₂₆₀ absorbance at 260 nm

AfSm1 Sm protein from Archeoglubus fulgidus

ASU asymmetric unit

Bicine 2-(bis(2-hydroxyethyl)amino)acetic acid

dn/dc refractive index increment

D_{max} maximal particle diameter

Hfq bacterial Lsm protein

I3C 5-amino-2,4,6-triiodoisophthalic acid

k_a association rate constant

K_A equilibrium association constant

k_{av} size exclusion distribution coefficient

k_d dissociation rate constant

K_D equilibrium dissociation constant

LB Luria-Bertani

LLG log likelihood gain

Lsm Sm-like

Lsm[2+3] Polyprotein consisting of fused Lsm2 and Lsm3

Lsm[4+1] Truncated polyprotein consisting of fused Lsm4 and Lsm1

Lsm[4+1*ext*] Polyprotein consisting of fused truncated Lsm4 and full length

Lsm1

m₇G cap 5' methyl guanosine cap

MALLS multi angle laser light scattering

Mt Lsmα *M. thermoautotrophicum* Lsmα

NSD normalized spatial discrepancies

OB-fold oligosaccharide/oligonucleotide binding fold

OD₆₀₀ optical density at 600 nm

PaSm1 Sm protein from Pyrobaculum aerophilum

P-body processing body

PDB Protein Data Bank

PEG polyethyleneglycole

pka acid dissociation constant

p(r) electron distance distribution function

R_{eq} surface Plasmon resonance signal at equilibrium

R_g radius of gyration

RNase P ribonuclease P

RNP ribonucleo-protein

rRNA ribosomal RNA

SAD single wavelength anomalous dispersion

SAXS small angle X-ray scattering

SEC size exclusion chromatography

SeMet L-selenomethionine

SIRAS single isomorphous replacement plus anomalous scattering

SMN Survival of Motor Neurons protein

snoRNA small nucleolar RNA

snRNP small nuclear ribonucleoprotein

sRNA small RNA

Tacsimate mixture of titrated organic acid salts

TCEP tris(2-carboxyethyl)phosphine

TRAP Trp RNA-binding attenuation protein

U units

V₀ void volume of size exclusion column